AMPK Subunit Antibody Sampler Kit

✓ 1 Kit (7 x 20 µl)



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For Research Use Only. Not For Use In Diagnostic Procedures.

Products Included	Product #	Quantity	Mol. Wt.	Isotype
AMPKα1 Antibody	2795	20 μΙ	62 kDa	Rabbit IgG
AMPKα2 Antibody	2757	20 μΙ	62 kDa	Rabbit IgG
AMPKβ1 (71C10) Rabbit mAb	4178	20 μΙ	38 kDa	Rabbit IgG
AMPKβ2 Antibody	4148	20 μΙ	30 kDa	Rabbit IgG
AMPKγ1 Antibody	4187	20 μΙ	37 kDa	Rabbit IgG
AMPKγ2 Antibody	2536	20 μΙ	75 kDa	Rabbit IgG
AMPKγ3 Antibody	2550	20 μΙ	54 kDa	Rabbit IgG
Anti-rabbit IgG, HRP-linked Antibody	7074	100 μΙ		Goat

See www.cellsignal.com for individual component applications, species cross-reactivity, dilutions and additional application protocols.

Description: The AMPK Subunit Antibody Sampler Kit provides an economical means to investigate the role played by all AMPK subunits in cellular energy homeostasis. The kit includes enough antibody to perform two western blot experiments with each primary antibody.

Background: AMP-activated protein kinase (AMPK) is highly conserved from yeast to plants and animals and plays a key role in the regulation of energy homeostasis (1). AMPK is a heterotrimeric complex composed of a catalytic α subunit and regulatory β and γ subunits, each of which is encoded by two or three distinct genes (α 1, 2: β 1, 2; γ 1, 2, 3) (2). The kinase is activated by an elevated AMP/ATP ratio due to cellular and environmental stress, such as heat shock, hypoxia and ischemia (1). The tumor suppressor LKB1, in association with accessory proteins STRAD and MO25, phosphorylates AMPK α at Thr172 in the activation loop and this phosphorylation is required for AMPK activation (3-5). AMPK α is also phosphorylated at Thr258 and Ser485 (for α 1; Ser491 for α 2). The upstream kinase and the biological significance of these phosphorylation events have yet to be elucidated (6). The β 1 subunit is post-translationally modified by myristoylation and multisite phosphorylation including Ser24/25, Ser96, Ser101, Ser108 and Ser182 (6,7). Phosphorylation at Ser108 of the $\beta 1$ subunit seems to be required for the activation of AMPK enzyme, while phosphorylation at Ser24/25 and Ser182 affects AMPK localization (7). Several mutations in AMPKγ subunits have been identified, most of which are located in the putative AMP/ATP binding sites (CBS or Bateman domains). Mutations at these sites lead to reduction of AMPK activity and cause glycogen accumulation in heart or skeletal muscle (1,2). Accumulating evidence indicates that AMPK not only regulates the metabolism of fatty acids and glycogen, but also modulates protein synthesis and cell growth through EF2 and TSC2/mTOR pathways, as well as blood flow via eNOS/nNOS (1).

Specificity/Sensitivity: Each of the antibodies in the AMPK Subunit Antibody Sampler Kit detects endogenous levels of the specified AMPK protein. Antibodies do not cross-react with related AMPK subunit proteins.

Source/Purification: Polyclonal antibodies are produced by immunizing animals with synthetic peptides corresponding to residues surrounding Leu519 near the carboxy

Storage: Supplied in 10 mM sodium HEPES (pH 7.5), 150 mM NaCl, 100 μ g/ml BSA, 50% glycerol and less than 0.02% sodium azide. Store at -20° C. *Do not aliquot the antibodies.*

Recommended Antibody Dilutions:

Western blotting

1:1000

Please visit www.cellsignal.com for validation data and a complete listing of recommended companion products.

terminus of human AMPK α 1, corresponding to residues surrounding Ser500 of human AMPK α 2, near the amino terminus of human AMPK γ 1, surrounding Ser60 of human AMPK γ 2, and corresponding to the sequences of human AMPK β 2 and AMPK γ 3. Antibodies are purified by protein A and peptide affinity chromatography. Monoclonal antibody is produced by immunizing animals with synthetic peptides corresponding to residues surrounding Val176 of human AMPK β 1.

Background References:

- (1) Hardie, D.G. (2004) J Cell Sci 117, 5479-87.
- (2) Carling, D. (2004) Trends Biochem Sci 29, 18-24.
- (3) Hawley, S.A. et al. (1996) J Biol Chem 271, 27879-87.
- (4) Lizcano, J.M. et al. (2004) EMBO J 23, 833-43.
- (5) Shaw, R.J. et al. (2004) *Proc Natl Acad Sci USA* 101, 3329–35.
- (6) Woods, A. et al. (2003) J Biol Chem 278, 28434-42.
- (7) Warden, S.M. et al. (2001) Biochem J 354, 275-83.



Western Immunoblotting Protocol

For western blots, incubate membrane with diluted primary antibody in either 5% w/v BSA or nonfat dry milk, 1X TBS, 0.1% Tween® 20 at 4°C with gentle shaking, overnight. **NOTE:** Please refer to primary antibody datasheet or product webpage for recommended primary antibody dilution buffer and recommended antibody dilution.

A. Solutions and Reagents

NOTE: Prepare solutions with reverse osmosis deionized (RODI) or equivalent grade water.

