

Antibodies and Kits
for the Study of

Chromatin and Epigenetic Regulation



Cell Signaling

TECHNOLOGY®

XP™ Monoclonal Antibodies for Chromatin and Epigenetic Regulation

XP™ monoclonal antibodies are a line of high quality rabbit monoclonal antibodies exclusively available from Cell Signaling Technology (CST). Any product labeled with XP has been carefully selected based on superior performance in all approved applications.

XP monoclonal antibodies are generated using XMT™, a proprietary monoclonal technology developed at CST. The technology provides access to a broad range of antibody-producing B cells unattainable with traditional monoclonal technologies, allowing more comprehensive screening and the identification of XP monoclonal antibodies with:

eXceptional specificity

As with all CST™ antibodies, the antibody is specific to your target of interest, saving you valuable time and resources.

+eXceptional sensitivity

The antibody will provide a stronger signal for your target protein in cells and tissues, allowing you to monitor expression of low levels of endogenous proteins, saving you valuable materials.

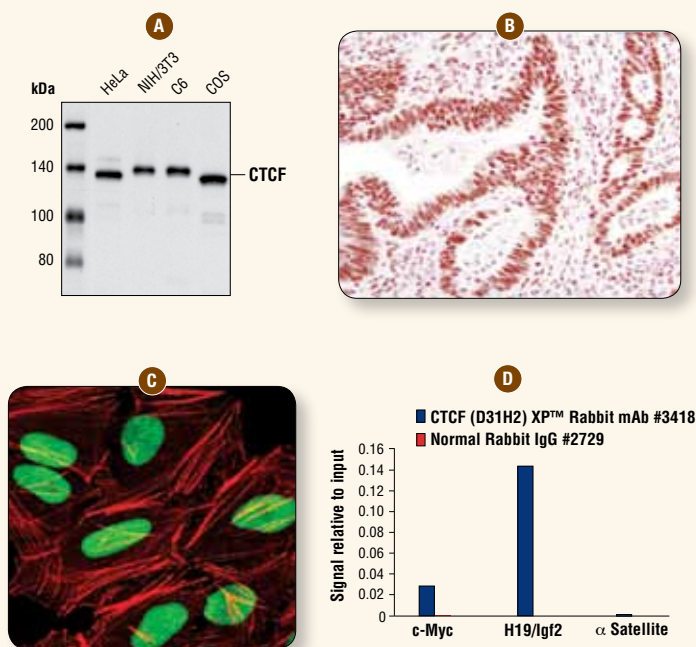
+eXceptional stability and reproducibility

XMT combined with our stringent quality control ensures maximum lot-to-lot consistency and the most reproducible results.

=eXceptional Performance™

XMT coupled with our extensive antibody validation and stringent quality control delivers XP monoclonal antibodies with eXceptional Performance in the widest range of applications.

CTCF (D31H2) XP™ Rabbit mAb #3418 is an example of an antibody with superior performance in a wide range of tested applications.



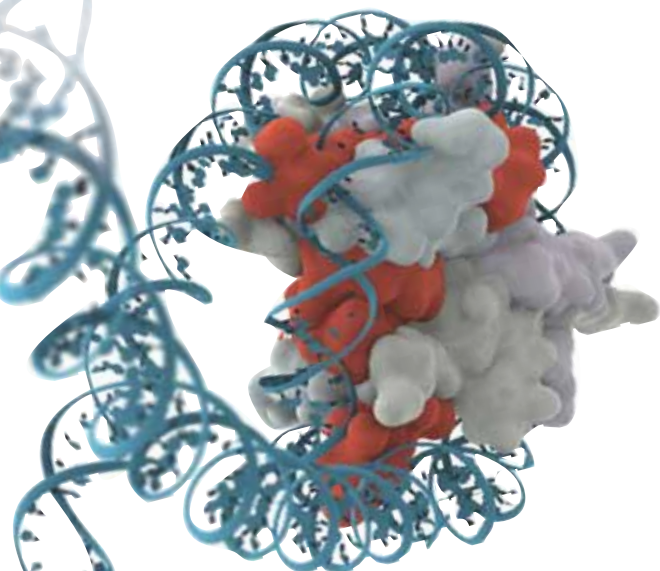
CTCF (D31H2) XP™ Rabbit mAb #3418: (A) WB analysis of extracts from various cell lines using #3418. (B) IHC analysis of paraffin-embedded human colon carcinoma using #3418. (C) Confocal IF analysis of HCT-116 cells using #3418 (green). Actin filaments were labeled with DY-554 phalloidin (red). (D) ChIP assays were performed with cross-linked chromatin from 4×10^6 HeLa cells, with either 10 μ l of #3418 or 2 μ l of Normal Rabbit IgG #2729, using SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using human c-Myc promoter primers, SimpleChIP® Human H19/lgt2 ICR Primers #5172, and SimpleChIP® Human α Satellite Repeat Primers #4486. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).

Visit our website for more experimental details, additional information, and a complete list of available XP Monoclonal Antibodies.

Antibodies and Kits for the Study of

Chromatin and Epigenetic Regulation

Cell Signaling Technology provides the highest quality activation-state and total protein antibodies for the study of chromatin and epigenetic regulation. CST™ antibodies have been extensively validated by our in-house scientists in applications including chromatin immunoprecipitation (ChIP), immunofluorescence, immunohistochemistry, and flow cytometry. Moreover, technical support is provided by the same scientists who develop and produce the antibodies and know them best. CST phosphorylation-specific antibodies are the most highly cited and serve as core reagents in multiple drug discovery platforms. Comprehensive and up-to-date information can be found at www.cellsignal.com.



The nucleosome, comprised of a histone octamer around which ~147 base-pairs of DNA are wrapped, is the basic unit of chromatin in the mammalian nucleus. Histone H3 is shown in red.

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SimpleChIP® Enzymatic Chromatin IP Kits

SimpleChIP® Enzymatic Chromatin IP Kits from Cell Signaling Technology (CST) are co-developed by CST and New England Biolabs and contain the highest quality research reagents. These kits are available with either Protein G agarose or Protein G magnetic beads and contain all buffers and reagents needed to perform up to 30 ChIP assays. The same reagents in these kits have been used for in-house ChIP validation at CST, which will

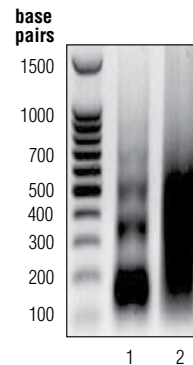
simplify your optimization. The kits can be used with any ChIP-validated antibody to detect endogenous levels of protein-DNA interactions and histone modifications in mammalian cells. Protein G agarose and Protein G magnetic beads are also available as separate products.

Please visit www.cellsignal.com/technologies/chip.html for information on ChIP and CST™ SimpleChIP products.

The SimpleChIP® Kit Advantage

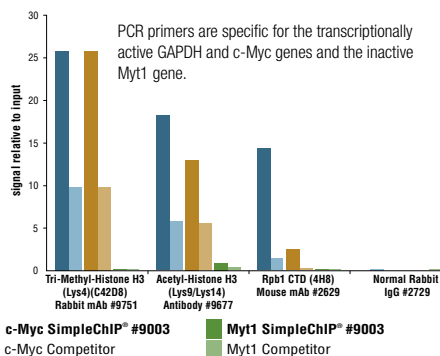
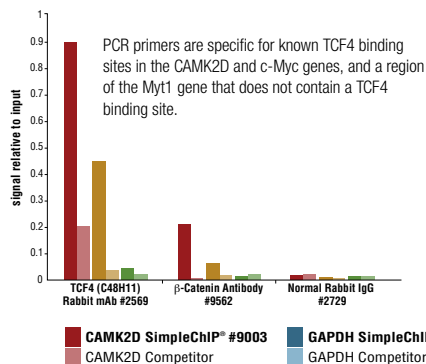
Enzyme-based Chromatin Digestion vs. Sonication-based Chromatin Fragmentation: Prior to performing chromatin IP, it is important to process chromatin to the appropriate DNA fragment size. CST SimpleChIP Enzymatic Chromatin IP Kits use micrococcal nuclease digestion to obtain chromatin fragments, whereas many competitor kits use a sonication-based method.

	SimpleChIP® from Cell Signaling Technology	Competitor Kits
Fragmentation Method	Enzymatic	Sonication
Chromatin Quality	High (mild sample treatment preserves chromatin integrity and antibody epitopes)	Low (rigorous sample treatment disrupts chromatin integrity and antibody epitopes)
IP Efficiency	High	Low
Detection	Higher sensitivity (especially crucial for transcription factors and cofactors)	Lower sensitivity (significantly decreased signal for less abundant chromatin binding proteins)



◀ **Enzyme-based and sonication-based ChIP kits produce chromatin fragments of a similar size, but differing chromatin integrity.** Chromatin was prepared from 4×10^7 HCT 116 human colorectal carcinoma cells according to the protocols included with the SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003 and from a competitor's sonication-based ChIP kit. DNA was purified from each chromatin sample and DNA fragment size was determined by electrophoresis on a 1% agarose gel. Both enzymatic digestion with the SimpleChIP® Kit (lane 1) and sonication with the competitor's kit (lane 2) produced chromatin fragments ranging from 150 to 700 bp, corresponding to one to five nucleosomes in length.

Enzyme-based Chromatin Digestion vs. Sonication-based Chromatin Fragmentation



◀ **SimpleChIP® digested chromatin is more conducive to immunoprecipitation than sonicated chromatin.** ChIP assays were performed with 10 µg of cross-linked HCT 116 chromatin and the indicated antibodies, using SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003 and a competitor's sonication-based ChIP Kit. The enriched DNA was quantified by qPCR. The amount of immunoprecipitated DNA in each sample is presented as a percent of the total input chromatin. For every target tested, enzyme-digested chromatin showed better enrichment of target DNA loci than did sonicated chromatin.

SimpleChIP® Related Products

	Reactivity
#9002 SimpleChIP® Enzymatic Chromatin IP Kit (Agarose Beads)	H, M, R, Mk
#9003 SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads)	H, M, R, Mk
#7017 6-Tube Magnetic Separation Rack	
#9007 ChIP-Grade Protein G Agarose Beads	
#9006 ChIP-Grade Protein G Magnetic Beads	
#2729 Normal Rabbit IgG	
#5415 Mouse (G3A1) mAb IgG1 Isotype Control	

SimpleChIP® Control PCR Primers

SimpleChIP Control PCR Primers featured throughout this brochure are a mix of forward and reverse primers that can be used to amplify DNA isolated using ChIP. These primers amplify positive control DNA sequences that contain known binding sites of the target protein detected by the antibody employed in the ChIP assay, and can also be used as a negative control to demonstrate antibody sensitivity.

- Primers are designed, tested, and optimized in-house in conjunction with our ChIP-validated antibodies and SimpleChIP Kits, saving time and reagents.
- Primers are optimized for use in real-time PCR with SYBR® Green dye, which simplifies quantification of DNA enrichment.
- Technical Support is provided by the scientists who designed and use these products, and know them best, assuring a fast and accurate response.

REACTIVITY KEY:

H human / M mouse / R rat / Hm hamster / Mk monkey / C chicken / Mi mink / Dm D. melanogaster / X Xenopus / Z zebra fish / B bovine / Dg dog / Pg pig / Sc S. cerevisiae / All all species expected / () 100% sequence homology

SimpleChIP® Enzymatic Chromatin IP Kits

30 assays

SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003:

#9002 SimpleChIP® Enzymatic Chromatin IP Kit (Agarose Beads)

Glycine Solution (10X)	ChIP-Grade Protein G Agarose Beads (blocked with BSA and sonicated salmon sperm DNA) #9007
Buffer A (4X)	DNA Purification Columns
Buffer B (4X)	Protease Inhibitor Cocktail (200X)
ChIP Buffer (10X)	Proteinase K
ChIP Elution Buffer (2X)	SimpleChIP® Human RPL30 Exon 3 Primers #7014
5 M NaCl	SimpleChIP® Mouse RPL30 Intron 2 Primers #7015
0.5 M EDTA	Histone H3 (D2B12) XP™ Rabbit mAb (ChIP Formulated) #4620
DNA Binding Buffer	Normal Rabbit IgG #2729
DNA Wash Buffer	1M DTT
DNA Elution Buffer	
RNAse A (10 mg/ml)	
Micrococcal Nuclease	

#9003 SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads)

This kit contains the same components as #9002 except #9003 contains ChIP-Grade Protein G Magnetic Beads #9006 (blocked with BSA).

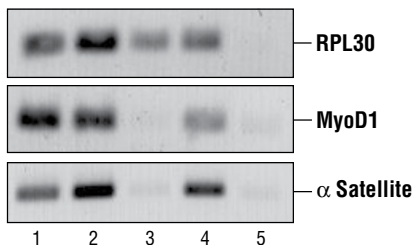
Selected Application References:

SimpleChIP® Enzymatic Chromatin IP Kit (Agarose Beads) #9002:

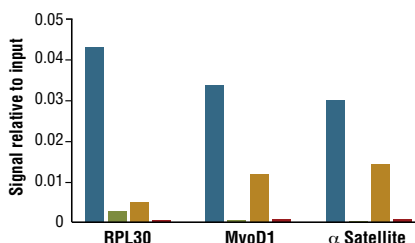
Czymai, T. et al. (2010) *J. Biol. Chem.* 285, 10163-10178. (ChIP)
 Ghosh, R. et al. (2010) *PLoS One* 5, e9575. (ChIP)

SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003:

Bommer, G.T. et al. (2010) *J. Biol. Chem.* 285, 1928-1938. (ChIP)
 De Bruyne, E. et al. (2010) *Blood* 115, 2430-2440. (ChIP)
 Onishi, Y. (2010) *Biosci. Rep.* 31, 57-62. (ChIP)



ChIP assays were performed using digested chromatin from HeLa cells and either Histone H3 (D2B12) XP™ Rabbit mAb (ChIP Formulated) #4620 (lane 2), Rpb1 CTD (4H8) Mouse mAb #2629 (lane 3), Di-Methyl Histone H3 (Lys9) Antibody #9753 (lane 4), or Normal Rabbit IgG #2729 (lane 5). Purified DNA was analyzed by standard PCR methods using SimpleChIP® Human RPL30 Exon 3 Primers #7014, SimpleChIP® Human MyoD1 Exon 1 Primers #4490, and SimpleChIP® Human alpha-Satellite Repeat Primers #4486. PCR products were observed for each primer set in the input sample (lane 1) and various ChIP samples, but not in the Normal Rabbit IgG ChIP sample (lane 5).



