

PathScan[®] Phospho-HER2/ErbB2 (Tyr1221/1222) Sandwich ELISA Antibody Pair



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Species Cross Reactivity:

UniProt ID: Entrez-Gene Id: #P04626 #2064

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Description

CST's PathScan® Phospho-HER2/ErbB2 (Tyr1221/1222) Sandwich ELISA Antibody Pair is offered as an economical alternative to our PathScan® Phospho-HER2/ErbB2 (Tyr1221/1222) Sandwich ELISA Kit #7148. Capture and Detection antibodies (100X stocks) and HRP-conjugated secondary antibody (1000X stock) are supplied. Sufficient reagents are supplied for 4 x 96 well ELISAs. The HER2/ErbB2 Capture Antibody is coated on a 96 well microplate in PBS overnight. After blocking, cell lysates are added followed by a Phospho-HER2/ErbB2 (Tyr1221/1222) Detection Antibody and anti-rabbit IgG, HRP conjugated antibody. HRP substrate, TMB, is added for color development. The magnitude of the absorbance for this developed color is proportional to the quantity of Phospho-HER2/ErbB2 (Tyr1221/1222) protein.

Antibodies in kit are custom formulations specific to kit.

Background

The ErbB2 (HER2) proto-oncogene encodes a 185 kDa transmembrane, receptor-like glycoprotein with intrinsic tyrosine kinase activity (1). While ErbB2 lacks an identified ligand, ErbB2 kinase activity can be activated in the absence of a ligand when overexpressed and through heteromeric associations with other ErbB family members (2). Amplification of the *ErbB2* gene and overexpression of its product are detected in almost 40% of human breast cancers (3). Binding of the c-Cbl ubiquitin ligase to ErbB2 at Tyr1112 leads to ErbB2 poly-ubiquitination and enhances degradation of this kinase (4). ErbB2 is a key therapeutic target in the treatment of breast cancer and other carcinomas and targeting the regulation of ErbB2 degradation by the c-Cbl-regulated proteolytic pathway is one potential therapeutic strategy. Phosphorylation of the kinase domain residue Tyr877 of ErbB2 (homologous to Tyr416 of pp60c-Src) may be involved in regulating ErbB2 biological activity. The major autophosphorylation sites in ErbB2 are Tyr1248 and Tyr1221/1222; phosphorylation of these sites couples ErbB2 to the Ras-Raf-MAP kinase signal transduction pathway (1,5).

Background References

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- 3. Dittadi, R. and Gion, M. (2000) J Natl Cancer Inst 92, 1443-4.
- 4. Klapper, L.N. et al. (2000) Cancer Res 60, 3384-8.
- 5. Kwon, Y.K. et al. (1997) J Neurosci 17, 8293-9.

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ELISA Antibody Pair

A. Solutions and Reagents

NOTE: Prepare solutions with reverse osmosis deionized (RODI) or equivalent grade water.

- 20X Phosphate Buffered Saline (PBS): (#9808) To prepare 1 L 1X PBS: add 50 ml 20X PBS to 950 ml dH₂O, mix.
- 2. **Wash Buffer**: 1X PBS/0.05% Tween[®] 20, (20X PBST #9809).
- 3. Blocking Buffer: 1X PBS/0.05% Tween® 20, 1% BSA.
- 4. **1X Cell Lysis Buffer**: 10X Cell Lysis Buffer (#9803): To prepare 10 ml of 1X Cell Lysis Buffer, add 1 ml of 10X Cell Lysis Buffer to 9 ml of dH₂O, mix. Buffer can be stored at 4°C for short-term use (1–2 weeks).

Recommended: Add 1 mM phenylmethylsulfonyl fluoride (PMSF) (#8553) immediately before use.

- 5. Bovine Serum Albumin (BSA): (#9998).
- 6. **TMB Substrate**: (#7004).
- 7. **STOP Solution**: (#7002)

NOTE: Reagents should be made fresh daily.

B. Preparing Cell Lysates

For adherent cells

- Aspirate media when the culture reaches 80-90% confluence. Treat cells by adding fresh media containing regulator for desired time.
- 2. Remove media and rinse cells once with ice-cold 1X PBS.
- 3. Remove PBS and add 0.5 ml ice-cold 1X Cell Lysis Buffer plus 1 mM PMSF to each plate (10 cm diameter) and incubate the plate on ice for 5 min.
- 4. Scrape cells off the plate and transfer to an appropriate tube. Keep on ice.
- 5. Sonicate lysates on ice.
- 6. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

For suspension cells

- 1. Remove media by low speed centrifugation (\sim 1,200 rpm) when the culture reaches 0.5–1.0 x 10⁶ viable cells/ml.
- Treat cells by adding fresh media containing regulator for desired time.

 2. Collect cells by low speed centrifugation (~1,200 rpm) and wash once with 5–10 ml ice-cold 1X PBS.
- 3. Cells harvested from 50 ml of growth media can be lysed in 2.0 ml of 1X cell lysis buffer plus 1 mM PMSF.
- 4. Sonicate lysates on ice.
- 5. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

C. Coating Procedure

- 1. Rinse microplate with 200 μ l of dH $_2$ O, discard liquid. Blot on paper towel to make sure wells are dry.
- 2. Dilute capture antibody 1:100 in $1\bar{X}$ PBS. For a single 96 well plate, add 100 μ l of capture antibody stock to 9.9 ml 1X PBS. Mix well and add 100 μ l/well. Cover plate and incubate overnight at 4°C (17–20 hr).
- 3. After overnight coating, gently uncover plate and wash wells:
 - 1. Discard plate contents into a receptacle.
 - 2. Wash four times with wash buffer, 200 µl each time per well. For each wash, strike plates on fresh paper towels hard enough to remove the residual solution in each well, but do not allow wells to completely dry at any time.
 - 3. Clean the underside of all wells with a lint-free tissue.
- 4. Block plates. Add 150 μ l of blocking buffer/well, cover plate, and incubate at 37°C for 2 hr.
- 5. After blocking, wash plate (Section C, Step 3). Plate is ready to use.

D. Test Procedure

1. Lysates can be used undiluted or diluted in blocking buffer. $100~\mu l$ of lysate is added per well. Cover plate and incubate at $37^{\circ}C$ for 2 hr.

- 2. Wash plate (Section C, Step 3).
- 3. Dilute detection antibody 1:100 in blocking buffer. For a single 96 well plate, add 100 µl of detection antibody Stock to 9.9 ml of blocking buffer. Mix well and add 100 µl/well. Cover plate and incubate at 37°C for 1 hr.
- 4. Wash plate (Section C, Step 3).
- 5. Secondary antibody, either streptavidin anti-mouse or anti-rabbit-HRP, is diluted 1:1000 in blocking buffer. For a single 96 well plate, add 10 μ l of secondary antibody stock to 9.99 ml of blocking buffer. Mix well and add 100 μ l/well. Cover and incubate at 37°C for 30 min.
- 6. Wash plate (Section C, Step 3).
 7. Add 100 μl of TMB substrate per well. Cover and incubate at 37°C for 10 min.
- 8. Add 100 µl of STOP solution per well. Shake gently for a few seconds.
- 9. Read plate on a microplate reader at absorbance 450 nm.
 - 1. Visual Determination: Read within 30 min after adding STOP solution.
 - 2. Spectrophotometric Determination: Wipe underside of wells with a lint-free tissue. Read absorbance at 450 nm within 30 min after adding STOP solution.

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