

IKK β Kinase

☑ 5 μ g



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This product is for *in vitro* research use only and is not intended for use in humans or animals.

Description: Purified recombinant full length human IKK β kinase, supplied as a GST fusion protein.

Background: The NF κ B/Rel transcription factors are present in the cytosol in an inactive state, complexed with the inhibitory I κ B proteins (1-3). Most agents that activate NF κ B do so through a common pathway based on phosphorylation-induced, proteasome-mediated degradation of I κ B (3-7). The key regulatory step in this pathway involves activation of a high molecular weight I κ B kinase (IKK) complex, whose catalysis is generally carried out by three tightly associated IKK subunits. IKK α and IKK β serve as the catalytic subunits of the kinase. IKK γ serves as the regulatory subunit (8-9). Activation of IKK depends on phosphorylation; serines 177 and 181 in the activation loop of IKK β (176 and 180 in IKK α) are the specific sites whose phosphorylation causes conformational changes resulting in kinase activation (10-13).

Source/Purification: The GST-Kinase fusion protein was produced using a baculovirus expression system from a construct containing a full length human IKK β cDNA kinase domain (Met1-Ser756) (GenBank accession No. AF029684) fragment amino-terminally fused to a GST-HIS $_6$ -Thrombin cleavage site. The protein was then purified by one-step affinity purification using glutathione-agarose.

Quality Control: The theoretical molecular weight of the GST-IKK β kinase fusion protein is 87 kDa (apparent molecular weight on SDS PAGE is 120 kDa). The purified kinase fusion protein was quality controlled for purity using SDS-PAGE Silver stain and Western blot [Fig.1]. IKK β kinase activity was determined using a radiometric assay [Fig.2]. A kinase dose dependency assay was performed to measure IKK β activity using HTScan™ IKK β Kinase Assay Kit #7549 [Fig.3].

Background References:

- (1) Baeuerle, P.A. et al. (1988) *Science* 242, 540-546.
- (2) Beg, A.A. et al. (1993) *Genes Dev.* 7, 2064-2070.
- (3) Finco, T.S. et al. (1994) *Proc. Natl. Acad. Sci. USA* 91, 11884-11888.
- (4) Brown, K. et al. (1995) *Science* 267, 1485-1488.
- (5) Brockman, J.A. et al. (1995) *Mol. Cell. Biol.* 15, 2809-2818.
- (6) Traenckner, E.B. et al. (1995) *EMBO J.* 14, 2876-2883.
- (7) Chen, Z.J. et al. (1996) *Cell* 84, 853-862.
- (8) Zandi, E. et al. (1997) *Cell* 91, 243-252.
- (9) Karin, M. et al. (1999) *Oncogene* 18, 6867-6874.
- (10) DiDonato, J.A. et al. (1997) *Nature* 388, 548-554.
- (11) Mercurio, F. et al. (1997) *Science* 278, 860-866.
- (12) Johnson, L.N. et al. (1996) *Cell* 85, 149-158.
- (13) Delhase, M. et al. (1999) *Science* 284, 309-313.

Storage: Enzyme is supplied in 50 mM Tris-HCl, pH 8.0; 100 mM NaCl, 5 mM DTT, 15 mM reduced glutathione, 20% glycerol. Store at -80°C.

Keep on ice during use.

Avoid repeated freeze-thaw cycles.

Companion Products:

HTScan™ IKK β Kinase Assay Kit #7549

I κ B- α (Ser32) Biotinylated Peptide #1146

Phospho-I κ B- α (Ser32/36) (5A5) Mouse mAb #9246

Kinase Buffer (10X) #9802

ATP (10 mM) #9804

Staurosporine #9953

Serine/Threonine Kinase Substrate Screening Kit #7400

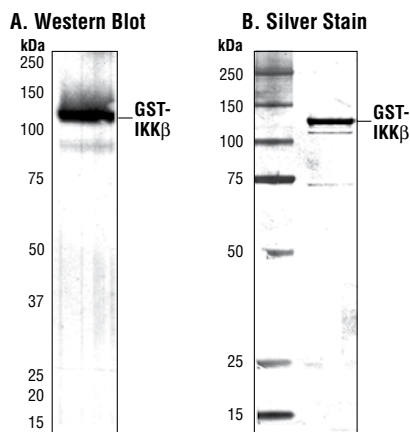


Figure 1. The purity of the GST-IKK β fusion protein was analyzed using SDS/PAGE followed by anti-IKK β Western blot (A) or Silver stain (B).

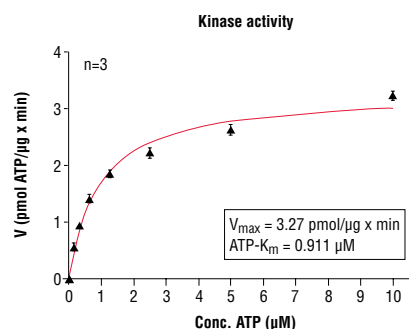


Figure 2. IKK β kinase activity was measured in a radiometric assay using the following reaction conditions: 60 mM HEPES-NaOH, pH 7.5, 3 mM MgCl₂, 3 mM MnCl₂, 3 μ M Na-orthovanadate, 1.2 mM DTT, ATP (variable), 2.5 μ g/50 μ l PEG20,000, Substrate: Rb CTF, 1.5 μ g/50 μ l, Recombinant IKK β : 50 ng/50 μ l.

