HTScan[™] ABL1 Kinase Assay Kit

100 Assays (96 Well Format)



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rev. 04/06/05

Products Included	Product #	Kit Quantity
Phospho-Tyrosine Monoclonal Antibody (P-Tyr-100)	9411	30 µl
HTScan [™] Tyrosine Kinase Buffer (4x)	9805	15 ml
DTT (1000x, 1.25 M)		80 µl
ATP (10 mM)	9804	1 ml
Peptide Substrate Biotin-Signal Transduction Protein (Tyr160)		1.25 ml
ABL1 Kinase (recombinant, human)	7700	1000 Units

Description: The kit provides a means of performing enzymatic assays with active human ABL1 kinase. It includes active ABL1 kinase (supplied as a GST fusion protein), a biotinylated substrate peptide and a phosphotyorsine monoclonal antibody for detection of the phosphorylated form of the substrate peptide.

Peptide Core Sequence: GIY*DV

Molecular Weights: Peptide Substrate, Biotin-Signal Transduction Protein (Tyr160): 1830 Daltons, GST-ABL1 Kinase domain: 74,140 Daltons

Unit Definition: 10 Units is defined as the amount of ABL1 kinase required to maximally phosphorylate 75 pmol of Signal Transduction Protein (Tyr160) #C03-719 biotinylated substrate peptide in 30 minutes at 25°C in a total reaction volume of 50 µl quantified by DELFIA[®].

Background: The c-Abl proto-oncogene encodes a nonreceptor type protein tyrosine kinase that is ubiquitously expressed and highly conserved in metazoan evolution. c-Abl protein is distributed in both the nucleus and the cytoplasm of cells. It has been implicated in regulation of cell proliferation, differentiation, apoptosis, cell adhesion and stress response (1–3). c-Abl kinase activity is increased *in vivo* by diverse physiological stimuli including integrin activation, PDGF stimulation and binding to c-Jun, Nck and RFX1 (2,4). The *in vivo* mechanism of regulation of c-Abl kinase activity is not completely



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2004

Figure 1. Time course of ABL1 kinase activity: DELFIA® data generated using Phospho-Tyrosine Monoclonal Antibody P-Tyr-100 #9411 to detect phosphorylation of ABL1 substrate peptide by GST-ABL1 kinase. In a 50 µl reaction, 10 Units GST-ABL1 and 1.5 µM substrate peptide were used per reaction well. Background reading is 768. (DELFIA® is a registered trademark of PerkinElmer, Inc.) understood. Tyrosine 245 is located in the linker region between the SH2 and catalytic domains, conserved among the AbI family. Phosphorylation of Tyr245 is involved in activation of c-AbI kinase activity (5).

Tyrosine 412 is located in the kinase activation loop of c-Abl, and phosphorylation of this residue is required for kinase activity (6). Thr735 is in a conserved 14-3-3 protein binding motif, which can be phosphorylated upon stress stimulation or TPA treatment (Wu, J. et al. unpublished data). Phosphorylation of Thr735 in c-Abl may play an important role in regulation of c-Abl localization as well as its function.

Source/Purification: The GST-Kinase fusion protein was produced using a baculovirus expression system from a construct containing a human ABL1 cDNA (Pro118-Ser553) fragment amino-terminally fused to a GST-HIS₆-Thrombin cleavage site. The protein was then purified by one-step affinity purification using glutathione-agarose.

The detection antibody, Phospho-Tyrosine Monoclonal Antibody (P-Tyr-100) #9411 was derived from mice immunized with phospho-tyrosine-containing peptides (KLH coupled).

Quality Control: Biotin-Signal Transduction Protein (Tyr160) peptide was selected by using Tyrosine Kinase Substrate Screening Kit #7450 to screen for ABL1 kinase substrates. Phospho-Tyrosine Monoclonal Antibody #9411 was used for detection (fig.2). The quality of the biotinylated peptides was evaluated by reverse-phase HPLC and by mass spectrometry.

Purified GST-ABL1 kinase was quality controlled for purity using silver stain SDS-PAGE and Western blot. ABL1 kinase V_{max} and K_m values were measured to determine specific enzymatic activity (fig.5).