ChIP assays were performed using digested chromatin from HeLa cells and the indicated ChIP-validated antibodies. Purified DNA was analyzed by quantitative real-time PCR using SimpleChIP® Human RPL30 Exon 3 Primers #7014 (control primer set), SimpleChIP® Human MyoD1 Exon 1 Primers #4490, and SimpleChIP® Human alpha-Satellite Repeat Primers #4486. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).

■ Histone H3 (D2B12) XP™ Rabbit mAb (ChIP Formulated) #4620
 ■ Di-Methyl-Histone H3 (Lys9) Antibody #9753
 ■ Rpb1 CTD (4H8) Mouse mAb #2629
 ■ Normal Rabbit IgG #2729

SimpleChIP® Assay Kits

New SimpleChIP Assay Kits combine several ChIP-validated antibodies with control PCR primer mixes that serve as markers for pluripotency or epigenetic status. Rigorous in-house quality control and testing ensure the antibodies included in the kits meet the highest standards for quality, validation, and lot-to-lot consistency. The kits provide all

reagents necessary to perform 10 ChIP assays and subsequent real-time PCR reactions. Pre-selected positive and negative primer sets are included in each kit, providing proven and appropriate controls for customer experiments.

10 assays

#8980 SimpleChIP® Stem Cell Master Regulator Assay Kit

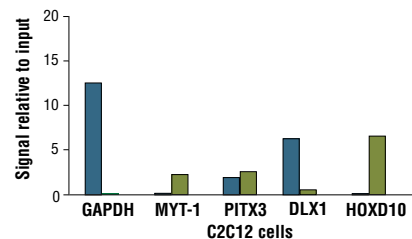
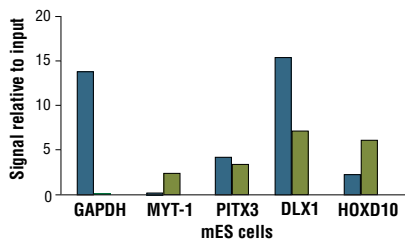
Nanog (D73G4) XP™ Rabbit mAb (ChIP Formulated) #5232
 Oct-4A (C30A3C1) Rabbit mAb (ChIP Formulated) #5677
 Sox2 (D6D9) XP™ Rabbit mAb (ChIP Formulated) #5024
 SimpleChIP® Human Oct-4 Promoter Primers #4641
 SimpleChIP® Human alpha-Satellite Repeat Primers #4486

#8982 SimpleChIP® Human Bivalent Promoter Assay Kit

Tri-Methyl-Histone H3 (Lys4) (C42D8) Rabbit mAb #9751
 Tri-Methyl-Histone H3 (Lys27) (C36B11) Rabbit mAb #9733
 SimpleChIP® Human GAPDH Exon 1 Primers #5516
 SimpleChIP® Human MYT-1 Exon 1 Primers #4493
 SimpleChIP® Human GATA6 Promoter Primers #5550

#8981 SimpleChIP® Mouse Bivalent Promoter Assay Kit

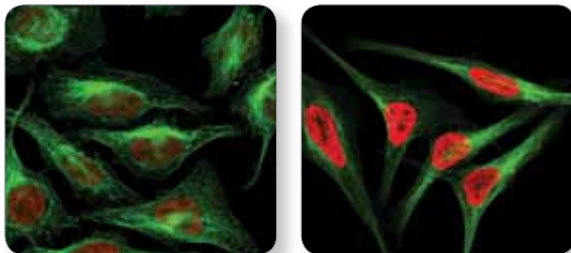
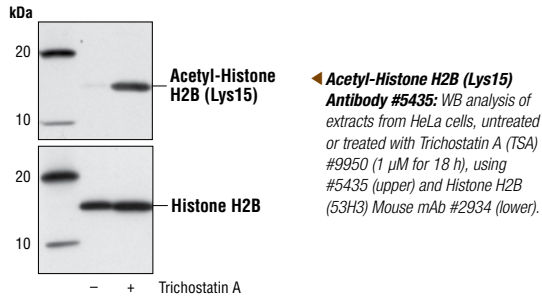
Tri-Methyl-Histone H3 (Lys4) (C42D8) Rabbit mAb #9751
 Tri-Methyl-Histone H3 (Lys27) (C36B11) Rabbit mAb #9733
 SimpleChIP® Mouse GAPDH Intron 2 Primers #8986
 SimpleChIP® Mouse MYT-1 Promoter Primers #8985
 SimpleChIP® Mouse PITX3 Intron 1 Primers #8984



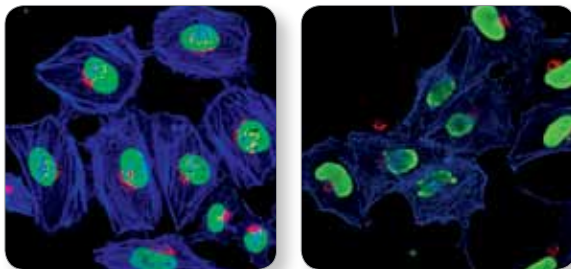
■ Tri-Methyl-Histone H3 (Lys4) (C42D8) Rabbit mAb #9751
 ■ Tri-Methyl-Histone H3 (Lys27) (C36B11) Rabbit mAb #9733
 ■ Normal Rabbit IgG #2729

SimpleChIP® Mouse Bivalent Promoter Assay Kit #8981: ChIP assays were performed with cross-linked chromatin from 4×10^6 mouse embryonic stem (mES) cells (left panel) or C2C12 cells (right panel) and Tri-Methyl-Histone H3 (Lys4) (C42D8) Rabbit mAb #9751, Tri-Methyl-Histone H3 (Lys27) (C36B11) Rabbit mAb #9733, or 2 μ l of Normal Rabbit IgG #2729, using SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using SimpleChIP® Mouse GAPDH Intron 2 Primers #8986, SimpleChIP® Mouse MYT-1 Promoter Primers #8985, SimpleChIP® Mouse PITX3 Intron 1 Primers #8984, mouse DLX1 promoter primers, and mouse HOXD10 intron 1 primers. The amount of immunoprecipitated DNA in each sample is normalized for enrichment of total histone H3 and represented as signal relative to the total amount of input chromatin (equivalent to one). Note that the PITX3, DLX1, and HOXD10 promoters are all bivalent in stem cells, while only PITX3 remains bivalent in the differentiated cell line C2C12.

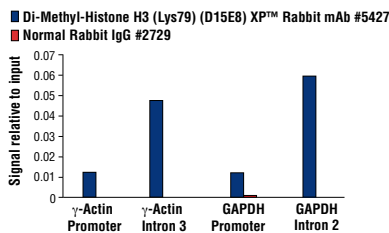
Histones and Histone Modifications



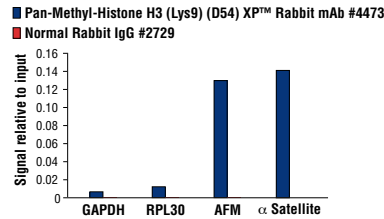
Acetyl-Histone H3 (Lys9) (C5B11) Rabbit mAb (Alexa Fluor[®] 555 Conjugate) #5489: Confocal IF analysis of HeLa cells, untreated (left) or treated with Trichostatin A (TSA) #9950 (right), using #5489 (red) and α -Tubulin (DM1A) Mouse mAb #3873 (green).



Acetyl-Histone H4 (Lys5) Antibody #9672: Confocal IF analysis of HeLa cells, untreated (left) or treated with Trichostatin A (TSA) #9950 (right), using #9672 (green) and Golgin-97 Antibody (red). Actin filaments were labeled with a dye-conjugated phalloidin (blue pseudocolor).



Di-Methyl-Histone H3 (Lys9) (D15E8) XP™ Rabbit mAb #5427: ChIP assays were performed with cross-linked chromatin from 4×10^6 HeLa cells and either 10 μ l of #5427 or 2 μ l of Normal Rabbit IgG #2729 using SimpleChIP[®] Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using SimpleChIP[®] Human γ -Actin Promoter Primers #5037, SimpleChIP[®] Human γ -Actin Intron 3 Primers #5047, SimpleChIP[®] Human GAPDH Promoter Primers #4471, and SimpleChIP[®] Human GAPDH Intron 2 Primers #4478. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).



Pan-Methyl-Histone H3 (Lys9) (D54) XP™ Rabbit mAb #4473: ChIP assays were performed with cross-linked chromatin from 4×10^6 HeLa cells and either 20 μ l of #4473 or 2 μ l of Normal Rabbit IgG #2729, using SimpleChIP[®] Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using SimpleChIP[®] Human GAPDH Exon 1 Primers #5516, SimpleChIP[®] Human RPL30 Exon 3 Primers #7014, SimpleChIP[®] Human AFM Intron 1 Primers #5098, and SimpleChIP[®] Human α Satellite Repeat Primers #4486. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).

Acetylation

		Application	Reactivity
NEW	#9814 Acetylated-Lysine (Ac-K2-100) Rabbit mAb	W, IP, ChIP, E-P	All
	#9681 Acetylated-Lysine Mouse mAb (Ac-K-103)	W, E-P	All
	#9441 Acetylated-Lysine Antibody	W, IP, IHC-P, IF-IC, ChIP, E-P	All
H2A	#2576 Acetyl-Histone H2A (Lys5) Antibody	W, IP, IHC-P	H, M, R, Mk
H2B	#2574 Acetyl-Histone H2B (Lys5) Antibody	W, IP, IHC-P	H, M, R, Mk
	NEW #5410 Acetyl-Histone H2B (Lys12) Antibody	W, IP, IF-IC	H, M, R, Mk
	NEW #5435 Acetyl-Histone H2B (Lys15) Antibody featured	W, IF-IC	H, M, R, Mk, (B, Pg)
	#2571 Acetyl-Histone H2B (Lys20) Antibody	W, IP, IHC-P, IF-IC, IC	H, M, R
H3	#9649 Acetyl-Histone H3 (Lys9) (C5B11) Rabbit mAb	W, IHC-P, IF-IC, F, ChIP	H, M, R, Mk, Z
	NEW #9683 Acetyl-Histone H3 (Lys9) (C5B11) Rabbit mAb (Alexa Fluor[®] 488 Conjugate)	IF-IC, F	H, M, Z, (R, Mk)
	NEW #5489 Acetyl-Histone H3 (Lys9) (C5B11) Rabbit mAb (Alexa Fluor[®] 555 Conjugate) featured	IF-IC	H, M, Z, (R, Mk)
	NEW #4484 Acetyl-Histone H3 (Lys9) (C5B11) Rabbit mAb (Alexa Fluor[®] 647 Conjugate)	IF-IC, F	H, M, Z, (R, Mk)
	#9711 Acetyl- and Phospho-Histone H3 (Lys9/Ser10) Antibody	W, IHC-P, IF-P	H, M, R
	#9671 Acetyl-Histone H3 (Lys9) Antibody	W, IP, IHC-P, ChIP	H, M, R, Mk, Dm, Sc
	#9677 Acetyl-Histone H3 (Lys9/Lys14) Antibody	W, IP, ChIP	H, M, R, Mk, (Z)
	NEW #4318 Acetyl-Histone H3 (Lys14) Antibody	W, IF-IC	H, M, R, Mk, (Drm)
	NEW #5275 Acetyl-Histone H3 (Lys14) Antibody (ChIP Formulated) featured	ChIP	H, (M, R, Mk)
	#9675 Acetyl-Histone H3 (Lys18) Antibody	W, IHC-P, ChIP	H, M, R
	#9674 Acetyl-Histone H3 (Lys23) Antibody	W, IHC-P	H, M, R
	NEW #4353 Acetyl-Histone H3 (Lys27) Antibody	W, IP, ChIP	H, M, R, Mk, (Hm, C, Dm, X, Z, B)
	NEW #4243 Acetyl-Histone H3 (Lys56) Antibody	W, IP, IF-IC	H, M, R, Mk
H4	NEW #9672 Acetyl-Histone H4 (Lys5) Antibody featured	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk, (C, Dm, X, Z, B, Pg)
	#2594 Acetyl-Histone H4 (Lys8) Antibody	W, IF-IC, F, ChIP	H, M, R, Mk
	#2591 Acetyl-Histone H4 (Lys12) Antibody	W, IHC-P, IF-IC, F	H, M, R, Mk

Selected Application References:

Acetylated-Lysine Mouse mAb (Ac-K-103) #9681:

VanDemark, A.P. et al. (2007) *Mol. Cell* 27, 817-828. (W)

Zhao, L.J. et al. (2006) *J. Biol. Chem.* 281, 36613-36623. (W)