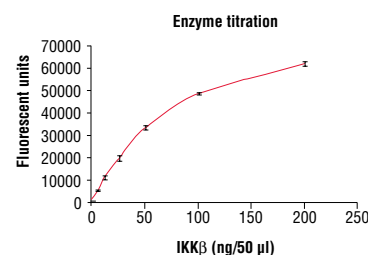


Figure 3. Dose dependence curve of IKK β kinase activity: DELFIA® data generated using Phospho-I κ B- α (Ser32/36) (5A5) Mouse mAb #9246 to detect phosphorylation of substrate peptide (#1146) by IKK β kinase. In a 50 μ l reaction, increasing amounts of IKK β and 1.5 μ M substrate peptide were used per reaction well at room temperature for 30 minutes. (DELFIA® is a registered trademark of PerkinElmer, Inc.)

Protocol for IKK β Kinase Assay

***IMPORTANT:** Use of an automated microplate washer as well as centrifugation of plates when appropriate, greatly improves reproducibility.

Kinase

Note: Lot-specific information for this kinase is provided on the enzyme vial. Optimal assay incubation times and enzyme concentrations must be determined empirically for each lot of kinase under specified conditions.

A Additional Solutions and Reagents (Not included)

1. **Wash Buffer:** 1X PBS, 0.05% Tween-20 (PBS/T)
2. Bovine Serum Albumin (BSA)
3. **Stop Buffer:** 50 mM EDTA pH 8
4. Phospho-I κ B- α (Ser32/36) (5A5) Mouse mAb #9246
5. Kinase Buffer (10X) #9802
6. ATP (10 mM) #9804
7. I κ B- α (Ser32) Biotinylated Peptide #1146
8. DELFIA® Europium-labeled Anti-mouse IgG antibody (PerkinElmer Life Sciences #AD0124)
9. DELFIA® Enhancement Solution (PerkinElmer Life Sciences #1244-105)
10. DELFIA® Streptavidin coated, 96-well, yellow plate (PerkinElmer Life Sciences AAAND-0005)

DELFIA® is a registered trademark of PerkinElmer Life Sciences

B Suggested Protocol for 100 Assays

1. Add 100 μ l 10 mM ATP to 1.25 ml 6 μ M substrate peptide. Dilute the mixture with dH₂O to 2.5 ml to make 2X ATP/substrate cocktail ([ATP]=400 μ M, [substrate] = 3 μ M).
2. Transfer enzyme from -80°C to ice. Allow enzyme to thaw on ice.
3. **Microcentrifuge briefly at 4°C to bring liquid to the bottom of the vial. Return immediately to ice.**
4. Add 1 ml 10X kinase buffer [250 mM Tris-HCl pH 7.5, 100 mM MgCl₂, 1 mM Na₃VO₄, 50 mM β -glycerophosphate, 20 mM dithiothreitol (DTT)] to 1.5 ml dH₂O to make 2.5 ml 4X reaction buffer.
5. Dilute enzyme in 1.25 ml of 4X reaction buffer to make 4X reaction cocktail ([enzyme]=4.0 ng/ μ l in 4X reaction cocktail).
6. Add 12.5 μ l of the 4X reaction cocktail to 12.5 μ l/well of prediluted compound of interest (usually around 10 μ M) and incubate for 5 minutes at room temperature.
7. Add 25 μ l of 2X ATP/substrate cocktail to 25 μ l/well preincubated reaction cocktail/compound.

Final Assay Conditions for a 50 μ l Reaction

25 mM Tris-HCl (pH 7.5)
10 mM MgCl₂
5 mM β -glycerophosphate
0.1 mM Na₃VO₄
2 mM DTT
200 μ M ATP
1.5 μ M peptide
50 ng IKK β Kinase

8. Incubate reaction plate at room temperature for 30 minutes.
9. Add 50 μ l/well Stop Buffer (50 mM EDTA, pH 8) to stop the reaction.
10. Transfer 25 μ l of each reaction to a 96-well streptavidin-coated plate containing 75 μ l dH₂O/well and incubate at room temperature for 60 minutes.
11. *Wash three times with 200 μ l/well PBS/T.
12. Dilute primary antibody in PBS/T with 1% BSA. Add 100 μ l/well primary antibody.
Please note: This protocol was validated using a I κ B- α (Ser32) Biotinylated Peptide and Phospho-I κ B- α (Ser32/36) (5A5) Mouse mAb diluted 1:1000 (see additional reagents). Primary antibody chosen should be specific to the substrate used.
13. Incubate at 37°C for 120 minutes.
14. *Wash three times with 200 μ l/well PBS/T.
15. Dilute Europium labeled secondary antibody 1:500 in PBS/T with 1% BSA. Add 100 μ l/well diluted antibody.
16. Incubate at room temperature for 30 minutes.
17. *Wash five times with 200 μ l/well PBS/T.
18. Add 100 μ l/well DELFIA® Enhancement Solution.
19. Incubate at room temperature for 5 minutes.
20. Detect 615 nm fluorescence emission with appropriate Time-Resolved Plate Reader.

Please contact Cell Signaling Technology for HTS-ready antibodies (PBS formulated and carrier-free), and detailed peptide substrate sequence information.
Email: drugdiscovery@cellsignal.com