

Rig-I Pathway Antibody Sampler Kit

✓ 1 Kit
 (9 x 20 µl)



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Products Included	Product #	Quantity	Mol. Wt.	Isotype
MDA-5 (D74E4) Rabbit mAb	5321	20 µl	135 kDa	Rabbit IgG
Rig-I (D14G6) Rabbit mAb	3743	20 µl	102 kDa	Rabbit IgG
MAVS Antibody	3993	20 µl	75, 52 kDa	Rabbit IgG
IRF-3 (D6I4C) XP® Rabbit mAb	11904	20 µl	50-55 kDa	Rabbit IgG
TBK1/NAK (D1B4) Rabbit mAb	3504	20 µl	84 kDa	Rabbit IgG
Phospho-TBK1/NAK (Ser172) (D52C2) XP® Rabbit mAb	5483	20 µl	84 kDa	Rabbit IgG
Phospho-IRF-3 (Ser396) (4D4G) Rabbit mAb	4947	20 µl	45-55 kDa	Rabbit IgG
Phospho-IKKε (Ser172) (D1B7) Rabbit mAb	8766	20 µl	80 kDa	Rabbit IgG
IKKε (D20G4) Rabbit mAb	2905	20 µl	80 kDa	Rabbit IgG
Anti-rabbit IgG, HRP-linked Antibody	7074	100 µl		Goat

See www.cellsignal.com for individual component applications, species cross-reactivity, dilutions and additional application protocols.

Description: The Rig-I Pathway Antibody Sampler Kit provides an economical means to evaluate the activation state and total protein levels of multiple members of the Rig-I pathway including Rig-I, MDA-5, MAVS, IRF-3, TBK1/NAK, and IKKε. The kit includes enough primary antibody to perform two western blot experiments for each primary antibody.

Background: Antiviral innate immunity depends on the combination of parallel pathways triggered by virus detecting proteins in the Toll-like receptor (TLR) family and RNA helicases, such as Rig-I (retinoic acid-inducible gene I) and MDA-5 (melanoma differentiation-associated antigen 5), which promote the transcription of type I interferons (IFN) and antiviral enzymes (1-3). TLRs and helicase proteins contain sites that recognize the molecular patterns of different virus types, including DNA, single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), and glycoproteins. These antiviral proteins are found in different cell compartments; TLRs (i.e. TLR3, TLR7, TLR8, and TLR9) are expressed on endosomal membranes and helicases are localized to the cytoplasm. Rig-I expression is induced by retinoic acid, LPS, IFN, and viral infection (4,5). Both Rig-I and MDA-5 share a DExD/H-box helicase domain that detects viral dsRNA and two amino-terminal caspase recruitment domains (CARD) that are required for triggering downstream signaling (4-7). Rig-I binds both dsRNA and viral ssRNA that contains a 5'-triphosphate end not seen in host RNA (8,9). Though structurally related, Rig-I and MDA-5 detect a distinct set of viruses (10,11). The CARD domain of the helicases, which is sufficient to generate signaling and IFN production, is recruited to the CARD domain of the MAVS/VISA/Cardif/IPS-1 mitochondrial protein, which triggers activation of NF-κB, TBK1/IKKε, and IRF-3/IRF-7

(12-15).

Specificity/Sensitivity: MDA-5 (D74E4) Rabbit mAb, Rig-I (D14G6) Rabbit mAb, MAVS Antibody, IRF-3 (D6I4C) XP® Rabbit mAb, TBK1/NAK (D1B4) Rabbit mAb, and IKKε (D20G4) Rabbit mAb detect endogenous levels of respective total proteins and do not cross-react with other proteins. Bands detected at 52 and 75 kDa by MAVS Antibody correlate with those described by Seth et al. (2005). Phospho-TBK1/NAK (Ser172) (D52C2) XP® Rabbit mAb detects endogenous levels of TBK1/NAK only when phosphorylated at Ser172. This antibody may cross-react with phospho-IKKε. Phospho-IRF-3 (Ser396) (4D4G) Rabbit mAb detects endogenous levels of IRF-3 only when phosphorylated at Ser396. Phospho-IKKε (Ser172) (D1B7) Rabbit mAb recognizes endogenous levels of IKKε protein only when phosphorylated at Ser172. This antibody may cross-react with phospho-TBK1/NAK.

