e at +4C	PathScan [®] RP Total c-Raf Sandwich ELIS Kit			LISA			
Stor				ο	rders:	877-616-CELL (2355) orders@cellsignal.com	
76	1 Kit (96 assays)			Su	.pport:	877-678-TECH (8324)	
87	Species Cross Reactivity : H M R Mk	UniProt ID: #P04049	Entrez-Gene Id: #5894	w	eb:	info@cellsignal.com cellsignal.com	
9#	esearch Lise Only, Not fo	or Use in Dia	anostic Procedures	3 Trask Lane Danv	rers Ma	issachusetts 01923 USA	

Product Includes Product # Quantity Color Storage Temp c-Raf Rabbit mAb Coated Microwells 83655 96 tests +4C Red (Lyophilized) c-Raf Mouse Detection mAb 25933 1 ea +4C **HRP** Diluent 13515 5.5 ml Red +4C **TMB** Substrate 7004 11 ml +4C **STOP Solution** 7002 11 ml +4C 2 ea +4C Sealing Tape 54503 ELISA Wash Buffer (20X) 9801 25 ml +4C Cell Lysis Buffer (10X) 9803 15 ml -20C

Kit contents scale proportionally with size, except sealing tape.

Example: The V1 kit contains 5X the listed quantities above, but will exclude the sealing tape.

The microwell plate is supplied as 12 8-well modules - Each module is designed to break apart for 8 tests.

Description	The rapid protocol (RP) PathScan [®] RP Total c-Raf Sandwich ELISA Kit is a solid phase sandwich enzyme- linked immunosorbent assay (ELISA) that detects endogenous levels of c-Raf protein in a reduced assay time of 1.5 hours. Incubation of cell lysates and detection antibody on the coated microwell plate forms a sandwich with c-Raf in a single step. The plate is then extensively washed and TMB reagent is added for signal development. The magnitude of absorbance for the developed color is proportional to the quantity of c-Raf. Learn more about all of your ELISA kit options here. *Antibodies in this kit are custom formulations specific to kit.
Specificity/Sensitivity	The PathScan [®] RP Total c-Raf Sandwich ELISA Kit detects endogenous levels of c-Raf protein. The kit sensitivity is shown in Figure 2. This kit detects proteins from the indicated species, as determined through in-house testing, but may also detect homologous proteins from other species.
Background	A-Raf, B-Raf, and c-Raf (Raf-1) are the main effectors recruited by GTP-bound Ras to activate the MEK- MAP kinase pathway (1). Activation of c-Raf is the best understood and involves phosphorylation at multiple activating sites, including Ser338, Tyr341, Thr491, Ser494, Ser497, and Ser499 (2). p21- activated kinase (PAK) has been shown to phosphorylate c-Raf at Ser338, and the Src family phosphorylates Tyr341 to induce c-Raf activity (3,4). Ser338 of c-Raf corresponds to similar sites in A-Raf (Ser299) and B-Raf (Ser445), although this site is constitutively phosphorylated in B-Raf (5). Inhibitory 14-3-3 binding sites on c-Raf (Ser259 and Ser621) can be phosphorylated by Akt and AMPK, respectively (6,7). While A-Raf, B-Raf, and c-Raf are similar in sequence and function, differential regulation has been observed (8). Of particular interest, B-Raf contains three consensus Akt phosphorylation sites (Ser364, Ser428, and Thr439) and lacks a site equivalent to Tyr341 of c-Raf (8,9). Research studies have shown that the B-Raf mutation V600E results in elevated kinase activity and is commonly found in malignant melanoma (10). Six residues of c-Raf (Ser29, Ser43, Ser289, Ser296, Ser301, and Ser642) become hyperphosphorylated in a manner consistent with c-Raf inactivation. The hyperphosphorylation of these six sites is dependent on downstream MEK signaling and renders c-Raf unresponsive to subsequent activation events (11).
Background References	1. Avruch, J. et al. (1994) <i>Trends Biochem Sci</i> 19, 279-83. 2. Chong, H. et al. (2001) <i>EMBO J</i> 20, 3716-27. 3. King, A.J. et al. (1998) <i>Nature</i> 396, 180-3. 4. Fabian, J.R. et al. (1993) <i>Mol Cell Biol</i> 13, 7170-9. 5. Mason, C.S. et al. (1999) <i>EMBO J</i> 18, 2137-48. 6. Zimmermann, S. and Moelling, K. (1999) <i>Science</i> 286, 1741-4. 7. Sprenkle, A.B. et al. (1997) <i>FEBS Lett</i> 403, 254-8. 8. Marais, R. et al. (1997) <i>J Biol Chem</i> 272, 4378-83.

	9. Guan, K.L. et al. (2000) <i>J Biol Chem</i> 275, 27354-9. 10. Davies, H. et al. (2002) <i>Nature</i> 417, 949-54. 11. Dougherty, M.K. et al. (2005) <i>Mol Cell</i> 17, 215-24.				
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#68776 PathScan[®] RP Total c-Raf Sandwich ELISA Kit



PathScan[®] Sandwich ELISA Protocol (Rapid Protocol)

NOTE: This protocol is for PathScan[®] kits that use an HRP directly conjugated to the detection antibody **(Rapid Protocol)**, rather than a 2-step method where the detection antibody and a secondary-HRP are added sequentially.

