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# FastScan<sup>™</sup> Total p44/42 MAPK (Erk1/2) ELISA Kit



1 Kit (96 assays)



**Support:** +1-978-867-2388 (U.S.) www.cellsignal.com/support

Orders: 877-616-2355 (U.S.) orders@cellsignal.com

**Entrez-Gene ID** #5595, 5594 **UniProt ID** #P27361, P28482

## New 10/18

# For Research Use Only. Not For Use In Diagnostic Procedures.

Species Cross-Reactivity: H, M, R, Mk

Description: The FastScan™ p44/42 MAPK (Erk1/2) ELISA Kit is a sandwich enzyme-linked immunosorbent assay (ELISA) that detects endogenous levels of p44/42 MAPK (Erk1/2). To perform the assay, sample is incubated with a capture antibody conjugated with a proprietary tag and a second detection antibody linked to HRP, forming a sandwich with p44/42 MAPK (Erk1/2) in solution. This entire complex is immobilized to the plate via an anti-tag antibody. The wells are then washed to remove unbound material. TMB is then added. The magnitude of observed signal is proportional to the quantity of p44/42 MAPK (Erk1/2).

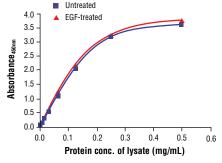
\*Antibodies in kit are custom formulations specific to kit.

Specificity/Sensitivity: The FastScan™ p44/42 MAPK (Erk1/2) ELISA Kit detects endogenous levels of p44/42 MAPK (Erk1/2) as shown in Figure 1. This kit detects proteins from the indicated species, as determined through in-house testing, but may also detect homologous proteins from other species.

**Background:** Mitogen-activated protein kinases (MAPKs) are a widely conserved family of serine/threonine protein kinases involved in many cellular programs, such as cell proliferation, differentiation, motility, and death. The p44/42 MAPK (Erk1/2) signaling pathway can be activated in response to a diverse range of extracellular stimuli including mitogens, growth factors, and cytokines (1-3), and research investigators consider it an important target in the diagnosis and treatment of cancer (4). Upon stimulation, a sequential three-part protein kinase cascade is initiated, consisting of a MAP kinase kinase kinase (MAPKKK or MAP3K), a MAP kinase kinase (MAPKK or MAP2K), and a MAP kinase (MAPK). Multiple p44/42 MAP3Ks have been identified, including members of the Raf family, as well as Mos and Tpl2/COT. MEK1 and MEK2 are the primary MAPKKs in this pathway (5,6). MEK1 and MEK2 activate p44 and p42 through phosphorylation of activation loop residues Thr202/Tyr204 and Thr185/Tyr187, respectively. Several downstream targets of p44/42 have been identified, including p90RSK (7) and the transcription factor Elk-1 (8,9). p44/42 are negatively regulated by a family of dual-specificity (Thr/Tyr) MAPK phosphatases, known as DUSPs or MKPs (10), along with MEK inhibitors, such as U0126 and PD98059

Product Includes	Item #	Kit Quantity	Color
FastScan™ ELISA Microwell Strip Plate*	53257	96 tests	
p44/42 MAPK (Erk1/2) Mouse Capture mAb	82448	1 each	Green (Lyophilized)
p44/42 MAPK (Erk1/2) Rabbit HRP-linked mAb	95915	1 each	Red (Lyophilized)
FastScan™ ELISA Capture Antibody Diluent	16076	3 ml	Green
FastScan™ ELISA HRP Antibody Diluent	28120	3 ml	
TMB Substrate	7004	11 ml	
STOP Solution	7002	11 ml	
Sealing Tape	54503	1 each	
ELISA Wash Buffer (20X)	9801	25 ml	
FastScan™ ELISA Cell Extraction Buffer (5X)	69905	10 ml	
FastScan™ ELISA Cell Extraction Enhancer Solution (50X)	25243	1 ml	
FastScan™ ELISA Kit #67404 Positive Control	35272	2 each	

<sup>\* 12 8-</sup>well modules -Each module is designed to break apart for 8 tests.