Acetylated-Lysine Antibody #9441:

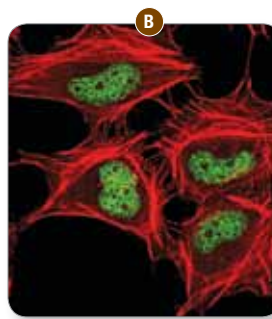
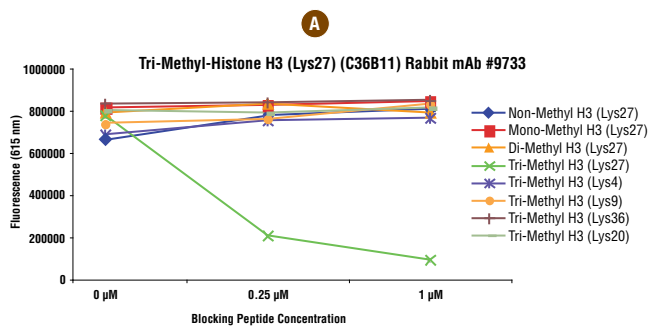
Werner, H.B. et al. (2007) *J. Neurosci.* 27, 7717-7730. (W)

Acetyl-Histone H3 (Lys9) (C5B11) Rabbit mAb #9649:

Kong, D.K. et al. (2010) *Mol. Biol. Cell* 21, 1335-1349. (ChIP)

APPLICATIONS KEY:

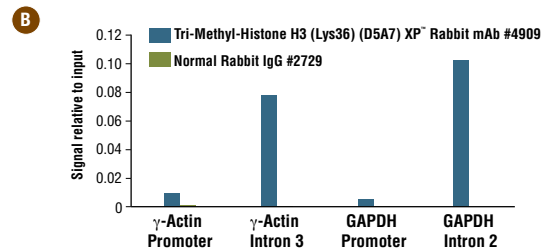
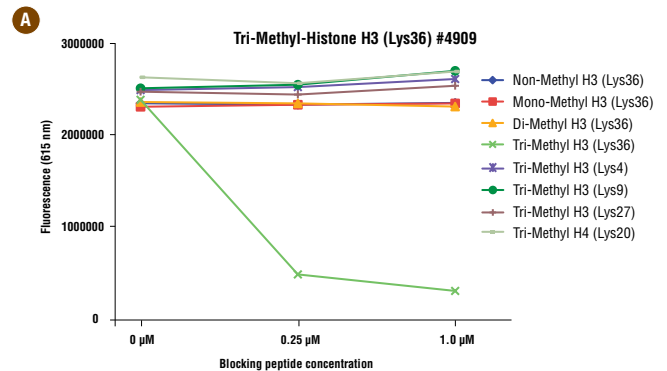
W Western / IP Immunoprecipitation / IHC Immunohistochemistry / IF Immunofluorescence / F Flow Cytometry / ChIP Chromatin Immunoprecipitation / (-IC Immunocytochemistry, -P Paraffin, -F Frozen) / E-P Peptide ELISA



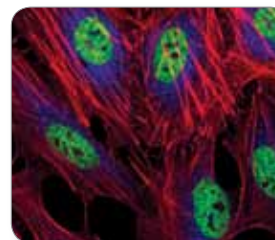
Tri-Methyl-Histone H3 (Lys27) (C36B11) Rabbit mAb #9733:
(A) Specificity of the antibody was determined by peptide ELISA. The graph depicts the binding of the antibody to pre-coated tri-methyl histone H3 (Lys27) peptide in the presence of increasing concentrations of various competitor peptides. As shown, only the tri-methyl histone H3 (Lys27) peptide (green) competed away binding of the antibody. **(B)** Confocal IF analysis of HeLa cells using #9733 (green). Actin filaments were labeled with DY-554 phalloidin (red).

Methylation

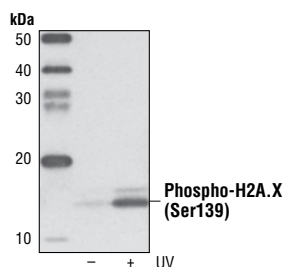
		Application	Reactivity
H3	#9707 Methyl-Histone H3 (Arg2) Antibody	W	H
NEW	#5326 Mono-Methyl-Histone H3 (Lys4) (D1A9) XP™ Rabbit mAb <i>featured</i>	W, IF-IC, ChIP	H, M, R, Mk
	#9723 Mono-Methyl-Histone H3 (Lys4) Antibody	W, IP, IF-IC	H, M, R, Mk, (X, Z)
	#9725 Di-Methyl-Histone H3 (Lys4) (C64G9) Rabbit mAb	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk
	#9726 Di-Methyl-Histone H3 (Lys4) Antibody	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk, (X, Z)
	#9751 Tri-Methyl-Histone H3 (Lys4) (C42D8) Rabbit mAb	W, IHC-P, IF-IC, ChIP	H, M, R, Mk, Dm, Sc, (X, Z)
	#9727 Tri-Methyl-Histone H3 (Lys4) Antibody	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk, (X, Z)
NEW	#4658 Di-Methyl-Histone H3 (Lys9) (D85B4) XP™ Rabbit mAb	W, IP, IF-IC, ChIP	H, M, R, Mk, (Dm, X, Z, B, Pg, Sc)
	#9753 Di-Methyl-Histone H3 (Lys9) Antibody	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk, Dm, Sc
NEW	#5327 Di/Tri-Methyl-Histone H3 (Lys9) (6F12) Mouse mAb	W, IP, IF-IC, ChIP	H, M, R, Mk
	#9754 Tri-Methyl-Histone H3 (Lys9) Antibody	W, IF-IC, ChIP	H, M, R, Mk, (Dm, Pg)
NEW	#4473 Pan-Methyl-Histone H3 (Lys9) (D54) XP™ Rabbit mAb <i>featured</i>	W, IP, IF-IC, ChIP	H, M, R, Mk, (C, Dm, X, Z, B, Pg, Sc)
	#4069 Pan-Methyl-Histone H3 (Lys9) Antibody	W, IP, IF-IC, ChIP	H, M, R, Mk, Z
	#9728 Di-Methyl-Histone H3 (Lys27) (D18C8) XP™ Rabbit mAb	W, IF-IC, ChIP	H, M, R, Mk
	#9755 Di-Methyl-Histone H3 (Lys27) Antibody	W, IP, IF-IC	H, M, R, Mk
	#9733 Tri-Methyl-Histone H3 (Lys27) (C36B11) Rabbit mAb <i>featured</i>	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk, (X, Z)
	#9756 Tri-Methyl-Histone H3 (Lys27) Antibody	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk, (X)
	#2901 Di-Methyl-Histone H3 (Lys36) (C75H12) Rabbit mAb	W, IHC-P, IF-IC	H, M, R, Mk
	#9758 Di-Methyl-Histone H3 (Lys36) Antibody	W, IP, IF-IC	H, M, R, Mk
NEW	#4909 Tri-Methyl-Histone H3 (Lys36) (D5A7) XP™ Rabbit mAb <i>featured</i>	W, ChIP	H, M, R, Mk, (Hm, C, Dm, X, Z, B)
	#9763 Tri-Methyl-Histone H3 (Lys36) Antibody	W, IHC-P, IF-IC	H, M, R, Mk
NEW	#5427 Di-Methyl-Histone H3 (Lys79) (D15E8) XP™ Rabbit mAb <i>featured</i>	W, ChIP	H, M, R, Mk
	#9757 Di-Methyl-Histone H3 (Lys79) Antibody	W, IP	H, M, R, Mk
H4	#9724 Mono-Methyl-Histone H4 (Lys20) Antibody	W	H, M, R, Mk, (Dm, X, Z, B, Pg)
	#9759 Di-Methyl-Histone H4 (Lys20) Antibody	W	H, M, R, Mk, (Dm, Z, B, Pg.)
NEW	#5737 Tri-Methyl-Histone H4 (Lys20) Rabbit mAb	W, IP, ChIP	H, M, R, Mk



Tri-Methyl-Histone H3 (Lys36) (D5A7) XP™ Rabbit mAb #4909: Specificity of the antibody was determined by peptide ELISA. The graph **(A)** depicts the binding of the antibody to pre-coated tri-methyl histone H3 (Lys36) peptide in the presence of increasing concentrations of various competitor peptides. As shown, only the tri-methyl histone H3 (Lys36) peptide (green) competed away binding of the antibody. **(B)** ChIP assays were performed with cross-linked chromatin from 4×10^6 HeLa cells and either 10 μ l of #4909 or 2 μ l of Normal Rabbit IgG #2729 using SimpleChIP™ Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using SimpleChIP™ Human γ -Actin Promoter Primers #5037, SimpleChIP™ Human γ -Actin Intron 3 Primers #5047, SimpleChIP™ Human GAPDH Promoter Primers #4471, and SimpleChIP™ Human GAPDH Intron 2 Primers #4478. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).

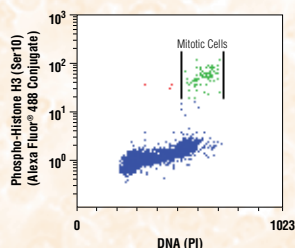
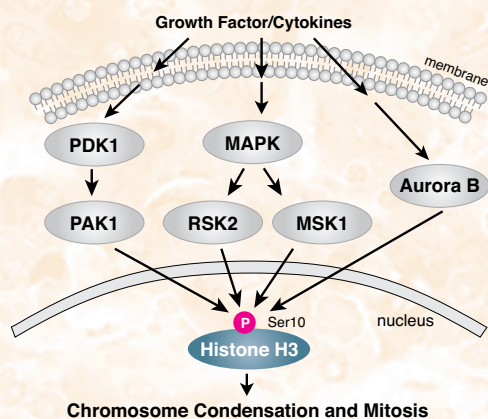


Mono-Methyl-Histone H3 (Lys4) (D1A9) XP™ Rabbit mAb #5326: Confocal IF analysis of HeLa cells using #5326 (green) and MEK1/2 (L38C12) Mouse mAb #4694 (blue). Actin filaments were labeled with DY-554 phalloidin (red).

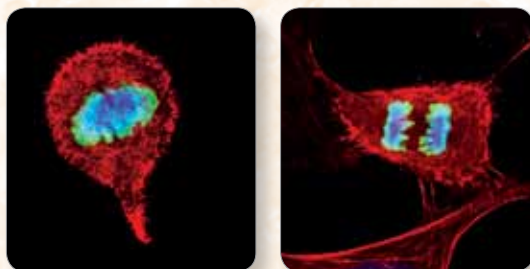


◀ **Phospho-Histone H2A.X (Ser139) (20E3) Rabbit mAb (Biotinylated) #4309:** WB analysis of extracts from HT-29 cells, untreated or UV-treated, using #4309 and detected with Streptavidin-HRP #3999.

Histone H3 Phosphorylation in Mitosis



◀ **Phospho-Histone H3 (Ser10) (D2C8) XP™ Rabbit mAb (Alexa Fluor® 488 Conjugate) #3465:** Flow cytometric analysis of Jurkat cells using #3465 compared to propidium iodide (DNA content). The box indicates phospho-histone H3 positive cells.



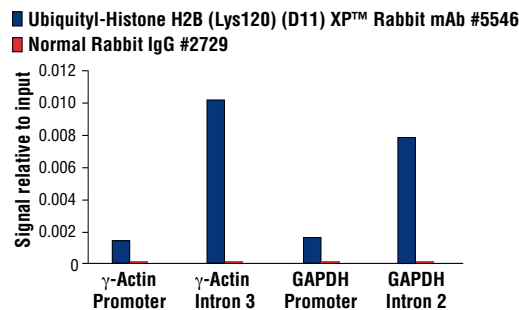
Phospho-Histone H3 (Ser10) (D2C8) XP™ Rabbit mAb #3377: Confocal IF analysis of HeLa cells using #3377 (green). Actin filaments were labeled with DY-554 phalloidin (red). Blue pseudocolor = DRAQ5® #4084 (fluorescent DNA dye).