A. Solutions and Reagents

NOTE: Prepare solutions with deionized/purified water or equivalent.

- 1. **Microwell strips:** Bring all to room temperature before opening bag/use. Unused microwell strips should be returned to the original re-sealable bag containing the desiccant pack and stored at 4°C.
- 2. Detection Antibody: Reconstitute lyophilized Detection Antibody (red colored cake) with 1 mL of HRP Diluent (red solution) to yield a concentrated stock solution. Incubate at room temperature for 5 min with occasional gentle mixing to fully reconstitute. To make the final working solution, add the full 1 mL of reconstituted Detection Antibody to 4.5 mL of HRP Diluent in a clean tube and gently mix. For best results, use immediately following antibody reconstitution. Unused reconstituted Detection Antibody may be stored for up to 4 weeks at 4°C, although there may be some loss of signal compared to freshly reconstituted antibody.
- 3. HRP Diluent: Red colored diluent for reconstitution and dilution of the Detection Antibody that is linked to HRP.
- 4. **1X ELISA Wash Buffer:** Prepare by diluting ELISA Wash Buffer (20X) (included in each kit) to 1X with deionized water.
- 5. 1X Cell Lysis Buffer: Prepare by diluting 10X Cell Lysis Buffer #9803 to 1X with deionized water. This buffer can be stored at 4°C for short-term use (1–2 weeks). Recommended: When using to prepare cell lysates, add Protease/Phosphatase Inhibitor Cocktail (#5872, not supplied) and 1 mM phenylmethyl- sulfonyl fluoride (PMSF, #8553, not supplied) immediately before use.
- 6. TMB Substrate (#7004): Bring to room temperature before use.
- 7. STOP Solution (#7002): Bring to room temperature before use.

B. Preparing Cell Lysates

For adherent cells

- 1. Aspirate media when the culture reaches 80–90% confluence. Treat cells by adding fresh media containing regulator for desired time.
- 2. Remove media and rinse cells once with ice-cold 1X PBS.
- 3. Remove PBS and add 0.5 mL ice-cold 1X Cell Lysis Buffer including 1 mM PMSF and Protease/Phosphatase Inhibitor Cocktail to each plate (10 cm diameter) and incubate the plate on ice for 5 min.
- 4. Scrape cells off the plate and transfer to an appropriate tube. Keep on ice.
- 5. Sonicate lysates on ice.
- Microcentrifuge for 10 min (14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

For suspension cells

- 1. Remove media by low speed centrifugation (\sim 1200 rpm) when the culture reaches 0.5–1.0 x 10⁶ viable cells/mL. Treat cells by adding fresh media containing regulator for desired time.
- 2. Collect cells by low speed centrifugation (~1200 rpm) and wash once with 5-10 mL ice-cold 1X PBS.
- 3. Cells harvested from 50 mL of growth media can be lysed in 2.0 mL of 1X Cell Lysis Buffer including 1 mM PMSF and Protease/Phosphatase Inhibitor Cocktail.
- 4. Sonicate lysates on ice.
- 5. Microcentrifuge for 10 min (14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

C. Test Procedure

NOTE: Equilibrate all materials and prepared reagents to room temperature prior to running the assay.

- 1. Prepare all reagents as indicated above (Section A).
- 2. Samples should be undiluted or diluted with 1X Cell Lysis Buffer to a 2X protein concentration in order to achieve a final 1X protein concentration upon addition of the Detection Antibody. Individual datasheets for each kit provide a sensitivity curve that serves as a reference for selection of an appropriate starting lysate concentration. The sensitivity curve shows typical results across a range of lysate concentration points.
- 3. Add 50 μ L of each sample to the appropriate wells.
- 4. Add 50 µL of the Detection Antibody to each well.
- 5. Seal the plate and incubate for 1 hour at room temperature on a plate shaker set to 400 rpm (moderate agitation).

- 6. Gently remove the tape and wash wells:
 - 1. Discard plate contents into a receptacle.
 - 2. Wash 4 times with 1X Wash Buffer, 200 μ L each time for each well.
 - 3. For each wash, strike plates on fresh towels hard enough to remove the residual solution in each well, but do not allow wells to completely dry at any time.
 - 4. Clean the underside of all wells with a lint-free tissue.
- 7. Add 100 μ L of TMB Substrate to each well. Seal with tape and incubate the plate in the dark for 15 min at room temperature on a plate shaker (400 rpm, moderate agitation) or alternatively for 10 min at 37°C without shaking.
- 8. Add 100 μL of STOP Solution to each well. Shake gently for a few seconds.
 - **NOTE:** Initial color of positive reaction is blue, which changes to yellow upon addition of STOP Solution.
- 9. Read results:
 - 1. Visual Determination: Read within 30 min after adding STOP Solution.
 - 2. **Spectrophotometric Determination:** Wipe underside of wells with a lint-free tissue. Read absorbance at 450 nm within 30 min after adding STOP Solution.

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