e at +4C	PathScan <sup>®</sup> Ph (Thr202/Tyr20	oody				
Stor	Pair				Orders:	877-616-CELL (2355) orders@cellsignal.com
.0	1 Kit (Reagents for	r 4 x 96 well plat	es)		Support:	877-678-TECH (8324)
7246	Species Cross Reactivity: H M	<b>UniProt ID:</b> #P27361, #P28482	<b>Entrez-Gene Id:</b> #5595, #5594		Web:	info@cellsignal.com cellsignal.com
#				3 Trask Lane   I	Danvers   Mass	sachusetts   01923   USA
For Re	esearch Use Only. Not fo	or Use in Diagnosti	c Procedures.			

Product Includes	Product #	Volume	Cap Color	Storage Temp
Phospho-p44/42 MAPK (Thr202/Tyr204) Capture Rabbit mAb (100X)	10184	400 µl	Pink	+4C
p44/42 MAPK Detection Mouse mAb (100X)	29005	400 µl	Blue	+4C
Anti-mouse IgG, HRP-linked Antibody (1000X)	16736	40 µl	Yellow	-20C

Please visit cellsignal.com for a complete listing of recommended companion products.

Description	CST's PathScan <sup>®</sup> Phospho-p44/42 MAPK (Thr202/Tyr204) Sandwich ELISA Antibody Pair is offered as an economical alternative to our PathScan <sup>®</sup> Phospho-p44/42 MAPK (Thr202/Tyr204) Sandwich ELISA Kit #7177. Capture and detection antibodies (100X stocks) and an HRP-conjugated secondary antibody (1000X stock) are supplied. Sufficient reagents are supplied for 4 x 96 well ELISAs. The Phospho-p44/42 MAPK Rabbit Capture Antibody is coated in PBS overnight in a 96 well microplate. After blocking, cell lysate is added followed by a p44/42 MAPK Mouse Detection Antibody and an HRP-conjugated Anti-Mouse IgG Antibody. HRP substrate, TMB, is added for color development. The magnitude of the absorbance for this developed color is proportional to the quantity of p44/42 MAPK phosphorylated at Thr202/Tyr204. Antibodies in kit are custom formulations specific to kit.
Reagents Not Supplied	Phosphate Buffered Saline (PBS-20X) #9808 Phosphate Buffered Saline with Tween-20 (PBST-20X) #9809 Cell Lysis Buffer (10X) #9803 TMB Substrate #7004 STOP Solution #7002 Blocking Buffer: 1X PBS/0.5% Tween-20, 1% BSA 96 Well Microplates** Microplate Reader ** Antibody Pairs have been validated on Corning© 96 Well Clear Polystyrene High Bind Stripwell <sup>™</sup> Microplates (#2592). Notes: Antibody pairs have been optimized using recommended buffers, reagents, plates and the included protocol. Solutions should be made fresh daily.
Background	Mitogen-activated protein kinases (MAPKs) are a widely conserved family of serine/threonine protein kinases involved in many cellular programs, such as cell proliferation, differentiation, motility, and death. The p44/42 MAPK (Erk1/2) signaling pathway can be activated in response to a diverse range of extracellular stimuli, including mitogens, growth factors, and cytokines (1-3), and research investigators consider it an important target in the diagnosis and treatment of cancer (4). Upon stimulation, a sequential three-part protein kinase cascade is initiated, consisting of a MAP kinase kinase kinase (MAPKK or MAP3K), a MAP kinase kinase (MAPKK or MAP2K), and a MAP kinase (MAPK). Multiple p44/42 MAP3Ks have been identified, including members of the Raf family, as well as Mos and Tpl2/COT. MEK1 and MEK2 are the primary MAPKKs in this pathway (5,6). MEK1 and MEK2 activate p44 and p42 through phosphorylation of activation loop residues Thr202/Tyr204 and Thr185/Tyr187, respectively. Several downstream targets of p44/42 have been identified, including p90RSK (7) and the transcription factor Elk-1 (8,9). p44/42 are negatively regulated by a family of dual-specificity (Thr/Tyr) MAPK phosphatases, known as DUSPs or MKPs (10), along with MEK inhibitors, such as U0126 and PD98059.
Background References	<ol> <li>Roux, P.P. and Blenis, J. (2004) <i>Microbiol Mol Biol Rev</i> 68, 320-44.</li> <li>Baccarini, M. (2005) <i>FEBS Lett</i> 579, 3271-7.</li> <li>Meloche, S. and Pouysségur, J. (2007) <i>Oncogene</i> 26, 3227-39.</li> <li>Roberts, P.J. and Der, C.J. (2007) <i>Oncogene</i> 26, 3291-310.</li> <li>Rubinfeld, H. and Seger, R. (2005) <i>Mol Biotechnol</i> 31, 151-74.</li> <li>Murphy, L.O. and Blenis, J. (2006) <i>Trends Biochem Sci</i> 31, 268-75.</li> <li>Dalby, K.N. et al. (1998) <i>J Biol Chem</i> 273, 1496-505.</li> <li>Marais, R. et al. (1993) <i>Cell</i> 73, 381-93.</li> <li>Kortenjann, M. et al. (1994) <i>Mol Cell Biol</i> 14, 4815-24.</li> </ol>

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# **#7246** PathScan<sup>®</sup> Phospho-p44/42 MAPK (Thr202/Tyr204) Sandwich ELISA Antibody Pair



## **ELISA Antibody Pair**

## A. Solutions and Reagents

NOTE: Prepare solutions with reverse osmosis deionized (RODI) or equivalent grade water.