Source/Purification: Polyclonal antibodies are produced by immunizing animals with a synthetic peptide corresponding to residues at the carboxy terminus of human MAVS protein. Polyclonal antibodies are purified by protein A and peptide affinity chromatography. Monoclonal antibodies are produced by immunizing animals with a synthetic peptide corresponding to residues surrounding Arg470 of human MDA-5 protein, Lys652 of human Rig-I protein, Ser645 of human TBK1/NAK protein, Val345 of human IKKε protein, or recombinant human IRF-3 protein. Activation state monoclonal antibodies are produced by immunizing animals with a synthetic phosphopeptide corresponding to residues surrounding Ser172 of human TBK1/NAK protein, Ser396 of

Storage: Supplied in 10 mM sodium HEPES (pH 7.5), 150 mM NaCl, 100 µg/ml BSA, 50% glycerol and less than 0.02% sodium azide. Store at -20°C. Do not aliquot the antibodies.

Recommended Antibody Dilutions:

Western blotting 1:1000

Please visit www.cellsignal.com for validation data and a complete listing of recommended companion products.

U.S. Patent No. 5,675,063

human IRF-3 protein, or Ser172 of human IKKε protein.

Background References:

- (1) Yoneyama, M. and Fujita, T. (2007) *J Biol Chem* 282, 15315-8.
- (2) Meylan, E. and Tschopp, J. (2006) *Mol Cell* 22, 561-9.
- (3) Thompson, A.J. and Locarnini, S.A. (2007) *Immunol Cell Biol* 85, 435-45.
- (4) Imaizumi, T. et al. (2002) *Biochem Biophys Res Commun* 292, 274-9.
- (5) Zhang, X. et al. (2000) *Microb Pathog* 28, 267-78.
- (6) Yoneyama, M. et al. (2005) *J Immunol* 175, 2851-8.
- (7) Yoneyama, M. et al. (2004) *Nat Immunol* 5, 730-7.
- (8) Hornung, V. et al. (2006) *Science* 314, 994-7.
- (9) Pichlmair, A. et al. (2006) *Science* 314, 997-1001.
- (10) Kato, H. et al. (2006) *Nature* 441, 101-5.
- (11) Childs, K. et al. (2007) *Virology* 359, 190-200.
- (12) Meylan, E. et al. (2005) *Nature* 437, 1167-72.
- (13) Xu, L.G. et al. (2005) *Mol Cell* 19, 727-40.
- (14) Kawai, T. et al. (2005) *Nat Immunol* 6, 981-8.
- (15) Seth, R.B. et al. (2005) *Cell* 122, 669-82.

Western Immunoblotting Protocol

For western blots, incubate membrane with diluted primary antibody in either 5% w/v BSA or nonfat dry milk, 1X TBS, 0.1% Tween® 20 at 4°C with gentle shaking, overnight.

NOTE: Please refer to primary antibody datasheet or product webpage for recommended primary antibody dilution buffer and recommended antibody dilution.

A. Solutions and Reagents

NOTE: Prepare solutions with reverse osmosis deionized (RODI) or equivalent grade water.

