



Orders: 877-616-CELL (2355)
orders@cellsignal.com

Support: 877-678-TECH (8324)

Web: info@cellsignal.com
cellsignal.com

3 Trask Lane | Danvers | Massachusetts | 01923 | USA

Store at +4C
#7274

PathScan[®] MAP Kinase Multi-Target Sandwich ELISA Kit

Species Cross
Reactivity:
H M

UniProt ID:
#Q16539, #P45983, #P27361,
#P28482, #Q02750

Entrez-Gene Id:
#1432, #5599, #5595,
#5594, #5604

For Research Use Only. Not for Use in Diagnostic Procedures.

Product Includes	Product #	Quantity	Color	Storage Temp
MAP Kinase Multi-Target	79155	96 tests		+4C
TMB Substrate	7004	11 ml	Colorless	+4C
STOP Solution	7002	11 ml	Colorless	+4C
Sealing Tape	54503	2 ea		+4C
ELISA Wash Buffer (20X)	9801	25 ml	Colorless	+4C
ELISA Sample Diluent	11083	25 ml	Blue	+4C
Cell Lysis Buffer (10X)	9803	15 ml	Yellowish	-20C

Kit contents scale proportionally with size, except sealing tape.

Example: The V1 kit contains 5X the listed quantities above, but will exclude the sealing tape.

The microwell plate is supplied as 12 8-well modules - Each module is designed to break apart for 8 tests.

Description

CST's PathScan[®] MAP Kinase Multi-Target Sandwich ELISA Kit is a solid phase sandwich enzyme-linked immunosorbent assay (ELISA) that combines the reagents necessary to detect endogenous levels of phospho-p44/42 MAPK (Thr202/Tyr204), phospho-p38 MAPK (Thr180/Tyr182), MEK1, phospho-MEK1 (Ser217/221), SAPK/JNK and phospho-SAPK/JNK (Thr183/Tyr185). These molecules represent convergence points and key regulatory proteins in signaling pathways controlling cellular events such as growth, differentiation and the response to stress and inflammation. Sixteen tests are provided for each target protein. Specific assay formulations for the indicated target proteins can be found in the datasheets associated with the individual PathScan[®] Sandwich ELISA Kits**. Briefly, a capture antibody has been coated onto the microwells. After incubation with cell lysates, the target protein is captured by the coated antibody. Following extensive washing, a detection antibody is added to detect the captured target protein. An HRP-linked secondary antibody is then used to recognize the bound detection antibody. HRP substrate, TMB, is added to develop color. The magnitude of absorbance for this developed color is proportional to the quantity of bound target protein. *Antibodies in kit are custom formulations specific to kit. **See companion products.

Specificity/Sensitivity

CST's PathScan[®] MAP Kinase Multi-Target Sandwich ELISA Kit #7274 detects endogenous levels of six proteins: phospho-p44/42 MAPK (Thr202/Tyr204), phospho-p38 MAPK (Thr180/Tyr182), MEK1, phospho-MEK1 (Ser217/221), SAPK/JNK and phospho-SAPK/JNK (Thr183/Tyr185). Differential phosphorylation of these proteins can be observed over time in response to various growth factor and cytokine treatments, as shown in Figure 1. The relationship between the protein concentration of the lysate and the absorbance at 450 nm can be found in the datasheets associated with the individual PathScan[®] Sandwich ELISA Kits**. **See companion products.

This kit detects proteins from the indicated species, as determined through in-house testing, but may also detect homologous proteins from other species.

Background

Both p44 and p42 MAP kinases (Erk1 and Erk2) function in a protein kinase cascade that plays a critical role in the regulation of cell growth and differentiation (1-6). MAP kinases are activated by a wide variety of extracellular signals including growth and neurotrophic factors, cytokines, hormones and neurotransmitters. Activation of MAP kinases occurs through phosphorylation of threonine and tyrosine (Thr202 and Tyr204 of human MAP kinase or Thr183 and Tyr185 of rat MAP kinase) at the sequence T*EY* by a single upstream MAP kinase kinase (MEK) (7,8).

