## IKKE Kinase





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This product is for in vitro research use only and is not intended for use in humans or animals.

**Description:** Purified recombinant full-length human IKK $\epsilon$  kinase, supplied as a GST fusion protein.

**Background:** The NFκB/Rel transcription factors are present in the cytosol in an inactive state, complexed with the inhibitory IxB proteins (1-3). Most agents that activate NFkB do so through a common pathway based on phosphorylation-induced, proteasome-mediated degradation of  $l\kappa B$  (3-7). The key regulatory step in this pathway involves activation of a high molecular weight IκB kinase (IKK) complex, whose catalysis is generally carried out by three tightly associated IKK subunits. IKK $\alpha$  and IKK $\beta$  serve as the catalytic subunits of the kinase. IKKy serves as the regulatory subunit (8-9). Activation of IKK depends on phosphorylation; serines 177 and 181 in the activation loop of IKKB (176 and 180 in IKK $\alpha$ ) are the specific sites whose phosphorylation causes conformational changes resulting in kinase activation (10-13).

Recently, two homologs of IKK $\alpha$  and IKK $\beta$  have been described, called IKK $\epsilon$  (also known as IKK-i) and TBK-1 (also known as T2K or NAK), and activation of either of these kinases results in NF $\kappa$ B activation. The kinase domain of IKK $\epsilon$  is located in its amino-terminus, which shares 30% sequence homology with both IKK $\alpha$  and IKK $\beta$ . IKK $\epsilon$  is expressed predominantly in immune cells, and may play a special role in the immune response (14–18).

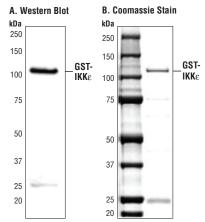


Figure 1. The purity of the GST-IKKe fusion protein was analyzed using SDS/PAGE followed by anti-GST Western blot (A) or Coomassie stain (B).

Source/Purification: The GST-Kinase fusion protein was produced using a baculovirus expression system with a construct expressing full length human IKKε (Met1-Val716) (GenBank Accession No. NM\_014002) with an amino-terminal GST tag. The protein was purified by one-step affinity chromatography using glutathione-agarose.

Quality Control: The theoretical molecular weight of the GST-IKKɛ fusion protein is 110 kDa. The purified kinase was quality controlled for purity using SDS-PAGE followed by Coomassie stain and Western blot [Fig.1]. IKKɛ kinase activity was determined using a radiometric assay [Fig.2]. A kinase dose dependency assay was performed to measure IKKɛ activity using HTScan™ IKKɛ Kinase Assay Kit #7556 [Fig.3].

### **Background References:**

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- (7) Chen, Z.J. et al. (1996) Cell 84, 853-862.
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- (11) Mercurio, F. et al. (1997) Science 278, 860-866.
- (12) Johnson, L.N. et al. (1996) *Cell* 85, 149–158.
- (13) Delhase, M. et al. (1999) Science 284, 309-313.
- (14) Shimada, T. et al. (1999) *Int. Immunol.* 11, 1357–1362.
- (15) Peters, R.T. et al. (2000) Mol. Cell. 5, 513-522.
- (16) Tojima, Y. et al. (2000) *Nature* 404, 778–782.
- (17) Bonnard, M. et al. (2000) EMBO J. 19, 4976-4985.
- (18) Peters, R.T. and Maniats, T. (2001) *Biochim. Biophys. Acta.* 1471, M57–62.

**Storage:** Enzyme is supplied in 50 mM Tris-HCl, pH 8.0; 100 mM NaCl, 5 mM DTT, 15 mM reduced glutathione, 20% glycerol. Store at -80° C.

Keep on ice during use.

Avoid repeated freeze-thaw cycles.

#### **Companion Products:**

HTScan™ IKKε Kinase Assay Kit #7556

PAK1 (Ser144)/ PAK2 (Ser141) Biotinylated Peptide #1134

Phospho-(Ser/Thr) Phe Antibody #9631

Kinase Buffer (10X) #9802

ATP (10 mM) #9804

Staurosporine #9953

Serine/Threonine Kinase Substrate Screening Kit #7400

Cell Signaling Technology offers a full line of protein kinases, substrates, and antibody detection reagents for high throughput screening. Please direct all inquiries to: <a href="mailto:drugdiscovery@cellsignal.com">drugdiscovery@cellsignal.com</a>



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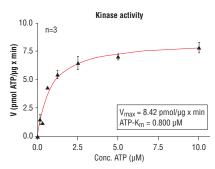


Figure 2. IKKε kinase activity was measured in a radiometric assay using the following reaction conditions: 60 mM HEPES-NaOH, pH 7.5, 3 mM MgCl<sub>2</sub>, 3 mM MnCl<sub>2</sub>, 3 μM Na-orthovanadate, 1.2 mM DTT, 1 μM ATP, 2.5 μg/50 μl PEG20,000, Substrate: Casein, 10 μg/50 μl and Recombinant IKKε: 100 ng/50 μl.