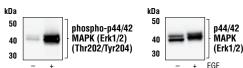


Figure 1. Treatment of A-431 cells with EGF stimulates phosphorylation of p44/42 MAPK (Erk1/2) at Thr202 and Tyr204, but does not affect the level of total p44/42 MAPK (Erk1/2). The relationship between lysate protein concentration from untreated and EGF-treated A-431 cells and the absorbance at 450 nm using the FastScan™ Total p44/42 MAPK (Erk1/2) ELISA Kit #67404 is shown in the upper figure. The corresponding western blots using phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) antibody (left panel) and p44/42 MAPK (Erk1/2) antibody (right panel) are shown in the lower figure. After serum starvation, A-431 cells were treated with 100 ng/ml EGF #8916 for 5 minutes at 37°C and then lysed.

# Background References:

- (1) Roux, P.P. and Blenis, J. (2004) *Microbiol Mol Biol Rev* 68, 320-44
- (2) Baccarini, M. (2005) FEBS Lett 579, 3271-7.
- (3) Meloche, S. and Pouysségur, J. (2007) *Oncogene* 26, 3227-39.
- (4) Roberts, P.J. and Der, C.J. (2007) Oncogene 26, 3291-310.
- (5) Rubinfeld, H. and Seger, R. (2005) Mol Biotechnol 31, 151-74.
- (6) Murphy, L.O. and Blenis, J. (2006) *Trends Biochem Sci* 31, 268-75.
- (7) Dalby, K.N. et al. (1998) J Biol Chem 273, 1496-505.
- (8) Marais, R. et al. (1993) Cell 73, 381-93.
- (9) Kortenjann, M. et al. (1994) Mol Cell Biol 14, 4815-24.
- (10) Owens, D.M. and Keyse, S.M. (2007) Oncogene 26, 3203-13.

U.S. Patents 7,429,487, 9,086,407, 9,261,500, and 9,476,874, foreign equivalents, and child patents deriving therefrom.

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#67404

# FastScan™ ELISA Protocol

## **A Solutions and Reagents**

**NOTE:** Prepare solutions with deionized/purified water or equivalent. Prepare only as much reagent as needed on the day of the experiment.

- FastScan™ ELISA Microwell Strip Plate, 96 well (#53257): Bring all to room temperature before opening bag/use. Unused microwell strips should be returned to the original re-sealable bag containing the desiccant pack and stored at 4°C.
- 1X ELISA Wash Buffer: Prepare by diluting ELISA Wash Buffer (20X) (included in each kit) to 1X with deionized water.
- 3. 1X Cell Extraction Buffer: Prepare by diluting FastScan™ ELISA Cell Extraction Buffer (5X) #69905 and FastScan™ ELISA Cell Extraction Enhancer Solution (50X) #25243\* to 1X with deionized water. This buffer can be stored at 4°C for short-term use (1-2 weeks). To make 10 mL 1X Cell Extraction Buffer, combine 7.8 mL deionized water, 2 mL FastScan™ ELISA Cell Extraction Buffer (5X), and 200 µL FastScan™ ELISA Cell Extraction Enhancer Solution (50X). Alternatively, Enhancer Solution may be added to the Cell Extraction Buffer after extraction of cells or tissue. When using the 1X Cell Extraction Buffer as a sample diluent for the assay, it is recommended to equilibrate it to room temperature prior to use.
  - \*IMPORTANT: The provided FastScan™ ELISA Cell Extraction Enhancer Solution (50X) may precipitate when stored at 4°C. To dissolve, warm briefly at 37°C and mix gently. The FastScan™ ELISA Cell Extraction Enhancer Solution (50X) can be stored at room temperature to avoid precipitation.

NOTE: The 1X Cell Extraction Buffer contains phosphatase inhibitors. Protease inhibitors should be added to the 1X Cell Extraction Buffer immediately prior to lysing cells. Additional phosphatase inhibitors can also be added (e.g. Protease/Phosphatase Inhibitor Cocktail (100X) #5872, not supplied).