MEK1 and MEK2 are dual-specificity protein kinases that function in a mitogen activated protein kinase cascade controlling cell growth and differentiation. Activation of MEK1 and MEK2 occurs through phosphorylation of Ser217 and Ser221 by Raf-like molecules. MEK activates p44 and p42 MAP kinase (1,9,10).

p38 MAP kinase (MAPK) participates in a signaling cascade controlling the cellular response to pro-inflammatory cytokines and a variety of cellular stresses. MKK3, MKK6 and SEK (MKK4) activate p38 MAP kinase by phosphorylation at Thr180 and Tyr182 (11-14).

The stress-activated protein kinase/Jun-amino-terminal kinase SAPK/JNK is activated by a variety of environmental stresses, including UV and gamma radiation, ceramides, inflammatory cytokines and in some instances, by growth factors and GPCR agonists (15-20). As with the other MAPKs, the core-signaling unit is composed of a MAPKKK, typically MEKK1-4, or by a mixed lineage kinase (MLK), which phosphorylates and activates MKK4-7, which then phosphorylates Thr183 and Tyr185 to activate the SAPK/JNK kinase (16). Stress signals are delivered to this cascade by small GTPases of the Rho family (Rac, Rho, cdc42) (17). Both Rac1 and cdc42 mediate the stimulation of MEKKs and MLKs (17). Alternatively, MKK4-7 can be activated by a pathway independent of small GTPases via stimulation of a member of the germinal center kinase (GCK) family (18).

Background References

1. McKay, M.M. and Morrison, D.K. (2007) *Oncogene* 26, 3113-21.
2. Pearson, G. et al. (2001) *Endocr Rev* 22, 153-83.
3. Marshall, C.J. (1995) *Cell* 80, 179-85.
4. Hunter, T. (1995) *Cell* 80, 225-36.
5. Hill, C.S. and Treisman, R. (1995) *Cell* 80, 199-211.
6. Cowley, S. et al. (1994) *Cell* 77, 841-52.
7. Sturgill, T.W. et al. (1988) *Nature* 334, 715-8.
8. Payne, D.M. et al. (1991) *EMBO J* 10, 885-92.
9. Alessi, D.R. et al. (1994) *EMBO J* 13, 1610-9.
10. Pearson, G. et al. (2001) *Endocr Rev* 22, 153-83.
11. Raingeaud, J. et al. (1995) *J Biol Chem* 270, 7420-6.
12. Raman, M. et al. (2007) *Oncogene* 26, 3100-12.
13. Zarubin, T. and Han, J. (2005) *Cell Res* 15, 11-8.
14. Roux, P.P. and Blenis, J. (2004) *Microbiol Mol Biol Rev* 68, 320-44.
15. Davis, R.J. (1999) *Biochem Soc Symp* 64, 1-12.
16. Ichijo, H. (1999) *Oncogene* 18, 6087-93.
17. Kyriakis, J.M. and Avruch, J. (2001) *Physiol Rev* 81, 807-69.
18. Kyriakis, J.M. (1999) *J Biol Chem* 274, 5259-62.
19. Leppä, S. and Bohmann, D. (1999) *Oncogene* 18, 6158-62.
20. Whitmarsh, A.J. and Davis, R.J. (1998) *Trends Biochem Sci* 23, 481-5.

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#7274

PathScan[®] MAP Kinase Multi-Target Sandwich ELISA Kit

ELISA Colorimetric

NOTE: Refer to product-specific datasheets or product webpage for assay incubation temperature.

A. Solutions and Reagents

NOTE: Prepare solutions with reverse osmosis deionized (RODI) or equivalent grade water.

