

X-34



#74193

5 mg

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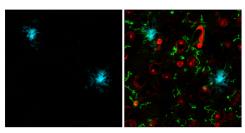
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Description: X-34 is a lipophilic and highly fluorescent marker used to examine pathological changes in Alzheimer's disease (AD). This cellular dye was developed as a derivative of Congo red to be more compatible with *in vivo* applications. By replacing the azo groups (nitrogen-to-nitrogen double bonds) in Congo red with carbon-to-carbon double bonds and having more lipophilic salicylic acid groups, X-34 has proven to be highly fluorescent and have increased membrane permeability. X-34 detects β-sheet structures and labels amyloid plaques, neurofibrillary tangles (NFTs), neuropil threads, and vascular amyloid in AD brains (1).

Molecular Formula: $C_{24}H_{18}O_6$ Molecular Weight: 402.4 g/mol

Purity: ≥ 90% **CAS:** 215294-98-7

Solubility: Soluble in DMSO at 2 mg/mL. **Excitation/Emission Max:** 367/497 nm



Confocal analysis of cortex from the 5XFAD mouse model of Alzheimer's disease, labeled with X-34 (left, cyan) and co-labeled with TMEM119 (E4B9S) Mouse mAb #98778 (right, green) and DRAQ5 #4084 (right, red). **Storage:** Store lyophilized at 4°C, desiccated. In lyophilized form, the product is stable for 12 months.

Please visit cellsignal.com for validation data and a complete listing of recommended companion products.

Directions for Use:

Supplied Reagents:

1. X-34: Create a 5 mM stock solution of X-34 (200X) using the X-34 diluent. Create a 25 µM staining solution by further diluting the 5 mM stock solution using the X-34 diluent.

Note: X-34 diluent (40% Et0H, pH 10) is the recommended solvent for staining. A slight precipitate may be present, but will not interfere with the product's performance.

Additional Reagants (Not Supplied)

- 1. X-34 diluent (40% EtOH, pH 10): Combine 20 mL of EtOH in 29.5 mL of dH $_2$ 0. Add 1 μ L of 5M NaOH dropwise until the solution reaches pH 10. Bring the final volume of X-34 diluent to 50 mL with dH $_2$ 0.
- 2. Differentiation buffer: 50 mM NaOH, 80% EtOH

Protocol:

- 1. Stain slides in 25 μ M X-34 staining solution for 30 min at room temperature in the dark.
- 2. Rinse slides three times in dH₂O.
- 3. Incubate for 2 min in the differentiation buffer.
- 4. Rinse slides three times in dH₂0.
- 5. If desired, proceed with immunostaining.
- 6. Mount and image.

Background References:

(1) Styren, S.D. et al. (2000) *J Histochem Cytochem* 48, 1223-32.

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