Revision 2

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PathScan [®] Total E-Cadherin Sandw ELISA Kit	vich		Orders:
Species Cross Reactivity: UniProt ID: Entrez-Gene Id: H #P12830 #999 For Research Use Only. Not for Use in Diagnostic Procedures.		3 Trask Lar	Support: Web: ne Danvers Massa
Product Includes	Product #	Quantity	Color
E-Cadherin Rabbit Detection mAb	14263	1 ea	Green (Lyophilized)
Anti-rabbit IgG, HRP-linked Antibody (ELISA Formulated)	13272	1 ea	Red (Lyophilized)
Detection Antibody Diluent	13339	11 ml	Green
HRP Diluent	13515	11 ml	Red



Kit contents scale proportionally with size, except sealing tape.

Example: The V1 kit contains 5X the listed quantities above, but will exclude the sealing tape.

The microwell plate is supplied as 12 8-well modules - Each module is designed to break apart for 8 tests.

Description	The PathScan [®] Total E-Cadherin Sandwich ELISA Kit #7886 is a solid phase sandwich enzyme-linked immunosorbent assay (ELISA) that detects endogenous levels of E-cadherin. An E-cadherin mouse antibody has been coated onto the microwells. After incubation with cell lysates, E-cadherin is captured by the coated antibody. Following extensive washing, an E-Cadherin rabbit detection antibody is added to detect the captured E-cadherin. Anti-rabbit IgG, HRP-linked antibody is then used to recognize the bound detection antibody. HRP substrate, TMB, is added to develop color. The magnitude of the absorbance for this developed color is proportional to the quantity of E-cadherin. Antibodies in kit are custom formulations specific to kit.
Specificity/Sensitivity	CST's PathScan [®] Total E-Cadherin Sandwich ELISA Kit #7886 detects endogenous levels of E-cadherin. As shown in Figure 1, using the Total E-Cadherin Sandwich ELISA Kit #7886, a high level of E-cadherin is detected in A431 and MCF-7 cells, but not in E-cadherin null Jurkat cells. The PathScan [®] Total E- Cadherin Sandwich ELISA Kit #7886 does not cross-react with human N-, P- or VE-cadherin. This kit detects proteins from the indicated species, as determined through in-house testing, but may also detect homologous proteins from other species.
Background	Cadherins are a superfamily of transmembrane glycoproteins that contain cadherin repeats of approximately 100 residues in their extracellular domain. Cadherins mediate calcium-dependent cell-cell adhesion and play critical roles in normal tissue development (1). The classic cadherin subfamily includes N-, P-, R-, B-, and E-cadherins, as well as about ten other members that are found in adherens junctions, a cellular structure near the apical surface of polarized epithelial cells. The cytoplasmic domain of classical cadherins interacts with β -catenin, γ -catenin (also called plakoglobin), and p120 catenin. β -catenin and γ -catenin associate with α -catenin, which links the cadherin-catenin complex to the actin cytoskeleton (1,2). While β - and γ -catenin play structural roles in the junctional complex, p120 regulates cadherin adhesive activity and trafficking (1-4). Investigators consider E-cadherin an active suppressor of invasion and growth of many epithelial cancers (1-3). Research studies indicate that cancer cells have upregulated N-cadherin in addition to loss of E-cadherin. This change in cadherin expression is called the "cadherin switch." N-cadherin cooperates with the FGF receptor, leading to overexpression of MMP-9 and cellular invasion (3). Research studies have shown that in endothelial cells, VE-cadherin signaling, expression, and localization correlate with vascular permeability and tumor angiogenesis (5,6). Investigators have also demonstrated that expression of P-cadherin, which is normally present in epithelial cells, is also altered in ovarian and other human cancers (7,8).
Background References	1. Wheelock, M.J. and Johnson, K.R. (2003) Annu Rev Cell Dev Biol 19, 207-35.



Storage Temp

+4C

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	2. Christofori, G. (2003) <i>EMBO J</i> 22, 2318-23. 3. Hazan, R.B. et al. (2004) <i>Ann N Y Acad Sci</i> 1014, 155-63. 4. Bryant, D.M. and Stow, J.L. (2004) <i>Trends Cell Biol</i> 14, 427-34. 5. Rabascio, C. et al. (2004) <i>Cancer Res</i> 64, 4373-7. 6. Yamaoka-Tojo, M. et al. (2006) <i>Arterioscler Thromb Vasc Biol</i> 26, 1991-7. 7. Patel, I.S. et al. (2003) <i>Int J Cancer</i> 106, 172-7. 8. Sanders, D.S. et al. (2000) <i>J Pathol</i> 190, 526-30.
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#7886 PathScan[®] Total E-Cadherin Sandwich ELISA Kit



ELISA Colorimetric (Lyophilized)

A. Solutions and Reagents

NOTE: Prepare solutions with purified water.