Phosphorylation

		Application	Reactivity
H2A.X	#9718	Phospho-Histone H2A.X (Ser139) (20E3) Rabbit mAb	W, IHC-P, IF-IC, F H, M, R, Mk
	#9719	Phospho-Histone H2A.X (Ser139) (20E3) Rabbit mAb (Alexa Fluor® 488 Conjugate)	IF-IC, F H, M
	#9720	Phospho-Histone H2A.X (Ser139) (20E3) Rabbit mAb (Alexa Fluor® 647 Conjugate)	IF-IC, F H, M
	NEW #4309	Phospho-Histone H2A.X (Ser139) (20E3) Rabbit mAb (Biotinylated) <i>featured</i>	W H, M, R, Mk
	#2577	Phospho-Histone H2A.X (Ser139) Antibody	W, IHC-P, IF-IC, F H, M, R
	NEW #5438	Phospho-Histone H2A.X (Ser139/Tyr142) Antibody	W, IP, IF, F H, M, R, Mk
H3	#9714	Phospho-Histone H3 (Thr3) Antibody	W, IHC-P H, M, R
	#3377	Phospho-Histone H3 (Ser10) (D2C8) XP™ Rabbit mAb <i>featured</i>	W, IF-IC, F H, M, R, Mk, Z
	#3465	Phospho-Histone H3 (Ser10) (D2C8) XP™ Rabbit mAb (Alexa Fluor® 488 Conjugate) <i>featured</i>	IF-IC, F H, M, R, Mk
	#3475	Phospho-Histone H3 (Ser10) (D2C8) XP™ Rabbit mAb (Alexa Fluor® 555 Conjugate)	IF-IC H, M, R, Mk
	#3458	Phospho-Histone H3 (Ser10) (D2C8) XP™ Rabbit mAb (Alexa Fluor® 647 Conjugate)	IF-IC, F H, M, R, Mk
	#3642	Phospho-Histone H3 (Ser10) (D2C8) XP™ Rabbit mAb (Biotinylated)	W, IF-F, IF-IC, F H, M, R, Mk
	#9706	Phospho-Histone H3 (Ser10) (6G3) Mouse mAb	W, IF-F, IF-IC, F H, M, R
	#9701	Phospho-Histone H3 (Ser10) Antibody	W, IP, IHC-P, IHC-F, IF-IC, F H, M, R, Mk, C, Dm, Z, Sc, (X)
	#9708	Phospho-Histone H3 (Ser10) Antibody (Alexa Fluor® 488 Conjugate)	IF-IC, F H, M, (X)
	#9716	Phospho-Histone H3 (Ser10) Antibody (Alexa Fluor® 647 Conjugate)	IF-IC, F H, M, R, Mk
	#9767	Phospho-Histone H3 (Thr11) (C2A6) Rabbit mAb	W, IP, F H, M, R, (X)
#9764	Phospho-Histone H3 (Thr11) Antibody	W, IP, IF-IC, F H, M, R, (X)	
#9713	Phospho-Histone H3 (Ser28) Antibody	W, IP, IF-F, IF-IC, F, H, M, Hm, Dm, (R, C, X, Z, B)	
CENP-A	#2187	Phospho-CENP-A (Ser7) Antibody <i>featured</i>	W, IP, IF-IC H, (Mk)

Ubiquitylation

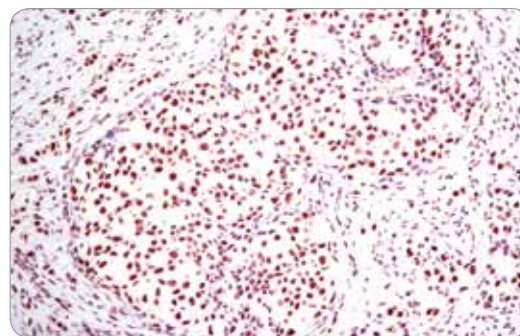
H2B	NEW #5546	Ubiquityl-Histone H2B (Lys120) (D11) XP™ Rabbit mAb <i>featured</i>	W, ChIP H, M, R, Mk
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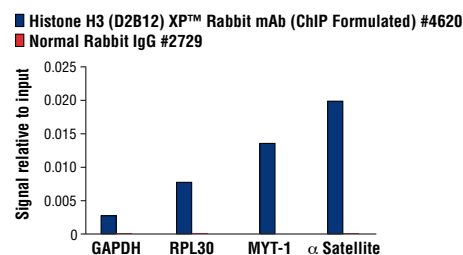
Ubiquityl-Histone H2B (Lys120) (D11) XP™ Rabbit mAb #5546: ChIP assays were performed with cross-linked chromatin from 4×10^6 HeLa cells and either 10 μ l of #5546 or 2 μ l of Normal Rabbit IgG #2729 using SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using SimpleChIP® Human γ -Actin Promoter Primers #5037, SimpleChIP® Human γ -Actin Intron 3 Primers #5047, SimpleChIP® Human GAPDH Promoter Primers #4471, and SimpleChIP® Human GAPDH Intron 2 Primers #4478. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).

Unmodified Histones and Variants

		Application	Reactivity
H2A	#3636 Histone H2A (L88A6) Mouse mAb	W, IHC-P	H, M, R, Mk, (C, Dm, Z, Pg)
	#2578 Histone H2A Antibody II	W, IP, IHC-P	H, M, R, Mk, (X)
	#2595 Histone H2A.X Antibody	W	H, Mk
	#2718 Histone H2A.Z Antibody	W, IP, IF-IC	H, M, R, Mk, Z, (C, X, B)
MACRO-H2A	NEW #4160 MacroH2A1.1 Antibody	W	H, M, R
	NEW #4827 MacroH2A1.2 Antibody <i>featured</i>	W, IF-IC	H, M, R, Mk, (C, B)
H2B	#2722 Histone H2B Antibody	W, IP	H, M, R, Mk
	#2934 Histone H2B (53H3) Mouse mAb	W	H, M, R, Mk, Z, (X, B)
H3	NEW #4499 Histone H3 (D1H2) XP™ Rabbit mAb <i>featured</i>	W, IHC-P, IF-IC	H, M, R, Mk, (Hm, C, Dm, X, Z, B)
	NEW #4620 Histone H3 (D2B12) XP™ Rabbit mAb (ChIP Formulated) <i>featured</i>	ChIP	H, M, (R, Hm, Mk, C, Dm, X, Z, B)
	#9717 Histone H3 (3H1) Rabbit mAb	W	H, M, R, Hm, Mk, Z, B, Pg
	NEW #5192 Histone H3 (3H1) Rabbit mAb (HRP Conjugate)	W	H, M, R, Hm, Mk, Z, B, Pg
	#3638 Histone H3 (96C10) Mouse mAb	W	H, M, R, Mk, (Dm, X)
	#3680 Histone H3 (96C10) Mouse mAb (IHC Formulated)	IHC-P	H, (M, R, Mk, Dm, X)
	#9715 Histone H3 Antibody	W, IHC-P, IF-IC	H, M, R, Mk, Z, B, Pg, (Dm)
	#2650 Histone H3 Antibody (ChIP Formulated)	ChIP	H, M, (R, Mk, C, Dm, X, Z, B)
H4	#2935 Histone H4 (L64C1) Mouse mAb	W, IHC-P	H, M, R, Mk, Z, (Dm, X, Z, B)
	#2960 Histone H4 (L64C1) Mouse mAb (ChIP Formulated)	ChIP	H, (M, R, Mk, Dm, X, Z, B)
	#2592 Histone H4 Antibody	W, IP, IHC-P	H, M, R, Mk, Dm, Z, Sc
CENP-A	#2048 CENP-A (C51A7) Rabbit mAb (Mouse Specific; IF Preferred)	W, IF-IC	M
	#2047 CENP-A (C5H3) Rabbit mAb (Mouse Specific)	W, IP	M
	#2186 CENP-A Antibody	W, IF-IC	H



Histone H3 (D1H2) XP™ Rabbit mAb #4499: IHC analysis of paraffin-embedded human breast carcinoma using #4499.



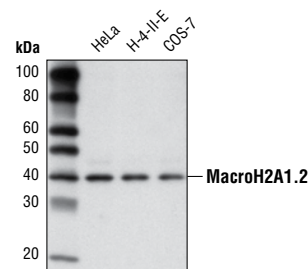
Histone H3 (D2B12) XP™ Rabbit mAb (ChIP Formulated) #4620: ChIP assays were performed with cross-linked chromatin from 4×10^6 HeLa cells and either 10 μ l of #4620 or 2 μ l of Normal Rabbit IgG #2729 using SimpleChIP® Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using SimpleChIP® Human GAPDH Exon 1 Primers #5516, SimpleChIP® Human RPL30 Exon 3 Primers #7014, SimpleChIP® Human MYT-1 Exon 1 Primers #4493, and SimpleChIP® Human α Satellite Repeat Primers #4486. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).



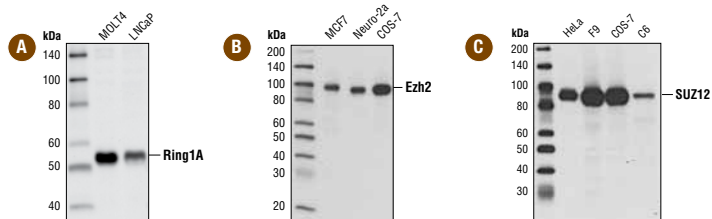
Phospho-CENP-A (Ser7) Antibody #2187: Confocal IF analysis of a mitotic HeLa cell using #2187 (green) and β -Tubulin (9F3) Rabbit mAb (Alexa Fluor® 555 Conjugate) #2116 (red). Phospho-CENP-A signal is localized to bright spots in the metaphase plate. Blue pseudocolor = DRAQ5® #4084 (fluorescent DNA dye).

Cell Signaling Technology offers the highest quality research products, rigorous validation, and technical support provided by the same scientists who produce and validate the products and know them best.

MacroH2A1.2 Antibody #4827: WB analysis of extracts from HeLa, H-4-II-E, and COS-7 cells using #4827.



Sampler Kits



Polycomb Group Antibody Sampler Kit #9788: WB analysis of extracts from various cell lines using (A) Ring1A Antibody #2820, (B) Ezh2 (D2C9) XP™ Rabbit mAb #5246, and (C) SUZ12 (D39F6) XP™ Rabbit mAb #3737.

- #9928 Histone Deacetylase (HDAC) Antibody Sampler Kit
- #9933 Acetyl-Histone Antibody Sampler Kit
- #9927 Acetyl-Histone H3 Antibody Sampler Kit
- #9847 Methyl-Histone H3 Antibody Sampler Kit
- #9849 Phospho-Histone H3 (Mitotic Marker) Antibody Sampler Kit
- NEW #9788 Polycomb Group Antibody Sampler Kit featured**

What does Antibody Validation Mean at Cell Signaling Technology?

Scientists at Cell Signaling Technology (CST) follow a stringent validation protocol using a combination of several approaches and applications to provide you with the highest quality antibodies. This ensures credible and reproducible results with the least expenditure of your costly time, samples, and reagents.

Antibody Validation at Cell Signaling Technology includes:

Testing in a Number of Applications to help you choose the antibody that works best in your experiment.

- ⌘ Western blot, Immunoprecipitation, Immunohistochemistry, Immunofluorescence, Flow cytometry, ChIP, Sandwich ELISA

Verifying Specificity and Reproducibility to ensure that the antibody performs consistently in all applications specified.

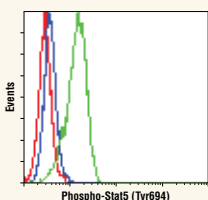
- ⌘ Treatment of cells with appropriate kinase-specific inhibitors to verify specificity
- ⌘ Analysis of a large panel of cell lines with known target expression levels to confirm target specificity
- ⌘ Phosphatase treatment to verify phospho-specificity
- ⌘ Comparison of antibody to isotype control antibody
- ⌘ Verification of target-specific signal in transfected cells, knock-out cells, or siRNA-treated cells
- ⌘ Blocking with antigen peptide
- ⌘ Verification of correct subcellular localization or treatment-induced translocation
- ⌘ Side-by-side comparison of a new lot with previous lots to ensure lot-to-lot consistency

Identifying Optimal Conditions to save your precious time, samples, and reagents.

- ⌘ Optimal dilutions and buffers predetermined
- ⌘ Positive and negative control cell extracts specified
- ⌘ Detailed protocols already optimized

Comparison of target-specific antibody to non-specific isotype control

Phospho-Stat5 (Tyr694) (C71E5) Rabbit mAb #9314: Flow cytometric analysis of K-562 cells, untreated (green) or gefitinib-treated (blue), using #9314 compared to concentration matched Rabbit (DA1E) mAb IgG XP™ Isotype Control #3900 (red).

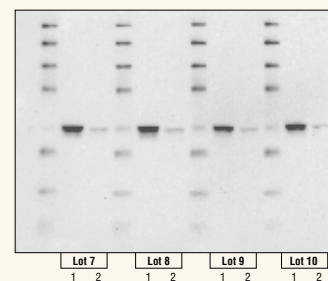


Side by side comparison of new lot with previous lots

Phospho-Akt (Ser473) Antibody #9271

Lot 7: 8/1/2002 Lot 9: 2/12/2004
Lot 8: 7/23/2003 Lot 10: 4/7/2006

- 1: C2C12 cells +insulin(100 nM for 10 min.)
- 2: C2C12 cells, untreated



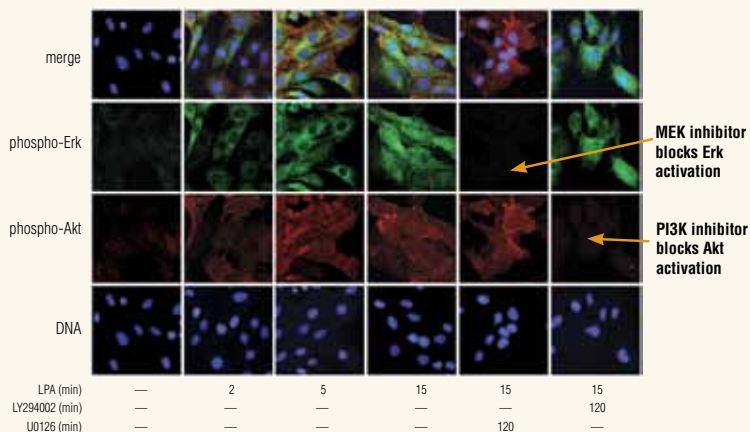
Verification of target-specificity using mouse models

Phospho-Akt (Ser473) (D9E) XP™ Rabbit mAb #4060:

IHC analysis of paraffin-embedded WT (left) and PTEN (-/-) (right) mouse prostate using #4060. Tissue courtesy of Dr. David Guertin, The Whitehead Institute for Biomedical Research, Cambridge, MA.