Assay conditions (time course [fig.1], kinase dosedependence [fig.3], substrate dose-dependence [fig.4], and staurosporine inhibition [fig.6]) for ABL1 kinase activity were verified using the ABL1 substrate peptide provided in this kit. Storage: Antibodies are supplied in in 10 mM sodium HEPES (pH 7.5), 150 mM NaCl, 100 μ g/ml BSA and 50% glycerol. Do not aliquot the antibodies. Peptides are supplied at 6 μ M in 0.001% DMSO. Enzymes are supplied in 50 mM Tris-HCL (pH 8.0), 100 mM NaCl, 5 mM DTT, 15 mM reduced glutathion and 20% glycerol. Store at -80°C.

Keep enzymes on ice during use.

Avoid repeated freeze-thaw cycles.

Companion Products:

Serine/Threonine Kinase Substrate Screening Kit #7400

Tyrosine Kinase Substrate Screening Kit #7450

Staurosporine #9953

ABL1 Kinase #7700



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Figure 2. Treatment of Tyrosine Kinase Substrate Screening Kit #7450 with GST-ABL1 Kinase. Positive control is in yellow and negative control is in blue. Several positive hits for ABL1 were identified using this approach (in red). The star indicates the peptide chosen for this HTScan™ ABL1 Kinase Assay Kit.



Figure 3. Dose dependence curve of ABL1 kinase activity: DELFIA® data generated using Phospho-Tyrosine Monoclonal Antibody P-Tyr-100 #9411 to detect phosphorylation of peptide C03-719 by GST-ABL1 kinase. In a 50 µl reaction, increasing amounts of GST-ABL1 and 1.5 µM substrate peptide were used per reaction well at 25°C for 30 minutes. Background reading is 1481. (DELFIA® is a registered trademark of PerkinElmer, Inc.)



Figure 4. Peptide concentration dependence of ABL1 kinase activity: DELFIA® data generated using Phospho-Tyrosine Monoclonal Antibody P-Tyr-100 #9411 to detect phosphorylation of peptide C03-719 by GST-ABL1 kinase. In a 50 µl reaction, 10 Units of GST-ABL1 and increasing concentrations of substrate peptide were used per reaction well at 25°C for 30 minutes. Background reading is 871. (DELFIA® is a registered trademark of PerkinElmer, Inc.)



Figure 5. ABL1 kinase activity was measured in a radioisotopic filtration assay using the following reaction conditions: 60 mM HEPES-NaOH, pH 7.5, 3 mM MgCl₂, 3 mM MnCl₂, 3 µM Na-orthovanadate, 1.2 mM DTT, ATP (variable), 2.5 µg / 50 µl PEG20.000, Substrate: PolyAEKY, 5 µg / 50 µl, Recombinant ABL1: 5 Units/50 µl.



Figure 6. Staurosporine inhibition of ABL1 kinase activity: DELFIA® data generated using Phospho-Tyrosine Monoclonal Antibody P-Tyr-100 #9411 to detect phosphorylation of ABL1 substrate peptide (C03-719) by GST-ABL1 kinase. In a 50 µI reaction, 10 Units GST-ABL1, 1.5 µM substrate peptide, 5 µM ATP and increasing amount of staurosporine were used per reaction well at 25°C for 30 minutes. (DELFIA® is a registered trademark of PerkinElmer, Inc.)



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Background References:

- (1) Wang, J. Y. et al. (2000) Oncogene 19, 5643–5650.
- (2) Van Etten, R. A. et al. (1999) *Trends Cell. Biol.* 9, 179–182.
- (3) Danial, N. N. et al. (2000) *Oncogene* 19, 2523–2531.
- (4) Shaul, Y. et al. (2000) Cell Death Differ. 7, 10–16.
- (5) Brasher, B. B. et al. (2000) J. Biol. Chem. 275, 35631–35637.
- (6) Pluk, H. et al. (2002) Cell 108, 247-259.