- 1. Microwell strips: Bring all to room temperature before use.
- 2. **Detection Antibody**: Supplied lyophilized as a green colored cake or powder. Add 1.0 ml of Detection Antibody Diluent (green solution) to yield a concentrated stock solution. Incubate at room temperature for 5 min with occasional gentle mixing to fully reconstitute. To make the final working solution, add the 1.0 ml volume of reconstituted Detection Antibody to 10.0 ml of Detection Antibody Diluent in a clean tube and gently mix. Unused working solution may be stored for 4 weeks at 4°C.
- 3. **HRP-Linked Antibody***: Supplied lyophilized as a red colored cake or powder. Add 1.0 ml of HRP Diluent (red solution) to yield a concentrated stock solution. Incubate at room temperature for 5 min with occasional gentle mixing to fully reconstitute. To make the final working solution, add the 1.0 ml volume of reconstituted HRP-Linked Antibody to 10.0 ml of HRP Diluent in a clean tube and gently mix. Unused working solution may be stored for 4 weeks at 4°C.
- 4. Detection Antibody Diluent: Green colored diluent for reconstitution and dilution of the detection antibody.
- 5. HRP Diluent: Red colored diluent for reconstitution and dilution of the HRP-Linked Antibody.
- 6. Sample Diluent: Blue colored diluent provided for dilution of cell lysates.
- 7. **1X Wash Buffer**: Prepare by diluting 20X Wash Buffer (included in each PathScan[®] Sandwich ELISA Kit) in purified water.
- 8. **Cell Lysis Buffer**: 10X Cell Lysis Buffer #9803: This buffer can be stored at 4°C for short-term use (1–2 weeks). Recommended: Add 1 mM phenylmethylsulfonyl fluoride (PMSF) immediately before use.
- 9. TMB Substrate (#7004).
- 10. **STOP Solution** (#7002).

*NOTE: Some PathScan® ELISA Kits may include HRP-Linked Streptavidin in place of HRP-Linked Antibody.

B. Preparing Cell Lysates

For adherent cells.

- 1. Aspirate media when the culture reaches 80–90% confluence. Treat cells by adding fresh media containing regulator for desired time.
- 2. Remove media and rinse cells once with ice-cold 1X PBS.
- 3. Remove PBS and add 0.5 ml ice-cold 1X Cell Lysis Buffer plus 1 mM PMSF to each plate (10 cm diameter) and incubate the plate on ice for 5 min.
- 4. Scrape cells off the plate and transfer to an appropriate tube. Keep on ice.
- 5. Sonicate lysates on ice.
- 6. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

For suspension cells

- 1. Remove media by low speed centrifugation (\sim 1200 rpm) when the culture reaches 0.5–1.0 x 10⁶ viable cells/ml. Treat cells by adding fresh media containing regulator for desired time.
- 2. Collect cells by low speed centrifugation (~1200 rpm) and wash once with 5-10 ml ice-cold 1X PBS.
- 3. Cells harvested from 50 ml of growth media can be lysed in 2.0 ml of 1X Cell Lysis Buffer plus 1 mM PMSF.
- 4. Sonicate lysates on ice.
- 5. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

C. Test Procedure

- 1. After the microwell strips have reached room temperature, break off the required number of microwells. Place the microwells in the strip holder. Unused microwells must be resealed and stored at 4°C immediately.
- 2. Cell lysates can be undiluted or diluted with Sample Diluent (supplied in each PathScan[®] Sandwich ELISA Kit, blue color). Individual datasheets for each kit provide a sensitivity curve that serves as a reference for selection of an appropriate starting lysate concentration. The sensitivity curve shows typical kit assay results across a range of lysate concentration points.
- 3. Add 100 µl of each undiluted or diluted cell lysate to the appropriate well. Seal with tape and press firmly onto top of microwells. Incubate the plate for 2 hr at 37°C. Alternatively, the plate can be incubated overnight at 4°C.

- 4. Gently remove the tape and wash wells:
 - 1. Discard plate contents into a receptacle.
 - 2. Wash 4 times with 1X Wash Buffer, 200 µl each time for each well.
 - 3. For each wash, strike plates on fresh towels hard enough to remove the residual solution in each well, but do not allow wells to completely dry at any time.
 - 4. Clean the underside of all wells with a lint-free tissue.
- 5. Add 100 µl of reconstituted Detection Antibody (green color) to each well (refer to Section A, Step 2). Seal with tape and incubate the plate at 37°C for 1 hr.
- 6. Repeat wash procedure (Section C, Step 4).
- 7. Add 100 μ l of reconstituted HRP-Linked secondary antibody (red color) to each well (refer to Section A, Step 3). Seal with tape and incubate the plate for 30 min at 37°C.
- 8. Repeat wash procedure (Section C, Step 4).
- 9. Add 100 μ l of TMB Substrate to each well. Seal with tape and incubate the plate for 10 min at 37°C or 30 min at 25°C.
- 10. Add 100 μl of STOP Solution to each well. Shake gently for a few seconds.

NOTE: Initial color of positive reaction is blue, which changes to yellow upon addition of STOP Solution.

- 11. Read results.
 - 1. Visual Determination: Read within 30 min after adding STOP Solution.
 - 2. Spectrophotometric Determination: Wipe underside of wells with a lint-free tissue. Read absorbance at 450 nm within 30 min after adding STOP Solution.

posted November 2013