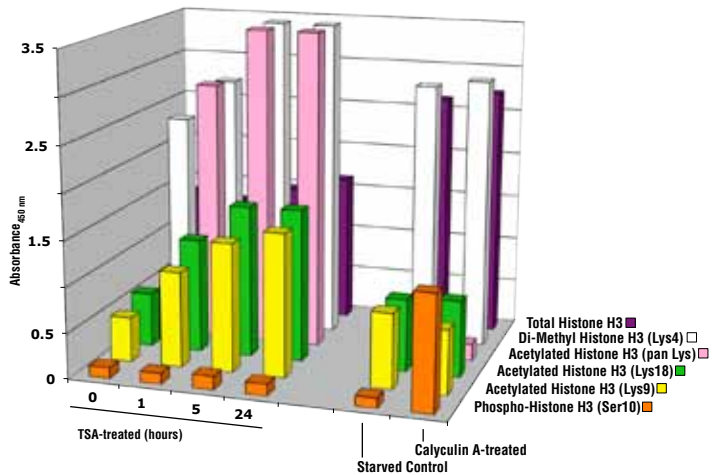
Verification of specificity using known target activators and inhibitors



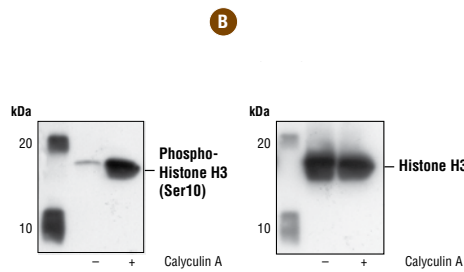
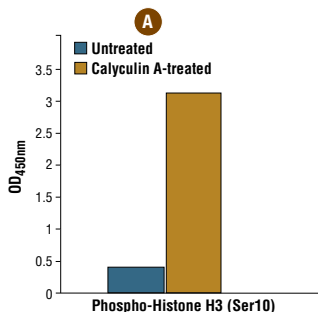
PathScan® Sandwich ELISA

	Reactivity
#7233 PathScan® Acetyl-Histone H2A Sandwich ELISA Kit	H, M, Mk
#7218 PathScan® Acetyl-Histone H2B (Lys5) Sandwich ELISA Kit	H, M, Mk
#7222 PathScan® Acetyl-Histone H2B (Lys20) Sandwich ELISA Kit	H, M, Mk
#7178 PathScan® Acetyl-Histone H2B Sandwich ELISA Kit	H, Mk
#7155 PathScan® Phospho-Histone H3 (Ser10) Sandwich ELISA Kit	H, M
#7207 PathScan® Phospho-Histone H3 (Ser10) Sandwich ELISA Antibody Pair	H, M
#7123 PathScan® Mono-Methyl-Histone H3 (Lys4) Sandwich ELISA Kit	H, M, Mk
#7124 PathScan® Di-Methyl-Histone H3 (Lys4) Sandwich ELISA Kit	H, M, Mk
#7125 PathScan® Tri-Methyl-Histone H3 (Lys4) Sandwich ELISA Kit	H, M, Mk
#7862 PathScan® Di-Methyl-Histone H3 (Lys9) Sandwich ELISA Kit	H, M, Mk
#7864 PathScan® Pan-Methyl-Histone H3 (Lys9) Sandwich ELISA Kit	H, M, Mk
#7866 PathScan® Tri-Methyl-Histone H3 (Lys27) Sandwich ELISA Kit	H, M, Mk
#7868 PathScan® Di-Methyl-Histone H3 (Lys36) Sandwich ELISA Kit	H, M, Mk
#7121 PathScan® Acetyl-Histone H3 (Lys9) Sandwich ELISA Kit	H, M, Mk
#7122 PathScan® Acetyl-Histone H3 (Lys18) Sandwich ELISA Kit	H, M, Mk
#7232 PathScan® Acetylated Histone H3 Sandwich ELISA Kit	H, M, Mk
#7209 PathScan® Acetylated Histone H3 Sandwich ELISA Antibody Pair	H, M, Mk
#7253 PathScan® Total Histone H3 Sandwich ELISA Kit	H, M, Mk
#7224 PathScan® Acetyl-Histone H4 (Lys8) Sandwich ELISA Kit	H, M, Mk
#7228 PathScan® Acetyl-Histone H4 (Lys12) Sandwich ELISA Kit	H, M, Mk
#7238 PathScan® Acetyl-Histone H4 Sandwich ELISA Kit	H, M, Mk

Scientists at CST develop the highest quality research tools that are robust, sensitive, and specific. Our line of PathScan® Sandwich ELISA products provides researchers with a selection of assays for the study of critical regulatory proteins. In-house development, production, and validation of these kits ensure the highest possible product quality. Technical support is provided by the same scientists that produce the products, and know them best.



COS cells were treated with either Trichostatin A #9950 or Calyculin A #9902 to inhibit HDACs or phosphatases, respectively. Histone acetylation or phosphorylation was analyzed using PathScan® Sandwich ELISA Kits: #7155, #7121, #7122, #7232, #7124, #7253.



◀ **PathScan® Phospho-Histone H3 (Ser10) Sandwich ELISA Kit #7155:** Treatment of NIH/3T3 cells with Calyculin A #9902 causes accumulation of phospho-histone H3 (Ser10), detected by #7155, but does not affect the level of total histone H3 protein, detected by western analysis. OD_{450nm} readings are shown in (A), while the corresponding western blots using Phospho-Histone H3 (Ser10) Antibody #9701 (left panel) or Histone H3 Antibody #9715 (right panel), are shown in (B).

Supporting Products



For your convenience, CST offers a wide selection of products to support your research needs. These products are the same used in-house for antibody validation in applications including western blotting, chromatin immunoprecipitation, immunohistochemistry, flow cytometry, and immunofluorescent analysis, and therefore work optimally with CST™ antibodies.

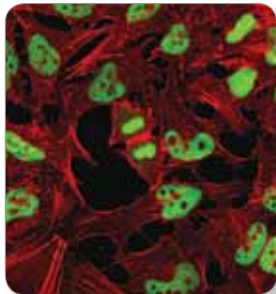
Complementary Products include:

- ▣ Growth Factors and Cytokines
- ▣ Inhibitors and Activators
- ▣ Secondary Antibodies
- ▣ Protein Markers
- ▣ Detection Systems
- ▣ Buffers
- ▣ Cellular Dyes

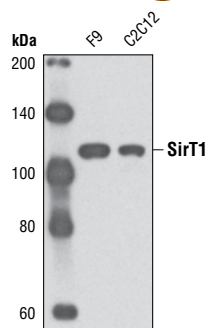
Control Products include:

- ▣ Cell Extracts
- ▣ Control Proteins
- ▣ Isotype Controls
- ▣ Blocking Peptides
- ▣ PCR Primers

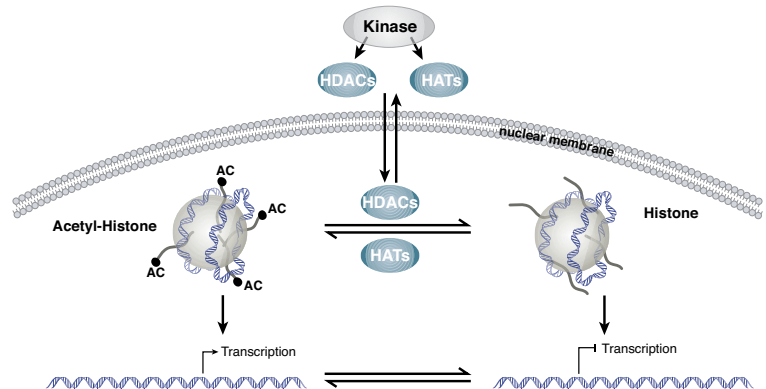
Epigenetic Regulators



HDAC1 (10E2) Mouse mAb #5356:
Confocal IF analysis of HeLa cells using #5356 (green). Actin filaments were labeled with DY-554 phalloidin (red).

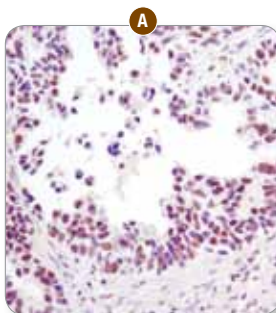


SirT1 (D60E1) Rabbit mAb (Mouse Specific) #3931: WB analysis of extracts from F9 and C2C12 cells using #3931.

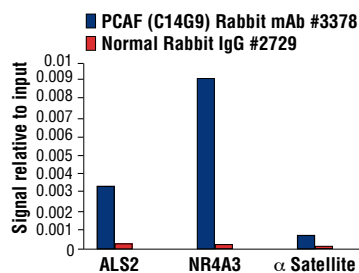
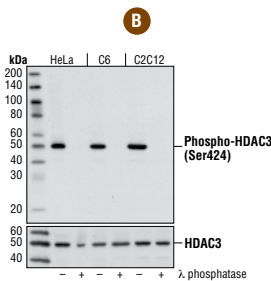


Histone Acetylases and Deacetylases

	Application	Reactivity
#4771 Acetyl-CBP (Lys1535)/p300 (Lys1499) Antibody	W, IP, ChIP	H, M, R, Mk
#4772 CBP Antibody	W	H, Mk
NEW #5157 CLOCK (D45B10) Rabbit mAb	W, IP	H, M, R, Mk
#3305 GCN5L2 (C26A10) Rabbit mAb	W, IP, IF-IC	H, M, R, Mk, (B)
#2062 Histone Deacetylase 1 (HDAC1) Antibody	W	H, M, R, Mk
NEW #5356 HDAC1 (10E2) Mouse mAb <i>featured</i>	W, IP, IF-IC	H, M, R, Mk
#2540 HDAC2 Antibody	W, IF-IC	H, M, R, Mk
#2545 HDAC2 Antibody (IP Preferred)	W, IP	H, M, Mk
NEW #5113 HDAC2 (3F3) Mouse mAb	W, IP, IF-IC	H, M, R, Mk
NEW #3815 Phospho-HDAC3 (Ser424) Antibody <i>featured</i>	W, IP, IHC-P, IF-IC	H, M, R, (Mk, C, X)
#2632 Histone Deacetylase 3 (HDAC3) Antibody	W	H, M, R, Mk
NEW #3949 HDAC3 (7G6C5) Mouse mAb	W, IP, IF-IC	H, M, R, Mk
#3443 Phospho-HDAC4 (Ser246)/HDAC5 (Ser259)/HDAC7 (Ser155) (D27B5) Rabbit mAb	W, IP	H, M
#3424 Phospho-HDAC4 (Ser632)/HDAC5 (Ser498)/HDAC7 (Ser486) Antibody	W, IP	H, M
#2072 Histone Deacetylase 4 (HDAC4) Antibody	W	H, M, R, Mk
NEW #5392 HDAC4 (4A3) Mouse mAb	W, IP	H, M, R, Mk
#2082 Histone Deacetylase 5 (HDAC5) Antibody	W, IP, IHC-P	H, M, R, Mk
#2882 Histone Deacetylase 7 (HDAC7) Antibody	W	H, M, R, Mk
#3378 PCAF (C14G9) Rabbit mAb <i>featured</i>	W, IP, ChIP	H, M, R, Mk, (B)
#2327 Phospho-SirT1 (Ser27) Antibody	W	H
#2314 Phospho-SirT1 (Ser47) Antibody	W, IP, IF-IC, F	H
NEW #3931 SirT1 (D60E1) Rabbit mAb (Mouse Specific) <i>featured</i>	W, IP	M
#2496 SirT1 (C14H4) Rabbit mAb	W, IP	H
#2310 SirT1 Antibody	W	H
#2493 SirT1 (D739) Antibody	W, IP, IF-IC	H, Mk
#2028 SirT1 Antibody (Mouse Specific)	W, IP, IF-IC	M
#2313 SirT2 Antibody	W	H, M, R, Mk
#2627 SirT3 (C73E3) Rabbit mAb	W, IP, IHC-P	H, R, Mk
#2590 SirT6 Antibody	W, IP, IF-IC	H
#4562 TFII-I Antibody	W, IP, IHC-P	H, M, Mk
NEW #4169 TFII-B (2F6A3H4) Mouse mAb	W, IP	H, M, R, Mk
#3966 TRRAP (D2966) Antibody	W, IP, IF-IC	H, M, R, Mk
#3967 TRRAP (P2032) Antibody	W, IP	H, M, Mk
#2152 WSTF Antibody	W, IP	H, M, R, Mk
#9950 Trichostatin A (TSA)		



Phospho-HDAC3 (S424) Antibody #3815: (A) IHC analysis of paraffin-embedded human lung carcinoma using #3815. (B) WB analysis of extracts from HeLa, C6, and C2C12 cells, untreated or λ -phosphatase-treated, using #3815 (upper) or HDAC3 Antibody #2632 (lower).



PCAF (C14G9) Rabbit mAb #3378: ChIP assays were performed with cross-linked chromatin from 4×10^6 293 cells treated with Forskolin #3828 (30 μ M) and either 20 μ l of #3378 or 2 μ l of Normal Rabbit IgG #2729 using SimpleChIP[®] Enzymatic Chromatin IP Kit (Magnetic Beads) #9003. The enriched DNA was quantified by real-time PCR using human ALS2 exon 1 primers, SimpleChIP[®] Human NR4A3 Promoter Primers #4829, and SimpleChIP[®] Human α Satellite Repeat Primers #4486. The amount of immunoprecipitated DNA in each sample is represented as signal relative to the total amount of input chromatin (equivalent to one).