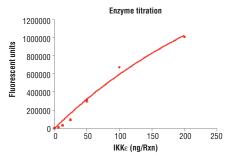


Figure 3. Dose dependence curve of IKKe kinase activity: DELFIA® data generated using Phospho-(Ser/Thr) Phe Antibody #9631 to detect phosphorylation of substrate peptide #1134 by IKKe kinase. In a 50 µl reaction, increasing amounts of IKKe and 1.5 µM substrate peptide were used per reaction at room temperature for 30 minutes. (DELFIA® is a registered trademark of PerkinElmer, Inc.)

# Protocol for IKKε Kinase Assay

#### **Kinase**

Note: Lot-specific information for this kinase is provided on the enzyme vial. Optimal assay incubation times and enzyme concentrations must be determined empirically for each lot of kinase under specified conditions.

## Additional Solutions and Reagents (Not included)

■ Wash Buffer: 1X PBS, 0.05% Tween-20 (PBS/T)

■ Bovine Serum Albumin (BSA)

■ Stop Buffer: 50 mM EDTA pH 8

■ Phospho-(Ser/Thr) Phe Antibody #9631

■ Kinase Buffer (10X) #9802

■ ATP (10 mM) #9804

■ PAK1 (Ser144)/ PAK2 (Ser141) Biotinylated Peptide #1134

■ DELFIA® Europium-labeled Anti-rabbit antibody (PerkinElmer Life Sciences #AD0105)

■ DELFIA® Enhancement Solution (PerkinElmer Life Sciences #1244-105)

■ DELFIA® Streptavidin coated, 96-well, yellow plate (PerkinElmer Life Sciences AAAND-0005)

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## Suggested Protocol For 100 Assays

- 1. Add 100  $\mu$ l 10 mM ATP to 1.25 ml 6  $\mu$ M substrate peptide. Dilute the mixture with dH<sub>2</sub>0 to 2.5 ml to make 2X ATP/substrate cocktail ([ATP]=400  $\mu$ M, [substrate]=3  $\mu$ m).
- 2. Transfer enzyme from  $-80^{\circ}\text{C}$  to ice. Allow enzyme to thaw on ice.
- Microcentrifuge briefly at 4°C to bring liquid to the bottom of the vial. Return immediately to ice.
- 4. Add 1 ml 10X kinase buffer [250 mM Tris-HCl pH 7.5, 100 mM MgCl $_2$ , 1 mM Na $_3$ VO $_4$ , 50 mM  $\beta$ -glycerophosphate, 20 mM dithiothreitol (DTT)] to 1.5 ml dH $_2$ 0 to make 2.5 ml 4X reaction buffer.
- Dilute enzyme in 1.25 ml of 4X reaction buffer to make 4X reaction cocktail ([enzyme]=4.0 ng/µl in 4X reaction cocktail).
- Add 12.5 μl of the 4X reaction cocktail to 12.5 μl/well of prediluted compound of interest (usually around 10 μM) and incubate for 5 minutes at room temperature.
- Add 25 µl of 2X ATP/substrate cocktail to 25 µl/well preincubated reaction cocktail/compound.

#### Final Assay Conditions for a 50 µl Reaction

25 mM Tris-HCI (pH7.5)

10 mM MgCl<sub>2</sub>

5 mM β-glycerophosphate

0.1 mM Na<sub>3</sub>VO<sub>4</sub>

2 mM DTT

200 μM ATP

1.5 µM peptide

50 ng IKKε Kinase

- 8. Incubate reaction plate at room temperature for 30 minutes.
- 9. Add 50  $\mu\text{I/well}$  Stop Buffer (50 mM EDTA, pH 8) to stop the reaction.
- 10. Transfer 25  $\mu$ I of each reaction to a 96-well streptavidin-coated plate containing 75  $\mu$ I dH<sub>2</sub>O/well and incubate at room temperature for 60 minutes.
- 11. \*Wash three times with 200 µl/well PBS/T.
- Dilute primary antibody in PBS/T with 1% BSA. Add 100 μl/well primary antibody.

**Please note:** This protocol was validated using a PAK1 (Ser144)/ PAK2 (Ser141) Biotinylated Peptide and Phospho-(Ser/Thr) Phe Antibody diluted 1:1000 (see additional reagents). Primary antibody chosen should be specific to the substrate used.

- 13. Incubate at 37°C for 120 minutes.
- 14. \*Wash three times with 200 ul/well PBS/T.
- 15. Dilute Europium labeled secondary antibody 1:1000 in PBS/T with 1% BSA. Add 100  $\mu$ I/well diluted antibody.
- 16. Incubate at room temperature for 30 minutes.
- 17. \*Wash five times with 200  $\mu$ I/well PBS/T.
- 18. Add 100 µl/well DELFIA® Enhancement Solution.
- 19. Incubate at room temperature for 5 minutes.
- Detect 615 nm fluorescence emission with appropriate Time-Resolved Plate Reader.

\*IMPORTANT: Use of an automated microplate washer as well as centrifugation of plates when appropriate, greatly improves reproducibility.