- FastScan™ ELISA Capture Antibody Diluent: Green diluent for reconstitution of the Capture Antibody.
- FastScan™ ELISA HRP Antibody Diluent: Diluent (amber bottle) for reconstitution of the HRP-linked Antibody. Protect from light.
- 6. 4X Capture Antibody: Reconstitute lyophilized Capture Antibody (green colored cake) with 3 mL FastScan™ ELISA Capture Antibody Diluent (green diluent). Incubate at room temperature for 5 min with occasional gentle mixing to fully reconstitute. For best results, use immediately following antibody reconstitution. Unused reconstituted 4X Capture Antibody may be stored for up to 4 weeks at 4°C, although there may be some loss of signal compared to freshly reconstituted antibody.
- 7. 4X HRP-linked Antibody: Reconstitute lyophilized HRP-linked Antibody (red colored cake) with 3 mL FastScan™ ELISA HRP Antibody Diluent. Incubate at room temperature for 5 min with occasional gentle mixing to fully reconstitute. For best results, use immediately following antibody reconstitution. Unused reconstituted 4X HRP-linked Antibody may be stored for up to 4 weeks at 4°C protected from light, although there may be some loss of signal compared to freshly reconstituted antibody.
- 8. Antibody Cocktail: Combine equal volumes of the reconstituted 4X Capture and 4X HRP-linked Antibodies immediately prior to assay and mix. To make 6 mL of the Antibody Cocktail (enough for 1x 96-well plate), combine 3 mL 4X Capture Antibody with 3 mL 4X HRP-linked Antibody.
- 9. Positive Control: Reconstitute 1 vial of lyophilized Positive Control by adding 250 µL deionized water. Mix thoroughly and gently, hold at room temperature for 1 minute and then follow the steps outlined below in the "Test Procedure" section. Positive Controls are recommended to be used immediately after reconstituting in deionized water, however remaining material may be stored at -80°C (there may be some loss of the positive control signal if freeze/ thawed). Positive Controls are supplied as a control reagent, not as an absolute quantitation measure.
- 10. TMB Substrate (#7004): Bring to room temperature before use.
- 11. STOP Solution (#7002): Bring to room temperature before use.

## **Preparing Cell Lysates**

### For adherent cells.

- **1.** Aspirate media when the culture reaches 80–90% confluence.
- Remove media and rinse cells once with ice-cold 1X PBS.
- Remove PBS and add 0.5 mL ice-cold 1X Cell Extraction Buffer (recommended to supplement with protease inhibitors and additional phosphatase inhibitors as needed) to each plate (10 cm diameter) and incubate the plate on ice for 5 min.
- **4.** Scrape cells off the plate and transfer to an appropriate tube. Keep on ice.
- 5. Sonicate lysates on ice.
- 6. Microcentrifuge for 5 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

#### For suspension cells

- Remove media by low speed centrifugation (~1200 rpm) when the culture reaches 0.5–1.0 x 10<sup>6</sup> viable cells/ml.
- 2. Wash once with ice-cold 1X PBS
- Cells harvested from 50 mL of growth media can be lysed in 2.0 mL of 1X Cell Extraction Buffer (recommended to supplement with protease inhibitors and additional phosphatase inhibitors as needed).
- 4. Sonicate lysates on ice.
- Microcentrifuge for 5 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at –80°C in single-use aliquots.

### C Test Procedure

**NOTE:** Equilibrate all materials and prepared reagents to room temperature prior to running the assay.

- 1. Prepare all reagents as indicated above (Section A).
- 2. Samples should be undiluted or diluted with 1X Cell Extraction Buffer to a 2X protein concentration in order to achieve a final 1X protein concentration upon addition of the antibody cocktail. Individual datasheets for each kit provide a sensitivity curve that serves as a reference for selection of an appropriate starting lysate concentration. The sensitivity curve shows typical results across a range of lysate concentration points.
- 3. Add 50 µL of each sample or Positive Control to the appropriate wells.
- 4. Add 50 µL of the Antibody Cocktail to each well.
- 5. Seal the plate with the supplied sealing tape and incubate for 1 hour at room temperature on a plate shaker set to 400 rpm (moderate agitation).
- **6.** Gently remove the tape and wash wells:
  - a. Discard plate contents into a receptacle.
  - b. Wash 3 times with 1X ELISA Wash Buffer, 200 μL each time for every well. After each wash, aspirate or decant from wells. Invert the plate and blot it against clean paper towels to remove the residual solution in each well, but do not allow wells to completely dry at any time.
  - c. Clean the underside of all wells with a lint-free tissue.
- Add 100 µL of TMB Substrate to each well. Seal with tape and incubate the plate in the dark for 15 min at room temperature on a plate shaker (400 rpm, moderate agitation) or alternatively for 10 min at 37°C without shaking.
- Add 100 µL of STOP Solution to each well. Shake gently for a few seconds. NOTE: Initial color of positive reaction is blue, which changes to yellow upon addition of STOP Solution.
- 9. Read results:
  - a. Visual Determination: Read within 30 min after adding STOP Solution.
  - b. Spectrophotometric Determination: Wipe underside of wells with a lint-free tissue. Read absorbance at 450 nm within 30 min after adding STOP Solution.