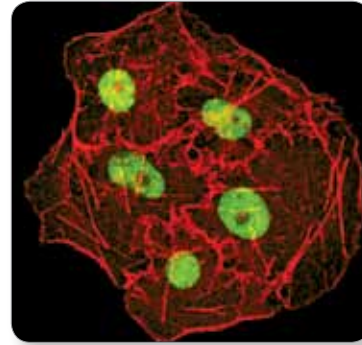
Selected Application References:

Histone Deacetylase 5 (HDAC5) Antibody #2082:

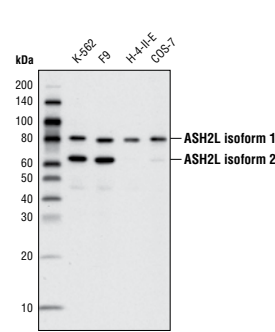
Xia, S. et al. (2010) *Development* 137, 1075-1084. (W)

Methyltransferases and Demethylases

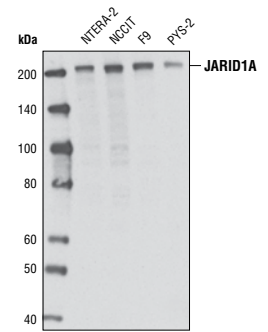
		Application	Reactivity
NEW	#5019 ASH2L (D93F6) XP™ Rabbit mAb <i>featured</i>	W, IP, IF-IC	H, M, R, Mk, (Dm)
NEW	#5032 DNMT1 (D63A6) XP™ Rabbit mAb <i>featured</i>	W, IF-IC	H, M, R, Mk, (B)
NEW	#5119 DNMT1 (D59A4) Rabbit mAb	W, IP	H, M, R, Mk, (B)
	#3598 DNMT3A (D23G1) Rabbit mAb	W, IP	H, M, R, Mk, (B)
	#2160 DNMT3A Antibody	W, IP	H, M, R, Mk, (B)
	#2161 DNMT3B Antibody	W, IP	H, M, R, Mk, (Z)
	#2196 ESET (C1C12) Rabbit mAb	W, IP, IF-IC	H, Mk
NEW	#5246 Ezh2 (D2C9) XP™ Rabbit mAb <i>featured</i>	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk
	#4905 Ezh2 Antibody	W, IP, ChIP	H, M, R, Pg
	#3147 Ezh2 (AC22) Mouse mAb	W, IF-IC	H, M, R, Mk
	#3306 G9a/EHMT2 (C6H3) Rabbit mAb	W, IF-IC	H, M, R, Mk, (B, Pg)
NEW	#3876 JARID1A (D28B10) XP™ Rabbit mAb <i>featured</i>	W, IP, IF-IC	H, M, (R, B)
	#3273 JARID1B Antibody	W, IP	H, Mk
NEW	#5361 JARID1C (D29B9) Rabbit mAb	W, IP	H, M, (Mk, Pg)
	#2621 JMJD1B/JHDM2B Antibody	W, IP, IF-IC	H, M, R, Mk
	#3314 JMJD1B (C69G2) Rabbit mAb	W, IP, IF-IC	H, Mk
	#3100 JMJD1B (C6D12) Rabbit mAb	W, IP, IHC-P	H, Mk
NEW	#5377 JMJD1B (6A1-1F5) Mouse mAb	W, IP, IF-IC	H, M, R, Mk
	#3393 JMJD2A (C70G6) Rabbit mAb	W, IP, IF-IC	H, M, R, (Mk)
	#2898 JMJD2B Antibody	W, IP	H, (Mk)
	#3457 JMJD3 Antibody	W	H, M, (Mk)
	#2184 LSD1 (C69G12) Rabbit mAb	W, IP, IHC-P, IHC-F, IF-IC	H, M, R, Mk
	#2139 LSD1 Antibody	W, IP, IHC-P, IF-IC, F	H, M, R, Mk
	#4064 LSD1 (1B2E5) Mouse mAb	W	H, M, R, Mk
NEW	#4218 LSD1 (1E5-H2) Mouse mAb	W, IP	H, Mk
	#2018 MEP50 (D56B8) Rabbit mAb	W, IP	H, M, R, Mk
	#2828 MEP50 (P328) Antibody	W, IP	H
	#2823 MEP50 Antibody	W, IP, IF-IC	H
NEW	#4789 Nucleomethylin Antibody	W	H
	#2449 PRMT1 (A33) Antibody	W, IP, IF-IC	H, M, R, Mk, (B)
	#2453 PRMT1 (F339) Antibody	W	H, M, R, Mk, (B)
	#3379 PRMT4/CARM1 (C31G9) Rabbit mAb	W, IP, IF-IC	H, M, R, Mk
	#4438 PRMT4/CARM1 Antibody	W, IP	H, M, R, Mk
	#2252 PRMT5/Skb1Hs Methyltransferase Antibody	W, IP	H, M, R, Mk
	#2825 SET7/SET9 (C24B1) Rabbit mAb	W	H, M, R, Mk
	#2813 SET7/SET9 Antibody	W, IF-IC	H, M, R, Mk
	#2996 SET8 (C18B7) Rabbit mAb	W, IF-IC	H, M, R, Mk, (B, Pg)
NEW	#4251 SMYD2 Antibody	W	H, M, R, Mk, (B, Pg)
	#2991 SUV39H1 Histone Methyltransferase Antibody	W	H, M
	#3737 SUZ12 (D39F6) XP™ Rabbit mAb	W, IP, IF-IC, ChIP	H, M, R, Mk



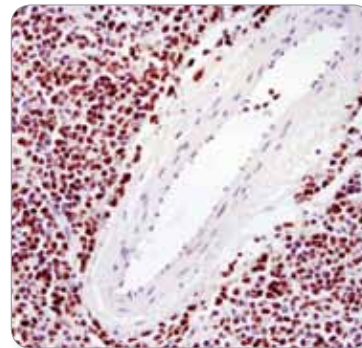
◀ **DNMT1 (D63A6) XP™ Rabbit mAb #5032:** Confocal IF analysis of COS-7 cells using #5032 (green). Actin filaments were labeled using DY-554 phalloidin (red).



ASH2L (D93F6) XP™ Rabbit mAb #5019: WB analysis of extracts from various cell lines using #5019.



JARID1A (D28B10) XP™ Rabbit mAb #3876: WB analysis of extracts from various cell lines using #3876.



◀ **Ezh2 (D2C9) XP™ Rabbit mAb #5246:** IHC analysis of paraffin-embedded human lymphoma using #5246.

Selected Application References:

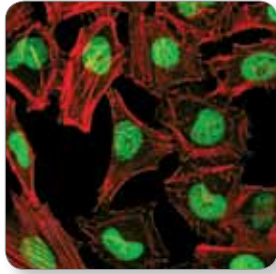
Ezh2 (AC22) Mouse mAb #3147:

Kalushkova, A. et al. (2010) *PLoS One* 5, e11483. (W)

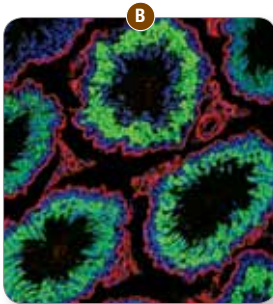
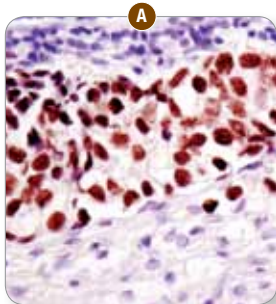
Van Dessel, N. et al. (2010) *Nucleic Acids Res.*, in press. (W)

UNPARALLELED PRODUCT QUALITY, VALIDATION, AND TECHNICAL SUPPORT

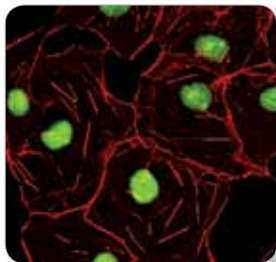
HP1 and Other Chromatin Proteins



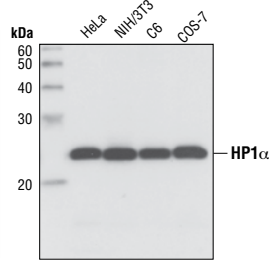
◀ CHAF1A (D77D5) XP™ Rabbit mAb #5480: Confocal IF analysis of HeLa cells using #5480 (green). Actin filaments were labeled with DY-554 phalloidin (red).



NUT (C52B1) Rabbit mAb #3625: (A) IHC analysis of paraffin-embedded human midline carcinoma using #3625. (B) Confocal IF analysis of rat testes using #3625 (green) and Pan-Keratin (C11) Mouse mAb #4545 (red). Blue pseudocolor = DRAQ5® #4084 (fluorescent DNA dye).



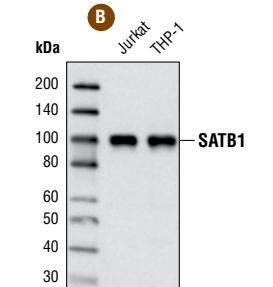
HMGB1 Antibody #3935: Confocal IF analysis of COS-7 cells using #3935 (green). Actin filaments were labeled using DY-554 phalloidin (red).



HP1α (C7F11) Rabbit mAb #2623: WB analysis of extracts from HeLa, NIH/3T3, C6, and COS-7 cells using #2623.



SATB1 (L745) Antibody #3650: (A) Confocal IF analysis of 293 cells using #3650 (green). Actin filaments were labeled with DY-554 phalloidin (red). (B) WB analysis of extracts from Jurkat and THP-1 cell lines using #3650.



HP1

		Application	Reactivity
#2623	HP1α (C7F11) Rabbit mAb <i>featured</i>	W, IP, IHC-P, IF-IC	H, M, R, Mk
#2616	HP1α Antibody	W, IP, IHC-P, IF-IC, F	H, M, R, Mk, (B)
#2613	HP1β Antibody	W	H, M, R, Mk, (B)
#2600	Phospho-HP1γ (Ser83) Antibody	W, IP, IF-IC	H, M, R, Mk, (Dm, B)
#2619	HP1γ Antibody	W, IP, IF-IC, F	H, M, R, Mk

Other Chromatin Proteins

#2990	ASF1A (C6E10) Rabbit mAb	W, IP, IHC-P, IF-IC	H, M, Mk, (C, B)
#2902	ASF1B (C70E2) Rabbit mAb	W, IP, IF-IC	H, Mk
#2769	ASF1B Antibody	W, IP	H, Mk, (M)
#2830	Bmi1 Antibody	W, IF-IC	H, R, Mk, (B)
#3508	Brg1 (A52) Antibody	W, IF-IC	H, M, Mk, (R)
#3514	Brg1 (P680) Antibody	W	H, M, R, Mk
NEW #5480	CHAF1A (D77D5) XP™ Rabbit mAb <i>featured</i>	W, IP, IF-IC	H, Mk
NEW #4170	CHD2 Antibody	W	H, M, R, Mk
NEW #4241	CHD3 Antibody	W, IP	H, M, Mk
NEW #4245	CHD4 Antibody	W	H, M, R, (Mk, B)
#3417	CTCF (D1A7) XP™ Rabbit mAb	W, IP, IF-IC, ChIP	H, R, Mk, (B)
#3418	CTCF (D31H2) XP™ Rabbit mAb	W, IP, IHC-P, IF-IC, ChIP	H, M, R, Mk, (B)
#2899	CTCF Antibody	W, IP, IF-IC, ChIP	H, M, R, Mk
NEW #4880	Phospho-DBC1 (Thr454) Antibody	W, IP, IF-IC	H
NEW #5693	DBC1 Antibody	W, IP	H, M, R, Mk
NEW #5269	HMG A2 Antibody	W, IP	H, M, R
NEW #3935	HMGB1 Antibody <i>featured</i>	W, IF-IC	H, M, R, Mk, (Hm, B, Pg)
#2088	LEDGF (C57G11) Rabbit mAb	W, IHC-P, IF-IC, F	H, M, R, (Mk)
#3896	MBD3 Antibody	W	H, M, R, Mk
#3456	MeCP2 (D4F3) XP™ Rabbit mAb	W, IP, IHC-P, IF-IC	H, M, R, Mk
NEW #3625	NUT (C52B1) Rabbit mAb <i>featured</i>	W, IP, IHC-P, IF-F	H, R, (Mk)
NEW #3497	PHF2 (D45A2) Rabbit mAb	W, IP	H, M, R, Mk
NEW #3934	PHF20 (D96F6) XP™ Rabbit mAb	W, IP, IF-IC	H, M, R, Mk, (B)
NEW #4522	RBAP46 Antibody	W	H, M, R, Mk, (B)
NEW #4633	RBAP46/RBAP48 Antibody	W	H, M, R, Mk, (C)
#2820	Ring1A Antibody	W	H, M, R, Mk
NEW #5694	RING1B (D22F2) XP™ Rabbit mAb	W, IP, IF-IC, ChIP	H, M, R, Mk
NEW #4028	Phospho-SATB1 (Ser47) Antibody	W	H, M, R, (Mk, B)
NEW #3650	SATB1 (L745) Antibody <i>featured</i>	W, IP, IF-IC	H, (Mk)
NEW #3643	SATB1 (P472) Antibody	W, IP	H, M, R, (Mk, B, Pg)
NEW #5329	SMC3 (D23C5) Rabbit mAb	W, IP	H, M, R, Mk
NEW #5394	SMC2 (D91E3) Rabbit mAb	W, IP	H, R, Mk
NEW #5696	SMC3 (D47B5) Rabbit mAb	W, IP, IF-IC	H, M, R, Mk
NEW #5547	SMC4 (D14E2) Rabbit mAb	W, IP	H, M, R, Mk, (X, B)
NEW #4239	STAG2 Antibody	W, IP	H, M, R, Mk
NEW #5433	TERF2IP (D9H4) Rabbit mAb	W, IP	H, M, R, Mk
#4733	Topoisomerase IIα Antibody	W, IP, F	H, M, R, Mk