- 1. 20X Phosphate Buffered Saline (PBS): (#9808) To prepare 1 L 1X PBS: add 50 ml 20X PBS to 950 ml dH<sub>2</sub>O,
- mix. 2. Wash Buffer: 1X PBS/0.05% Tween<sup>®</sup> 20, (20X PBST #9809).
- 3. Blocking Buffer: 1X PBS/0.05% Tween<sup>®</sup> 20, 1% BSA.
- 4. **1X Cell Lysis Buffer**: 10X Cell Lysis Buffer (#9803): To prepare 10 ml of 1X Cell Lysis Buffer, add 1 ml of 10X Cell Lysis Buffer to 9 ml of dH<sub>2</sub>O, mix. Buffer can be stored at 4°C for short-term use (1–2 weeks).

Recommended: Add 1 mM phenylmethylsulfonyl fluoride (PMSF) (#8553) immediately before use.

- 5. Bovine Serum Albumin (BSA): (#9998).
- 6. TMB Substrate: (#7004).
- 7. **STOP Solution**: (#7002)

NOTE: Reagents should be made fresh daily.

### **B.** Preparing Cell Lysates

#### For adherent cells

- 1. Aspirate media when the culture reaches 80–90% confluence. Treat cells by adding fresh media containing regulator for desired time.
- 2. Remove media and rinse cells once with ice-cold 1X PBS.
- 3. Remove PBS and add 0.5 ml ice-cold 1X Cell Lysis Buffer plus 1 mM PMSF to each plate (10 cm diameter) and incubate the plate on ice for 5 min.
- 4. Scrape cells off the plate and transfer to an appropriate tube. Keep on ice.
- 5. Sonicate lysates on ice.
- 6. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

#### For suspension cells

- 1. Remove media by low speed centrifugation ( $\sim$ 1,200 rpm) when the culture reaches 0.5–1.0 x 10<sup>6</sup> viable cells/ml. Treat cells by adding fresh media containing regulator for desired time.
- 2. Collect cells by low speed centrifugation (~1,200 rpm) and wash once with 5-10 ml ice-cold 1X PBS.
- 3. Cells harvested from 50 ml of growth media can be lysed in 2.0 ml of 1X cell lysis buffer plus 1 mM PMSF.
- 4. Sonicate lysates on ice.
- 5. Microcentrifuge for 10 min (x14,000 rpm) at 4°C and transfer the supernatant to a new tube. The supernatant is the cell lysate. Store at -80°C in single-use aliquots.

## **C. Coating Procedure**

- 1. Rinse microplate with 200  $\mu$ l of dH<sub>2</sub>O, discard liquid. Blot on paper towel to make sure wells are dry.
- 2. Dilute capture antibody 1:100 in 1X PBS. For a single 96 well plate, add 100 μl of capture antibody stock to 9.9 ml 1X PBS. Mix well and add 100 μl/well. Cover plate and incubate overnight at 4°C (17-20 hr).
- 3. After overnight coating, gently uncover plate and wash wells:
  - 1. Discard plate contents into a receptacle.
  - 2. Wash four times with wash buffer, 200  $\mu$ l each time per well. For each wash, strike plates on fresh paper towels hard enough to remove the residual solution in each well, but do not allow wells to completely dry at any time.
  - 3. Clean the underside of all wells with a lint-free tissue.
- 4. Block plates. Add 150  $\mu$ l of blocking buffer/well, cover plate, and incubate at 37°C for 2 hr.
- 5. After blocking, wash plate (Section C, Step 3). Plate is ready to use.

#### **D. Test Procedure**

- 1. Lysates can be used undiluted or diluted in blocking buffer. 100  $\mu$ l of lysate is added per well. Cover plate and incubate at 37°C for 2 hr.
- 2. Wash plate (Section C, Step 3).
- 3. Dilute detection antibody 1:100 in blocking buffer. For a single 96 well plate, add 100  $\mu$ l of detection antibody Stock to 9.9 ml of blocking buffer. Mix well and add 100  $\mu$ l/well. Cover plate and incubate at 37°C for 1 hr.
- 4. Wash plate (Section C, Step 3).
- 5. Secondary antibody, either streptavidin anti-mouse or anti-rabbit-HRP, is diluted 1:1000 in blocking buffer. For a single 96 well plate, add 10 µl of secondary antibody stock to 9.99 ml of blocking buffer. Mix well and add 100 µl/well. Cover and incubate at 37°C for 30 min.
- 6. Wash plate (Section C, Step 3).
- 7. Add 100  $\mu l$  of TMB substrate per well. Cover and incubate at 37°C for 10 min.
- 8. Add 100 µl of STOP solution per well. Shake gently for a few seconds.
- 9. Read plate on a microplate reader at absorbance 450 nm.
  - 1. Visual Determination: Read within 30 min after adding STOP solution.
  - 2. **Spectrophotometric Determination**: Wipe underside of wells with a lint-free tissue. Read absorbance at 450 nm within 30 min after adding STOP solution.

posted January 2008

revised Sepetember 2013