Protocol for HTScan™ ABL1 Kinase Assay Kit

Suggested Kinase Assay Conditions:

■ Combine 12.5 µl of 6 µM peptide substrate and 12.5 µl H₂O with kinase inhibitor or activator of interest. Add 2x kinase reaction cocktail (see below). Incubate at 25°C for 30 minutes.

Peptide substrate is supplied in H_2O plus 0.001 % DMSO.

Note: Lot-specific information for this kinase is provided on the enzyme vial. Optimal assay incubation times and enzyme concentrations must be determined empirically for each kinase.

Additional Solutions and Reagents (Not included)

- Wash Buffer: 1X PBS, 0.05% Tween-20 (PBS/T)
- Bovine Serum Albumin (BSA)
- Stop Buffer: 50 mM EDTA pH 8
- DELFIA[®] Europium-labeled Anti-mouse IgG (PerkinElmer Life Sciences #AD0124)
- DELFIA[®] Enhancement Solution (PerkinElmer Life Sciences #1244-105)

■ DELFIA[®] Streptavidin coated, 96-well, yellow plate (PerkinElmer Life Sciences AAAND-0005)

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Suggested Protocol For 100 Assays

- 1. Transfer enzyme from -80°C immediately to ice. Allow enzyme to thaw on ice.
- 2. Microcentrifuge briefly at 4°C to bring all liquid to the bottom of the vial. Return immediately to ice.
- 3. Add 1.25 ml 6 μ M peptide substrate to 1.25 ml H₂O (include kinase inhibitor or activator of interest if desired). Aliquot 25 μ l/well into a 96 well plate.
- 4. Set up 2x Reaction cocktails (Mock/No Kinase and Plus Kinase) in tubes on ice.

2.5 ml 4X HTScan[™] Tyrosine Kinase Buffer (240 mM HEPES pH 7.5, 20 mM MgCl₂, 20 mM MnCl₂, 12 μ M Na₃VO₄)

20-200 µl ATP (10 mM)

10 µl DTT (1.25 M)

Bring to 5 ml with H₂0. Mix gently by inversion

Transfer enzyme to 2.5 ml 2X reaction cocktail. Final enzyme concentration should be 10 Units/25 µl in 2X reaction cocktail.

5. Add 25 μ l 2x kinase reaction cocktail (containing kinase) to 25 μ l diluted peptide in each well. Mix by gentle agitation.

Final Assay Conditions for a 50 μI Reaction

60 mM HEPES pH 7.5 5 mM MgCl₂ 5 mM MnCl₂ 3 μM Na₃VO₄ 1.25 mM DTT 20–200 μM ATP 1.5 μM peptide 10 Units Kinase

- 4. Incubate reaction plate at 25°C for 30 minutes.
- 5. Add 50 µl Stop Buffer (50 mM EDTA, pH 8) to each well and mix by pipetting.
- 6. Transfer 25 μI of each reaction and 75 μI H_2O/well to a 96-well streptavidin-coated plate and incubate at room temperature for 60 minutes.
- 7. *Wash three times with 200 µl/well PBS/T.
- Dilute primary antibody, Phospho-Tyrosine Monoclonal Antibody (P-Tyr-100) #9411, 1:1000 in PBS/T with 1% BSA. Add 100 μl primary antibody per well.
- 9. Mix and incubate at room temperature with rocking for 60 minutes.
- 10. *Wash three times with 200 µl/well PBS/T.
- 11. Dilute Europium labeled anti-mouse IgG 1:500 in PBS/T with 1% BSA. Add 100 µl diluted antibody/well.
- 12. Mix and incubate at room temperature for 30 minutes.
- 13. *Wash five times with 200 µl/well PBS/T.
- 14. Add 100 µl/well DELFIA® Enhancement Solution.
- 15. Incubate at room temperature for 5 minutes.
- 16. Detect 615 nm fluorescence emission with appropriate Time-Resolved Plate Reader.

*IMPORTANT: Use of an automated microplate washer as well as centrifugation of plates when appropriate, greatly improves reproducibility.

Please contact Cell Signaling Technology for HTS-ready antibodies (PBS formulated and carrier-free), and detailed peptide substrate sequence information. Email: <u>drugdiscovery@cellsignal.com</u>