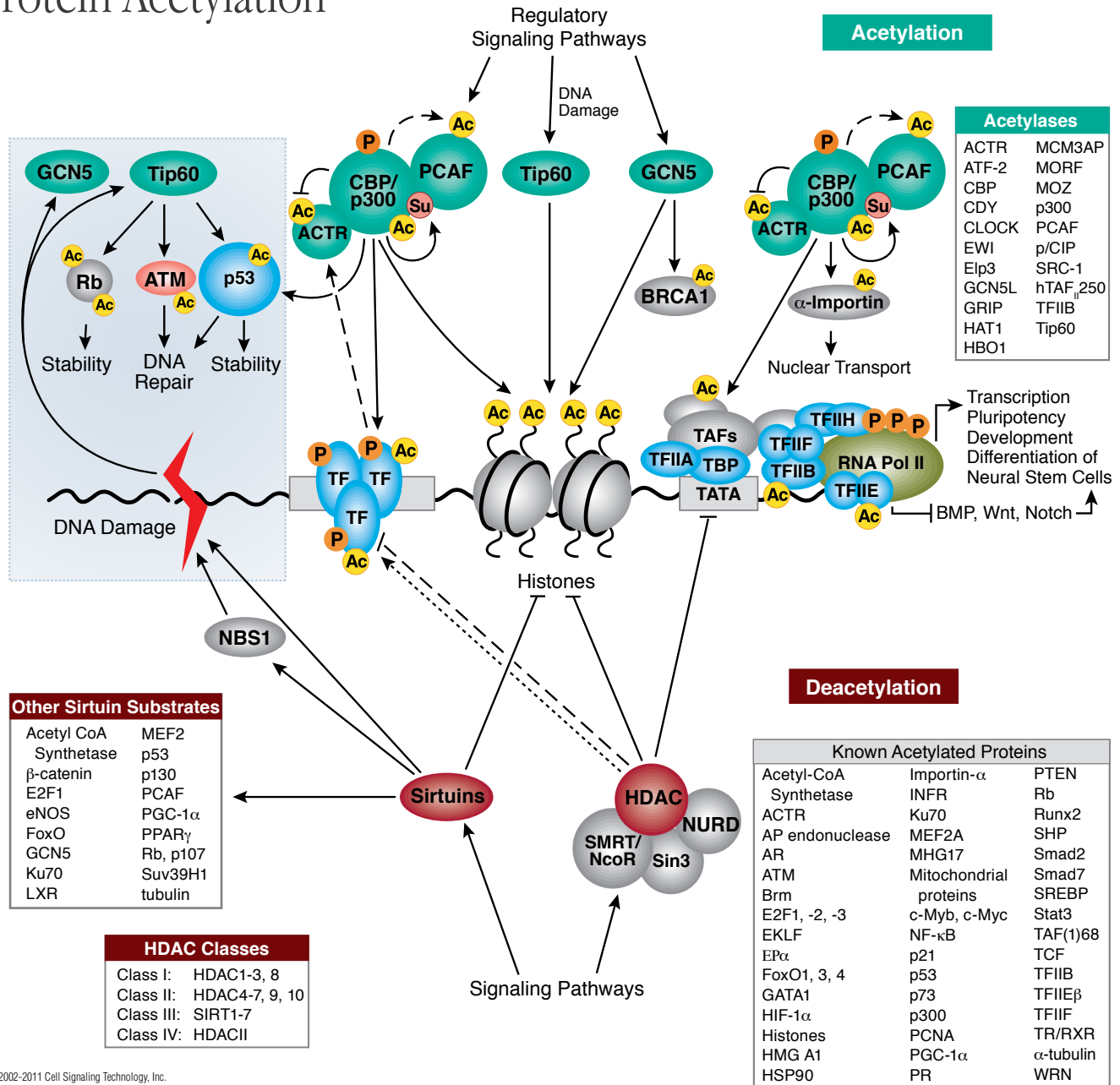
Selected Application References:

ASF1B (C70E2) Rabbit mAb #2902: Jasencakova, Z. et al. (2010) *Mol. Cell* 37, 736-743. (IF-IC)

NUT (C52B1) Rabbit mAb #3625: Haack, H. et al. (2009) *Am. J. Surg. Pathol.* 33, 984-991. (IHC-P)

Signaling Pathways

Protein Acetylation



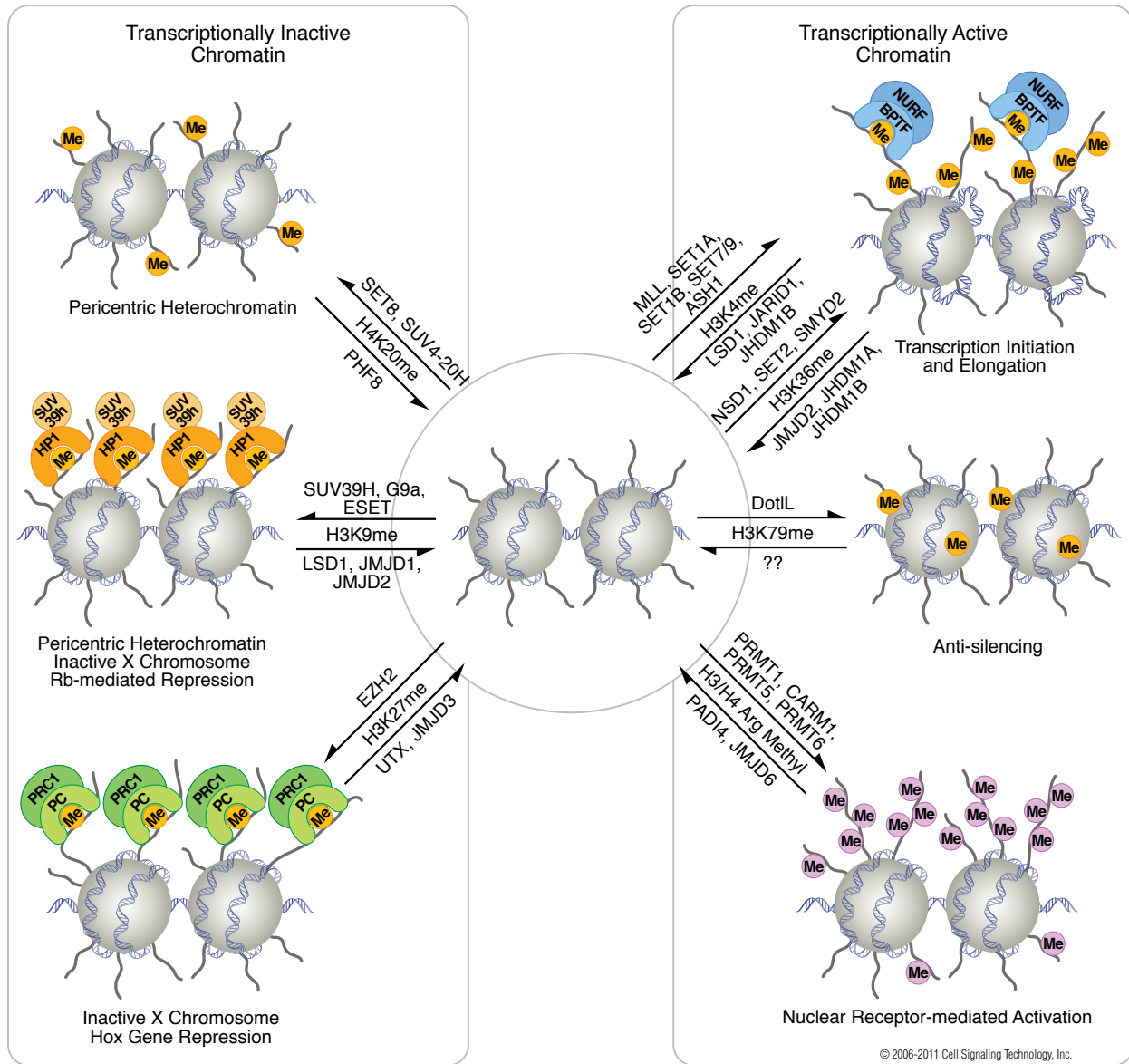
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Pathway Description: Protein acetylation plays a crucial role in regulating chromatin structure and transcriptional activity. Many transcriptional coactivators possess intrinsic acetylase activity, while transcriptional corepressors are associated with deacetylase activity. Acetylation complexes (such as CBP/p300 and PCAF) or deacetylation complexes (such as Sin3, NuRD, NcoR and SMRT) are recruited to DNA-bound transcription factors (TFs) in response to signaling pathways. Histone hyperacetylation by histone acetyltransferases (HATs) is associated with transcriptional activation, whereas histone deacetylation by histone deacetylases (HDACs) is associated with transcriptional repression. Histone acetylation stimulates transcription by remodeling higher order chromatin structure, weakening histone-DNA interactions, and providing binding sites for transcriptional activation complexes containing proteins that possess bromodomains, which bind acetylated lysine. Histone deacetylation represses transcription through

an inverse mechanism involving the assembly of compact higher order chromatin and the exclusion of bromodomain-containing transcription activation complexes. Histone hypoacetylation is a hallmark of silent heterochromatin. Site-specific acetylation of a growing number of non-histone proteins, including p53 and E2F, has been shown to regulate their activity, localization, specific interactions, and stability/degradation, therefore controlling a variety of cellular processes, such as transcription, proliferation, apoptosis, and differentiation. At an organismal level, acetylation plays an important role in immunity, circadian rhythmicity, and memory formation. Protein acetylation is becoming a favorable target in drug design for numerous disease conditions.

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Histone Methylation



Pathway Description: The nucleosome, made up of four histone proteins (H2A, H2B, H3, and H4), is the primary building block of chromatin. Originally thought to function as a static scaffold for DNA packaging, histones have more recently been shown to be dynamic proteins, undergoing multiple types of post-translational modifications. Two such modifications, methylation of arginine and lysine residues are major determinants for formation of active and inactive regions of the genome. Arginine methylation of histones H3 (Arg2, 17, 26) and H4 (Arg3) promotes transcriptional activation and is mediated by a family of protein arginine methyltransferases (PRMTs), including the co-activators PRMT1 and CARM1 (PRMT4). In contrast, a more diverse set of histone lysine methyltransferases has been identified, all but one of which contain a conserved catalytic SET domain originally identified in the *Drosophila* Su(var)3-9, Enhancer of zeste, and Trithorax proteins. Lysine methylation has been implicated in both transcriptional activation (H3 Lys4, 36, 79) and silencing (H3 Lys9, 27, H4 Lys20).

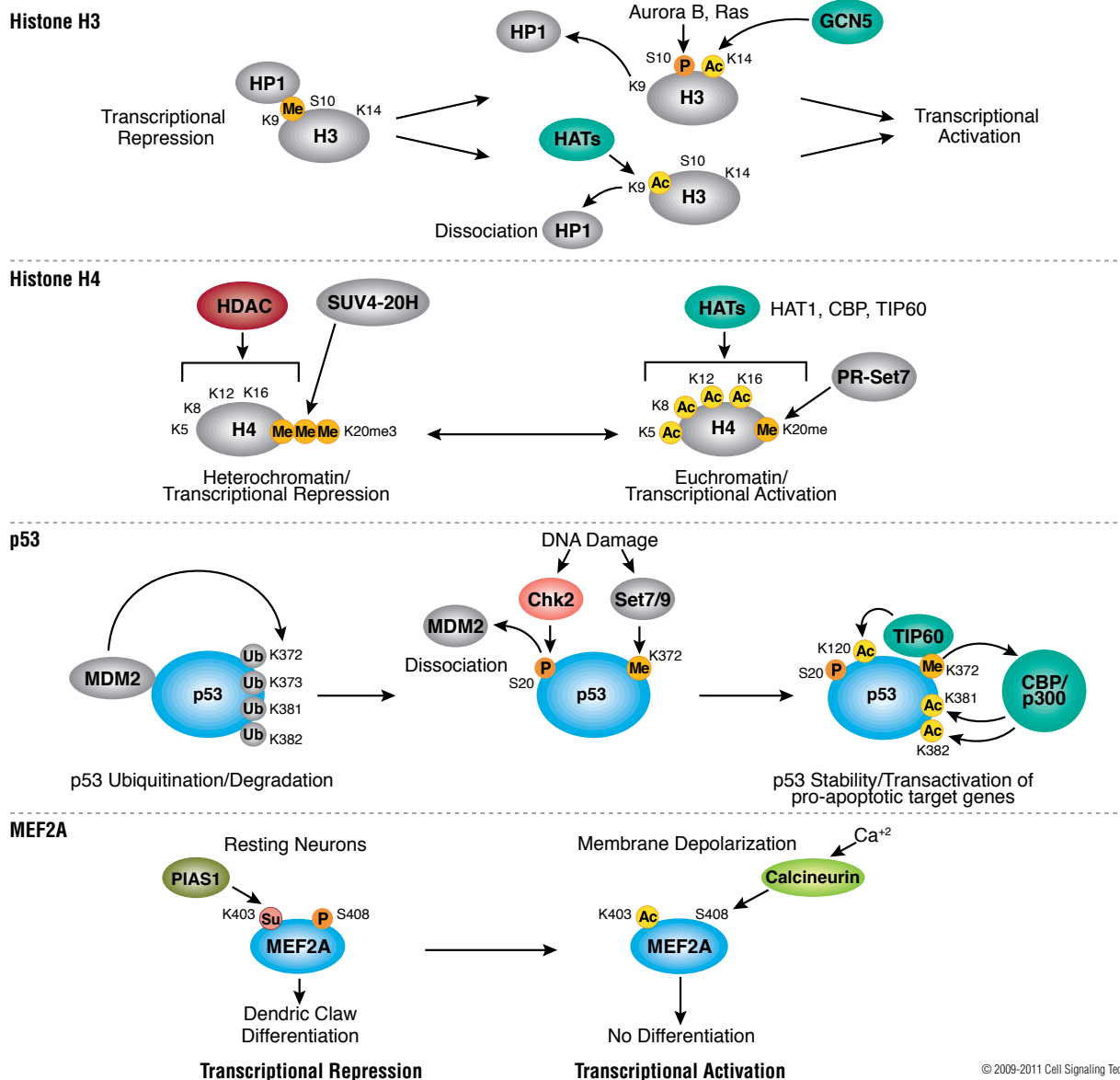
Unlike acetylation, methylation does not alter the charge of arginine and lysine residues and is unlikely to directly modulate nucleosomal interactions required for chromatin folding. While the mechanisms by which arginine methylation regulates transcription are unknown, lysine methylation coordinates the recruitment of chromatin modifying enzymes. Chromodomains (HP1, PRC1), PHD fingers (BPTF, ING2), Tudor domains (53BP1), and WD-40 domains (WDR5) are among a growing list of methyl-lysine binding

modules found in histone acetyltransferases, deacetylases, methylases and ATP-dependent chromatin remodeling enzymes. Lysine methylation provides a binding surface for these enzymes, which then regulate chromatin condensation and nucleosome mobility in order to maintain local regions of active or inactive chromatin. In addition, lysine methylation can block binding of proteins that interact with unmethylated histones or directly inhibit catalysis of other regulatory modifications on neighboring residues. The presence of methyl-lysine binding modules in the DNA repair protein 53BP1 suggests roles for lysine methylation in other cellular processes.