- 20X Phosphate Buffered Saline (PBS): (#9808) To prepare 1 L 1X PBS: add 50 ml 20X PBS to 950 ml dH₂O, mix.
- 2. 10X Tris Buffered Saline (TBS): (#12498) To prepare 1 L 1X TBS: add 100 ml 10X to 900 ml dH_2O , mix.
- 3. 1X SDS Sample Buffer: Blue Loading Pack (#7722) or Red Loading Pack (#7723) Prepare fresh 3X reducing loading buffer by adding 1/10 volume 30X DTT to 1 volume of 3X SDS loading buffer. Dilute to 1X with dH₂O.
- 4. 10X Tris-Glycine SDS Running Buffer: (#4050) To prepare 1 L 1X running buffer: add 100 ml 10X running buffer to 900 ml dH₂0, mix.
- 5. 10X Tris-Glycine Transfer Buffer: (#12539) To prepare 1 L 1X transfer buffer: add 100 ml 10X transfer buffer to 200 ml methanol + 700 ml dH₂O, mix.
- 6. 10X Tris Buffered Saline with Tween® 20 (TBST): (#9997) To prepare 1 L 1X TBST: add 100 ml 10X TBST to 900 ml dH₂O, mix.
- 7. Nonfat Dry Milk: (#9999)
- Blocking Buffer: 1X TBST with 5% w/v nonfat dry milk; for 150 ml, add 7.5 g nonfat dry milk to 150 ml 1X TBST and mix well.
- 9. Wash Buffer: (#9997) 1X TBST
- 10. Bovine Serum Albumin (BSA): (#9998)
- 11. Primary Antibody Dilution Buffer: 1X TBST with 5% BSA or 5% nonfat dry milk as indicated on primary antibody datasheet; for 20 ml, add 1.0 g BSA or nonfat dry milk to 20 ml 1X TBST and mix well.
- 12. Biotinylated Protein Ladder Detection Pack: (#7727)
- 13. Prestained Protein Marker, Broad Range (Premixed Format): (#7720)
- 14. Blotting Membrane and Paper: (#12369) This protocol has been optimized for nitrocellulose membranes. Pore size 0.2 µm is generally recommended.
- **15. Secondary Antibody Conjugated to HRP:** anti-rabbit (#7074); anti-mouse (#7076)
- 16. Detection Reagent: LumiGLO® chemiluminescent reagent and peroxide (#7003) or SignalFire™ ECL Reagent (#6883)

B. Protein Blotting

A general protocol for sample preparation.

- 1. Treat cells by adding fresh media containing regulator for desired time.
- 2. Aspirate media from cultures; wash cells with 1X PBS; aspirate.
- 3. Lyse cells by adding 1X SDS sample buffer (100 µl per well of 6-well plate or 500 µl for a 10 cm diameter plate). Immediately scrape the cells off the plate and transfer the extract to a microcentrifuge tube. Keep on ice.
- Sonicate for 10–15 sec to complete cell lysis and shear DNA (to reduce sample viscosity).
- 5. Heat a 20 μ l sample to 95–100°C for 5 min; cool on ice.
- 6. Microcentrifuge for 5 min.
- 7. Load 20 µl onto SDS-PAGE gel (10 cm x 10 cm). NOTE: Loading of prestained molecular weight markers (#7720, 10 µl/lane) to verify electrotransfer and biotinylated protein ladder (#7727, 10 µl/lane) to determine molecular weights are recommended.
- 8. Electrotransfer to nitrocellulose membrane (#12369).

C. Membrane Blocking and Antibody Incubations

NOTE: Volumes are for 10 cm x 10 cm (100 cm²) of membrane; for different sized membranes, adjust volumes accordingly.

I. Membrane Blocking

- (Optional) After transfer, wash nitrocellulose membrane with 25 ml TBS for 5 min at room temperature
- 2. Incubate membrane in 25 ml of blocking buffer for 1 hr at room temperature.
- 3. Wash three times for 5 min each with 15 ml of TBST.

II. Primary Antibody Incubation

- Incubate membrane and primary antibody (at the appropriate dilution and diluent as recommended in the product datasheet) in 10 ml primary antibody dilution buffer with gentle agitation overnight at 4°C.
- 2. Wash three times for 5 min each with 15 ml of TBST.
- 3. Incubate membrane with the species appropriate HRP-conjugated secondary anti-body (#7074 or #7076 at 1:2000) and anti-biotin, HRP-linked Antibody (#7075 at 1:1000–1:3000) to detect biotinylated protein markers in 10 ml of blocking buffer with gentle agitation for 1 hr at room temperature.
- 4. Wash three times for 5 min each with 15 ml of TBST.
- 5. Proceed with detection (Section D).

D. Detection of Proteins

- 1. Incubate membrane with 10 ml LumiGLO® (0.5 ml 20X LumiGLO® #7003, 0.5 ml 20X peroxide, and 9.0 ml purified water) or 10 ml SignalFire™ #6883 (5 ml Reagent A, 5 ml Reagent B) with gentle agitation for 1 min at room temperature.
- 2. Drain membrane of excess developing solution (do not let dry), wrap in plastic wrap and expose to x-ray film. An initial 10 sec exposure should indicate the proper exposure time. NOTE: Due to the kinetics of the detection reaction, signal is most intense immediately following incubation and declines over the following 2 hr.