- 20X Phosphate Buffered Saline (PBS):** (#9808) To prepare 1 L 1X PBS: add 50 ml 20X PBS to 950 ml dH₂O, mix.
- 10X Tris Buffered Saline (TBS):** (#12498) To prepare 1 L 1X TBS: add 100 ml 10X to 900 ml dH₂O, mix.
- 1X SDS Sample Buffer:** Blue Loading Pack (#7722) or Red Loading Pack (#7723)
Prepare fresh 3X reducing loading buffer by adding 1/10 volume 30X DTT to 1 volume of 3X SDS loading buffer. Dilute to 1X with dH₂O.
- 10X Tris-Glycine SDS Running Buffer:** (#4050) To prepare 1 L 1X running buffer: add 100 ml 10X running buffer to 900 ml dH₂O, mix.
- 10X Tris-Glycine Transfer Buffer:** (#12539) To prepare 1 L 1X transfer buffer: add 100 ml 10X transfer buffer to 200 ml methanol + 700 ml dH₂O, mix.
- 10X Tris Buffered Saline with Tween® 20 (TBST):** (#9997) To prepare 1 L 1X TBST: add 100 ml 10X TBST to 900 ml dH₂O, mix.
- Nonfat Dry Milk:** (#9999)
- Blocking Buffer:** 1X TBST with 5% w/v nonfat dry milk; for 150 ml, add 7.5 g nonfat dry milk to 150 ml 1X TBST and mix well.
- Wash Buffer:** (#9997) 1X TBST
- Bovine Serum Albumin (BSA):** (#9998)
- Primary Antibody Dilution Buffer:** 1X TBST with 5% BSA or 5% nonfat dry milk as indicated on primary antibody datasheet; for 20 ml, add 1.0 g BSA or nonfat dry milk to 20 ml 1X TBST and mix well.
- Biotinylated Protein Ladder Detection Pack:** (#7727)
- Prestained Protein Marker, Broad Range (Premixed Format):** (#7720)
- Blotting Membrane and Paper:** (#12369) This protocol has been optimized for nitrocellulose membranes. Pore size 0.2 µm is generally recommended.
- Secondary Antibody Conjugated to HRP:** anti-rabbit (#7074); anti-mouse (#7076)
- Detection Reagent:** LumiGLO® chemiluminescent reagent and peroxide (#7003) or SignalFire™ ECL Reagent (#6883)

B. Protein Blotting

A general protocol for sample preparation.

- Treat cells by adding fresh media containing regulator for desired time.
- Aspirate media from cultures; wash cells with 1X PBS; aspirate.
- Lyse cells by adding 1X SDS sample buffer (100 µl per well of 6-well plate or 500 µl for a 10 cm diameter plate). Immediately scrape the cells off the plate and transfer the extract to a microcentrifuge tube. Keep on ice.
- Sonicate for 10–15 sec to complete cell lysis and shear DNA (to reduce sample viscosity).
- Heat a 20 µl sample to 95–100°C for 5 min; cool on ice.
- Microcentrifuge for 5 min.
- Load 20 µl onto SDS-PAGE gel (10 cm x 10 cm). **NOTE:** Loading of prestained molecular weight markers (#7720, 10 µl/lane) to verify electrotransfer and biotinylated protein ladder (#7727, 10 µl/lane) to determine molecular weights are recommended.
- Electrotransfer to nitrocellulose membrane (#12369).

C. Membrane Blocking and Antibody Incubations

NOTE: Volumes are for 10 cm x 10 cm (100 cm²) of membrane; for different sized membranes, adjust volumes accordingly.

I. Membrane Blocking

- (Optional) After transfer, wash nitrocellulose membrane with 25 ml TBS for 5 min at room temperature.
- Incubate membrane in 25 ml of blocking buffer for 1 hr at room temperature.
- Wash three times for 5 min each with 15 ml of TBST.

II. Primary Antibody Incubation

- Incubate membrane and primary antibody (at the appropriate dilution and diluent as recommended in the product datasheet) in 10 ml primary antibody dilution buffer with gentle agitation overnight at 4°C.
- Wash three times for 5 min each with 15 ml of TBST.
- Incubate membrane with the species appropriate HRP-conjugated secondary antibody (#7074 or #7076 at 1:2000) and anti-biotin, HRP-linked Antibody (#7075 at 1:1000–1:3000) to detect biotinylated protein markers in 10 ml of blocking buffer with gentle agitation for 1 hr at room temperature.
- Wash three times for 5 min each with 15 ml of TBST.
- Proceed with detection (Section D).

D. Detection of Proteins

- Incubate membrane with 10 ml LumiGLO® (0.5 ml 20X LumiGLO® #7003, 0.5 ml 20X peroxide, and 9.0 ml purified water) or 10 ml SignalFire™ #6883 (5 ml Reagent A, 5 ml Reagent B) with gentle agitation for 1 min at room temperature.
- Drain membrane of excess developing solution (do not let dry), wrap in plastic wrap and expose to x-ray film. An initial 10 sec exposure should indicate the proper exposure time. **NOTE:** Due to the kinetics of the detection reaction, signal is most intense immediately following incubation and declines over the following 2 hr.

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