Histone methylation is crucial for proper programming of the genome during development and misregulation of the methylation machinery can lead to diseased states such as cancer. Until recently, methylation was believed to be an irreversible, stable epigenetic mark that is propagated through multiple cell divisions, maintaining a gene in an active or inactive state. While there is no argument that methylation is a stable mark, recent identification of histone demethylases such as LSD1/AOF2, JMJD1, JMJD2 and JHDM1 has shown that methylation is reversible and provides a rationale for how genomes might be reprogrammed during differentiation of individual cell lineages.

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Examples of Crosstalk Between Post-translational Modifications



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Pathway Description: Post-translational modifications (PTMs) have recently emerged as major regulators of protein function. Originally described in histones, these various chemical modifications (methylation, acetylation, phosphorylation, sumoylation, and more) have now been identified in non-histone proteins as well. Early work defined a putative role for each of these modifications, for instance, acetylation correlates with activation and methylation with repression. However, more recent studies indicate that some of these modifications could trigger either activation or silencing in a context dependent manner. For instance, methylation of histone H3 Lys9 correlates with repression, while methylation of H3 Lys4 correlates with activation. Furthermore, each of these moieties can be either mono-, di- or tri-methylated, and depending on the degree of methylation, the biological output will be completely different. Until recently, PTMs were considered independently, under the assumption that their functions would not be related to one another. It is now clear that PTMs work in concert, and the crosstalk between different modifications determines the final biological read-out. In this context, some modifications can influence others, and it appears that specific combinations of these modifications can form a dynamic "code". We provide a few examples of this type of crosstalk above. Although each of the modifications

shown here are occurring in cis, there are now clear examples, at least for histones, where modifications in one histone molecule can regulate modifications in other histones in trans. Although there are now many examples of these "functional networks", it is likely that we have just begun to scratch the surface. Better antibodies and novel technologies will help to complete this crosstalk puzzle, for which the specific fine-tuning appears critical to determine life as we know it.

Selected Reviews:

Latham, J.A. and Dent, S.Y. (2007) Cross-regulation of histone modifications. *Nat. Struct. Mol. Biol.* 14, 1017–1024.
 Yang, X.J., and Seto, E. (2008) Lysine acetylation: codified crosstalk with other posttranslational modifications. *Mol. Cell* 31, 449–461.
 Kouzarides, T. (2007) Chromatin modifications and their function. *Cell* 128, 693–705.
 Berger, S.L. (2007) The complex language of chromatin regulation during transcription. *Nature* 447, 407–412.

Histone Modification Table

Acetylation

Histone	Site	Histone-modifying Enzymes	Proposed Function	Ref. #	
H2A	Lys4 (<i>S. cerevisiae</i>)	Esa1	transcriptional activation	(1)	
	Lys5 (mammals)	Tip60, p300/CBP	transcriptional activation	(2,3)	
	Lys7 (<i>S. cerevisiae</i>)	Hat1 Esa1	unknown transcriptional activation	(4) (1)	
H2B	Lys5	p300, ATF2	transcriptional activation	(3,5)	
	Lys11 (<i>S. cerevisiae</i>)	Gcn5	transcriptional activation	(6)	
	Lys12 (mammals)	p300/CBP, ATF2	transcriptional activation	(3,5)	
	Lys15 (mammals)	p300/CBP, ATF2	transcriptional activation	(3,5)	
	Lys16 (<i>S. cerevisiae</i>)	Gcn5, Esa1	transcriptional activation	(6)	
	Lys20	p300	transcriptional activation	(3)	
H3	Lys4 (<i>S. cerevisiae</i>)	Esa1	transcriptional activation	(1)	
		Hpa2	unknown	(7)	
	Lys9	unknown	histone deposition	(8)	
		Gcn5, SRC-1	transcriptional activation	(9,10)	
	Lys14	unknown	histone deposition	(8)	
		Gcn5, PCAF	transcriptional activation	(3,11)	
		Esal, Tip60	transcriptional activation	(1,2)	
			DNA repair	(11,12)	
		SRC-1	transcriptional activation	(10)	
		Elp3	transcriptional activation (elongation)	(13)	
		Hpa2	unknown	(7)	
		hTFIIIC90	RNA polymerase III transcription	(14)	
		TAF1	RNA polymerase II transcription	(15)	
		Sas2	euchromatin	(16)	
		Sas3	transcriptional activation (elongation)	(17)	
		p300	transcriptional activation	(3)	
		Lys18	Gcn5	transcriptional activation, DNA repair	(9)
	p300/CBP		DNA replication, transcriptional activation	(3,18)	
	Lys23	unknown	histone deposition	(8)	
		Gcn5	transcriptional activation, DNA repair	(9)	
		Sas3	transcriptional activation (elongation)	(17)	
		p300/CBP	transcriptional activation	(3,18)	
	Lys27	Gcn5	transcriptional activation	(6)	
	Lys56 (<i>S. cerevisiae</i>)	Spt10	transcriptional activation DNA repair	(19) (20)	
	H4	Lys5	Hat1	histone deposition	(21)
			Esal, Tip60	transcriptional activation DNA repair	(1,2) (11,12)
			ATF2	transcriptional activation	(5)
Hpa2			unknown	(7)	
p300			transcriptional activation	(3)	
Lys8		Gcn5, PCAF	transcriptional activation	(3,22)	
		Esal, Tip60	transcriptional activation DNA repair	(1,2) (11,12)	
		ATF2	transcriptional activation	(5)	
		Elp3	transcriptional activation (elongation)	(13)	
		p300	transcriptional activation	(3)	
Lys12		Hat1	histone deposition telomeric silencing	(21) (23)	
		Esal, Tip60	transcriptional activation DNA repair	(1,2) (11,12)	
		Hpa2	unknown	(7)	
		p300	transcriptional activation	(3)	
		Lys16	Gcn5	transcriptional activation	(22)
MOF (<i>D. melanogaster</i>)			transcriptional activation	(24)	
Esal, Tip60			transcriptional activation DNA repair	(1,2) (11,12)	
ATF2			transcriptional activation	(5)	
Sas2			euchromatin	(2,6)	
Lys91 (<i>S. cerevisiae</i>)			Hat1/Hat2	chromatin assembly	(25)

Methylation

Histone	Site	Histone-modifying Enzymes	Proposed Function	Ref. #
H1	Lys26	Ezh2	transcriptional silencing	(48,49)
H3	Lys4	Set1 (<i>S. cerevisiae</i>)	permissive euchromatin (di-Me)	(26)
		Set7/9 (vertebrates)	transcriptional activation (tri-Me)	(27)
		MLL, ALL-1	transcriptional activation	(28,29)
		Ash1 (<i>D. melanogaster</i>)	transcriptional activation	(30)
	Arg8	PRMT5	transcriptional repression	(31)
		Suv39h, Clr4	transcriptional silencing (tri-Me)	(32,33)
	Lys9	G9a	transcriptional repression genomic imprinting	(34)
		SETDB1	transcriptional repression (tri-Me)	(35)
	Lys27	Dim-5 (<i>N. crassa</i>), Kryptonite (<i>A. thaliana</i>)	DNA methylation (tri-Me)	(36,37)
		Ash1 (<i>D. melanogaster</i>)	transcriptional activation	(30)
		Arg17	CARM1	transcriptional activation
	Lys27	Ezh2	transcriptional silencing X inactivation (tri-Me)	(38)
		G9a	transcriptional silencing	(34)
	Lys36	Set2	transcriptional activation (elongation)	(39)
	Lys79	Dot1	euchromatin	(40)
			transcriptional activation (elongation) checkpoint response	(41) (42)
	H4	Arg3	PRMT1	transcriptional activation
PRMT5			transcriptional repression	(31)
Lys20		PR-Set7	transcriptional silencing (mono-Me)	(44)
		Suv4-20h	heterochromatin (tri-Me)	(45)
Lys59		Ash1 (<i>D. melanogaster</i>)	transcriptional activation	(30)
		Set9 (<i>S. pombe</i>)	checkpoint response	(46)
Lys59		unknown	transcriptional silencing	(47)

Phosphorylation

Histone	Site	Histone-modifying Enzymes	Proposed Function	Ref. #
H1	Ser27	unknown	transcriptional activation, chromatin decondensation	(48,49)
H2A	Ser1	unknown	mitosis, chromatin assembly	(50)
		MSK1	transcriptional repression	(51)
	Thr119 (<i>D. melanogaster</i>)	NHK1	mitosis	(52)
	Ser122 (<i>S. cerevisiae</i>)	unknown	DNA repair	(53)
	Ser129 (<i>S. cerevisiae</i>)	Mec1, Tel1	DNA repair	(54,55)
	Ser139 (mammalian H2A.X)	ATR, ATM, DNA-PK	DNA repair	(56-58)
H2B	Ser10 (<i>S. cerevisiae</i>)	Ste20	apoptosis	(59)
	Ser14 (vertebrates)	Msl1	apoptosis	(60)
		unknown	DNA repair	(61)
Ser33 (<i>D. melanogaster</i>)	TAF1	transcriptional activation	(62)	
H3	Thr3	Haspin/Gsg2	mitosis	(63)
	Ser10	Aurora-B kinase	mitosis, meiosis	(64,65)
		MSK1, MSK2	immediate-early gene activation	(66)
		IKK- α	transcriptional activation	(67)
		Snf1	transcriptional activation	(68)
		Thr11 (mammals)	Dlk/Zip	mitosis
Ser28 (mammals)	Aurora-B kinase	mitosis	(70)	
	MSK1, MSK2	immediate-early activation	(66,71)	
H4	Ser1	unknown	mitosis, chromatin assembly	(50)
		CK2	DNA repair	(72)

Ubiquitylation

Histone	Site	Histone-modifying Enzymes	Proposed Function	Ref. #
H2A	Lys119 (mammals)	Ring2	spermatogenesis	(73)
H2B	Lys120 (mammals)	UbcH6	meiosis	(74)
	Lys123 (S. cerevisiae)	Rad6	transcriptional activation euchromatin	(75)

Sumoylation

Histone	Site	Histone-modifying Enzymes	Proposed Function	Ref. #
H2A	Lys126 (S. cerevisiae)	Ubc9	transcriptional repression	(76)
H2B	Lys6 or Lys7 (S. cerevisiae)	Ubc9	transcriptional repression	(76)
	N-terminal tail (S. cerevisiae)	Ubc9	transcriptional repression	(77)

Biotinylation

Histone	Site	Histone-modifying Enzymes	Proposed Function	Ref. #
H2A	Lys9	biotinidase	unknown	(78)
	Lys13	biotinidase	unknown	(78)
H3	Lys4	biotinidase	gene expression	(79)
	Lys9	biotinidase	gene expression	(79)
	Lys18	biotinidase	gene expression	(79)
H4	Lys12	biotinidase	DNA damage response	(80,81)

Histone Modification Table Description: The nucleosome, made up of four core histone proteins (H2A, H2B, H3, and H4), and linker histone H1 are the primary building blocks of chromatin. Originally thought to function as a static scaffold for DNA packaging, histones have more recently been shown to be dynamic proteins, undergoing multiple types of post-translational modifications that regulate chromatin condensation and DNA accessibility. For example, acetylation of lysine residues has long been associated with histone deposition and transcriptional activation, and more recently found to be associated with DNA repair. Phosphorylation of serine and threonine residues facilitates chromatin condensation during mitosis and transcriptional activation of immediate-early genes. Methylation of lysine and arginine residues function as a major determinant for formation of transcriptionally active and inactive regions of chromatin and is crucial for proper programming of the genome during development. This table provides a referenced list of many known histone modifications, the associated modifying enzymes, and proposed functions.

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