1. **20X Phosphate Buffered Saline (PBS):** (#9808) To prepare 1 L PBS: add 50 ml 10X PBS to 950 ml dH₂O, mix.
2. Bring all microwell strips to room temperature before use.
3. Prepare 1X Wash Buffer by diluting 20X Wash Buffer (included in each PathScan[®] Sandwich ELISA Kit) in dH₂O.
4. **1X Cell Lysis Buffer:** 10X Cell Lysis Buffer (#9803): To prepare 10 ml of 1X Cell Lysis Buffer, add 1 ml of 10X Cell Lysis Buffer to 9 ml of dH₂O, mix. Buffer can be stored at 4°C for short-term use (1-2 weeks).

Recommended: Add 1 mM phenylmethylsulfonyl fluoride (PMSF) (#8553) immediately before use.

NOTE: Refer to product-specific datasheet or webpage for lysis buffer recommendation.

5. **TMB Substrate:** (#7004).
6. **STOP Solution:** (#7002).

B. Preparing Cell Lysates

For adherent cells

1. Aspirate media when the culture reaches 80–90% confluence. Treat cells by adding fresh media containing regulator for desired time.
2. Remove media and rinse cells once with ice-cold 1X PBS.
3. Remove PBS and add 0.5 ml ice-cold 1X cell lysis buffer plus 1 mM PMSF to each plate (10 cm diameter) and incubate the plate on ice for 5 min.
4. Scrape cells off the plate and transfer to an appropriate tube. Keep on ice.
5. Sonicate lysates on ice.
6. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

For suspension cells

1. Remove media by low speed centrifugation (~1,200 rpm) when the culture reaches 0.5–1.0 x 10⁶ viable cells/ml. Treat cells by adding fresh media containing regulator for desired time.
2. Collect cells by low speed centrifugation (~1,200 rpm) and wash once with 5–10 ml ice-cold 1X PBS.
3. Cells harvested from 50 ml of growth media can be lysed in 2.0 ml of 1X cell lysis buffer plus 1 mM PMSF.
4. Sonicate lysates on ice.
5. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

C. Test Procedure

1. After the microwell strips have reached room temperature, break off the required number of microwells. Place the microwells in the strip holder. Unused microwells must be resealed in the storage bag and stored at 4°C immediately.
2. Cell lysates can be undiluted or diluted with sample diluent (supplied in each PathScan[®] Sandwich ELISA Kit, blue color). Individual datasheets or product webpage for each kit provide information regarding an appropriate dilution factor for lysates and kit assay results.
3. Add 100 µl of each undiluted or diluted cell lysate to the appropriate well. Seal with tape and press firmly onto top of microwells. Incubate the plate for 2 hr at 37°C. Alternatively, the plate can be incubated overnight at 4°C.
4. **Gently remove the tape and wash wells:**
 1. Discard plate contents into a receptacle.
 2. Wash 4 times with 1X wash buffer, 200 µl each time per well.
 3. For each wash, strike plates on fresh paper towels hard enough to remove the residual solution in each well, but do not allow wells to completely dry at any time.
 4. Clean the underside of all wells with a lint-free tissue.
5. Add 100 µl of detection antibody (green color) to each well. Seal with tape and incubate the plate at 37°C for 1 hr.

6. Repeat wash procedure (Section C, Step 4).
7. Add 100 μ l of HRP-linked secondary antibody (red color) to each well. Seal with tape and incubate the plate for 30 min at 37°C.
8. Repeat wash procedure (Section C, Step 4).
9. Add 100 μ l of TMB substrate to each well. Seal with tape and incubate the plate for 10 min at 37°C or 30 min at 25°C.
10. Add 100 μ l of STOP solution to each well. Shake gently for a few seconds.

NOTE: Initial color of positive reaction is blue, which changes to yellow upon addition of STOP solution.

11. Read results

1. **Visual Determination:** Read within 30 min after adding STOP solution.
2. **Spectrophotometric Determination:** Wipe underside of wells with a lint-free tissue. Read absorbance at 450 nm within 30 min after adding STOP solution.

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